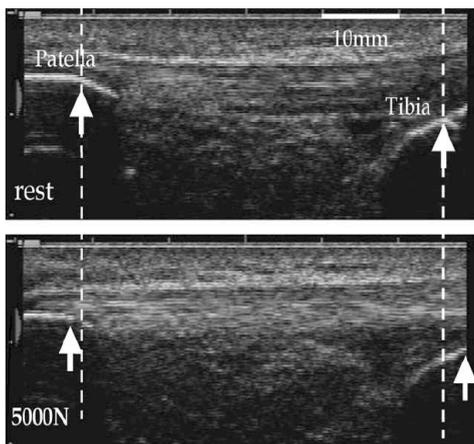
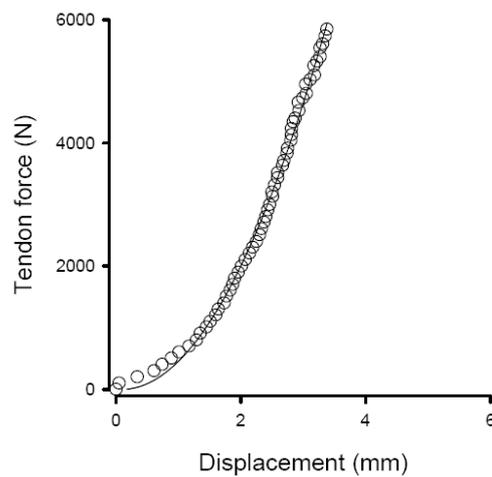
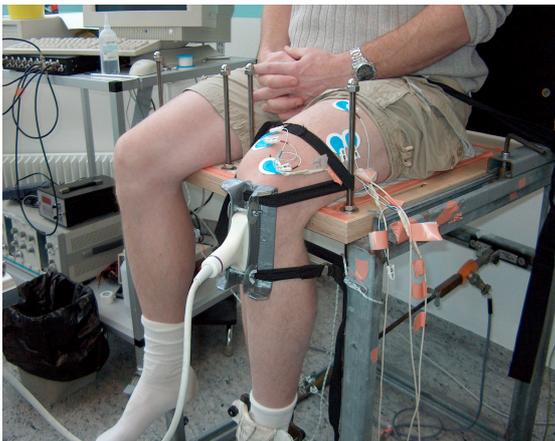


## Ph.D. thesis

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# *Human tendon response to loading, unloading and aging*



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Copenhagen, Denmark, 2010



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*To my son William*

## Abbreviations:

AGE	Advanced glycation endproduct
CSA	Cross-sectional area
HPLC	High pressure/performance liquid chromatography
HP	Hydroxylysylpyridinoline
LP	Lysylpyridinoline
MRI	Magnetic resonance imaging
N	Newton
Nm	Newton meter
US	Ultrasonography
$\varepsilon$	Strain
$\sigma$	Stress
E	Young's modulus of elasticity
Pa	Pascal
PT	Patellar tendon

## List of included papers

This thesis rests on three studies. The studies are listed below referred to in the thesis by their roman numerals.

**I. Habitual loading results in tendon hypertrophy and increased stiffness of the human patellar tendon.** Couppé C, Kongsgaard M, Aagaard P, Hansen P, Bojsen-Møller J, Kjær M, Magnusson SP. *J Appl Physiol.* 2008 Sep;105(3):805-10.

**II. Mechanical properties and collagen cross-linking of the patellar tendon in old and young men.** Couppé C, Hansen P, Kongsgaard M, Kovanen V, Suetta C, Aagaard P, Kjær M, Magnusson SP. *J Appl Physiol.* 2009 Sep;107(3):880-6.

**III. The effects of immobilization on the mechanical properties of the patellar tendon in old men.** Couppé C, Suetta C, Kongsgaard M, Justesen L, Hansen P, Hviid L, Bojsen-Møller J, Aagaard P, Kjær M, Magnusson SP. (Unpublished).

## **Introduction**

Human movement is produced by the force of contracting muscles, which is transmitted to bone through aponeurosis and tendon. It is well known that mechanical properties of bone and muscle are influenced by loading and aging. Repetitive loading (exercise) improves physical function and prevents development of disease. Unloading has the opposite effect. Aging reduces the physical function and increases the rate of disease. Although tendon properties influence the function of the muscle–tendon complex, there is little human data that describe activity and age-associated changes in mechanical properties of tendon. In addition, it is largely unknown if changes in mechanical properties of tendon contribute to the decreased physical function. Therefore, it is important to understand tendon response to physical activity.

## **Tendon Structure and Composition**

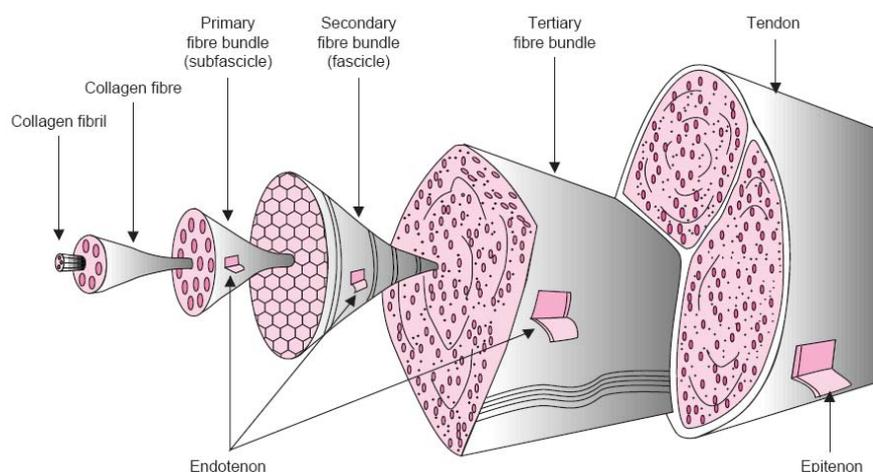
The connective tissue leaves the muscle to be converted into the tendon that connects muscle to bone. Tendon is white, glossy and smooth upon assessment. Tendons have various sizes and forms. Some are long and flat. Others are round and thick (4). Some tendons are enclosed in connective tissue sheath to facilitate sliding (such as the digital flexor tendons) whereas others slide relative to adjacent tissues and skin (such as Achilles tendon). Some tendons wrap around connective tissue "pulleys" that function to increase functional excursion (such as the dorsiflexor retinaculum of the ankle), whereas others pull in a direct line from muscle to bone (such as the patellar tendon or triceps brachii tendon).

Tendons are made up by fibroblasts and an extracellular tendon matrix composed of a mix of collagen and elastin fibers along with ground substance and inorganic components (5). Tendon contain 55-77% water, which to a large extent is associated with proteoglycans of the ECM (Extracellular Matrix) (6). Tendons are composed primarily of the structural protein, collagen (~90% of the total protein in tendon) or 65-75% of the dry mass of tendons (6). Collagen is the most abundant protein in animal world and constitutes more than 30% of the total protein in human body (7;8). The smallest units of collagen is made of polypeptide chains that composed of three amino acids arranged in series: glycine (~30%), proline (~10%) and hydroxyproline (~10%) (9). These chains, due to the three-dimensional nature of the repeating amino acid structure, polymerize in groups of three to create the collagen molecule.

The bonds between the collagen molecules are called cross-links. Cross-linking within and between collagen molecules provides structural integrity to the tendon fibrils in order to secure appropriate force-transduction (10-13). The tendon strength is obtained from cross-links that are assumed to be strengthened through interaction between the collagen and proteoglycan and ground substance. Cross-linking involves two different mechanisms, one precisely controlled enzymatic modifications by the enzyme lysyl oxidase (11;13;14) during maturation and one non-enzymatic mechanism that seems to increase with age and involves reactions with glucose and is generally referred to glycation (1;12;15). Please see a further description in “Age and tendon properties”.

To accurately specify the type of collagen, the structure of the three component chains must be identified. There are numerous chains and thus, collagen itself may exist in one of many isoforms or types. In other words, collagen is classified according to its molecular organization as type I, type II, type III etc. (16). At least 27 types of collagen have been reported (7). Type I collagen is found in skin, bone, tendon, ligament and cornea and is the most abundant type of collagen in the body. Adult tendons consist predominately of collagen type I and type III (6). Collagen type I accounts for about 86% of the dry weight in tendon. Type II collagen is primarily found in cartilage.

Figure 1 shows the generic structure of a tendon (17-20). Collagen molecules are generally aligned in parallel rows to form a collagen fibril. The structural hierarchy of collagen tissue is based on bundling of collagen fibrils. Fibrils are gathered into fascicles that are assembled into the whole tendons. Other extracellular matrix proteins are present in tendons that give it some its viscoelastic properties. These materials include highly charged glycosaminoglycans and proteoglycans that attract water and elastin that presumably provides some of the elasticity of normal tendons (16;21).



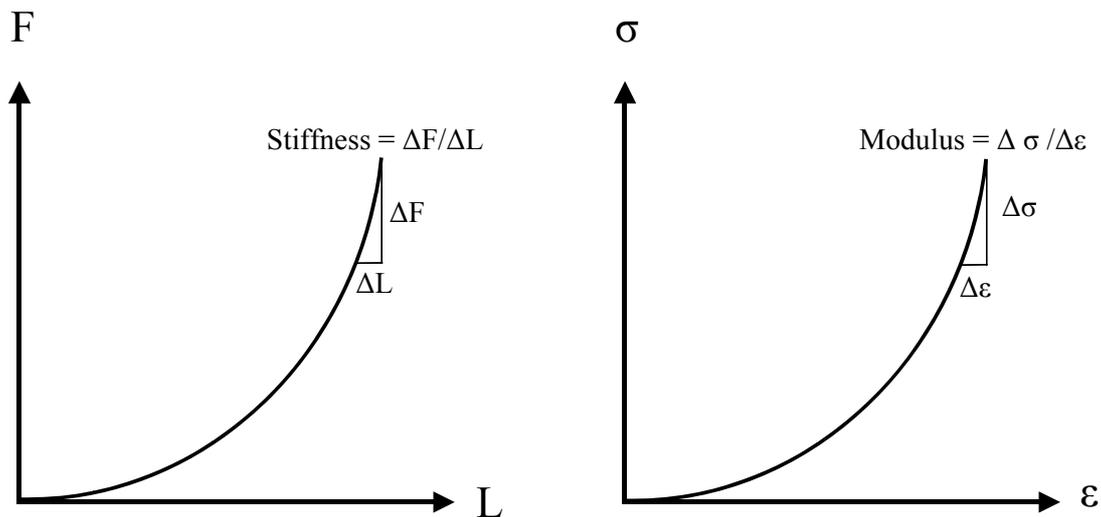
**Figure 1.** Tendon hierarchy. *From K. Khan (3).*

## Tendon biomechanics

The tensile properties of tendons and tendon substructures can be determined by tensile testing that simultaneously record changes in force and elongation of the tendon. Traditionally, whole tendon testing has been performed *in vitro* (1), however ultrasonographic and MRI methods allow for measurement of mechanical properties, *in vivo* (6). For all measurement modalities some fundamental conventions and parameters are used to describe the mechanical properties.

When tensile force is imposed on a tendon it will elongate. The ratio of force to elongation ( $\Delta F/\Delta L$ ) defines the tendon *stiffness*. The unit of stiffness is N/m. The shape of the force-elongation curve is curvilinear with an initial nonlinear “toe-region” and a subsequent “linear region” (figure 2). Straightening of crimps corresponds with the toe-region phenomenon (3; 4). Crimping may provide a mechanical ‘buffer’ against sudden elongation to prevent fibrous damage through shock absorption (7; 8). The linear region likely derives from complex microstructural mechanisms such as elongation of the collagen molecule, increase in the gap region between collagen molecules in the longitudinal direction and relative slippage of laterally neighbouring molecules along the fibril axis (10). Also, non-uniform reorganization and straightening of fibrils may contribute (9).

The structure of a tendon significantly influences the mechanical properties. If the tissue increases cross-sectional area (CSA), the stiffness of the tendon will increase, but reduce elongation for a given force level. Increased tendon length decreases stiffness and increases elongation, for a given force level (Figure 2) (1; 5). To compare material properties for tendons of different dimensions, a common approach is to normalise  $\Delta L$  to the initial tendon length ( $L_0$ ), which yields the unit-less parameter *strain*. Strain is most commonly reported as a percentage. Likewise, a change in force ( $\Delta F$ ) is normalised to tendon CSA to provide tendon *stress*. The unit for stress is Pa. The ratio  $\Delta \text{stress}/\Delta \text{strain}$  yields the *Young's modulus* of elasticity (unit: Pa). Stress, strain, and Young's modulus are utilised to characterise the material properties e.g. of tendon irrespective of material dimensions.



**Figure 2.** Typical force-elongation curve of a tendon on the left.  $F$  = Force (N) and  $L$  = elongation (m). Normalisation of  $\Delta F$  and  $\Delta L$  by tendon dimensions yields a corresponding stress-strain relation on the right.  $\sigma$  = Stress ( $\text{N}/\text{m}^2$ ) and  $\epsilon$  = Strain (%).

Tendons have viscoelastic properties i.e. they exhibit a load-dependent (*time-independent*) linear elastic component and a viscous rate-dependent (*time-dependent*) component (1). Simple spring, dashpot, and frictional elements are required to model the viscoelastic behaviour of tendon (1).

### **Tendon Function**

The primary role of tendon is to transmit force with minimal energy loss from the contracting muscle to the bone in order to produce movement. Tendons do have other functions than acting as “force transmitters”. Tendons also work as “shock absorbers” that can store kinetic energy to be released again (22) and thereby provide considerable elastic energy to locomotion. Animal studies have reported that the tendon is capable of returning over 90% of the stored energy, but some of the stored energy is not recovered due to energy dissipation (23). Further, it appears that the strain energy recovery is larger during running than normal walking demonstrating the importance of elastic saving of energy for economy for motion (24;25). In addition, long and thin tendons (e.g. Achilles tendon) favors energy storage compared to thick and short tendon (e.g. Patellar tendon)(26;27). In sport, these features are critical, as they enhance athletic function. Further, tendons play an essential role in humans provided by evolutionary advantage for bipedal gaits such as walking and running (28). In contrast to apes, human legs have many long spring-like tendons connected to short muscle fascicle that can generate force economically (29). It was recently shown that during stance phase in human walking, the gastrocnemius muscle operates near isometric conditions while it stretches (30). These and related findings during countermovement exercise involving human jumping, support the concept that the human, *in vivo*, gastrocnemius tendon

(31;32) can store and release elastic energy, and thereby confirms previous animal studies. It has been estimated that human tendon provide 52-60% of the total work during locomotion (33). Direct measures, in human, *in vivo*, gastrocnemius tendinous tissue show that it may release ~1.3 J during walking, which corresponds to 6% of the respective total external work (34), ~4 J during two leg vertical jump (32), which corresponds to 16% of the respective total external work (35) with the elastic strain energy increasing with increasing dropping height (36). However, it should be noted that thick tendon would reduce the stress across the tendon, which will raise the safety factor. Moreover, tendon also provide protection from muscle fibre tensile injury (37-39).

### **Tendon strength**

The strength of the tendons is determined by its CSA and by the material properties. The material properties are determined by many factors such the intrafibrillar cross-links and collagen type distribution (40). In this respect the fibrillar organization is also of great importance (41).

Tendons can withstand very high forces. The failure stress of tendons is generally considered to be close 100 MPa (40;42), which corresponds to 10.000 N per cm<sup>2</sup> or ~1000 Kg (43). Therefore most adult PT with a CSA around 1 cm<sup>2</sup> can withstand loads of 10.000 N, but a case report revealed that the PT of a weightlifter was able to withstand 14.500 N (or more than 17.5 times the body weight of the weightlifter) before rupturing (44).

### **Tendon Loading**

The ultimate loads in tendons can be extremely large, especially in tendons such as Achilles and PT. Most studies have determined tendon loading by estimation (45-48), since it difficult to determine the tensile forces of the tendons *in vivo* (49). In the Achilles tendon peak forces can reach ~11000 N/cm<sup>2</sup> in activities such as running and jumping. The loading forces of the PT have not been investigated as much as the Achilles tendon. However, it has been estimated that forces within PT may reach 5000 N during walking, 8000 N during landing from a jump and 9000 N during fast running (47). Ishikawa et al. (50) investigated the PT forces with the fiber optique technique (49) during various types of jumps (squat jumps, countermovement jumps, submaximal drop jumps) on a sledge apparatus. They found that drop jump had the highest values ~7 kN, whereas in squat jump and counter movement jump the PT forces was within 3-6.5 kN. Others have calculated the knee moment of ~500 Nm, (Newtonmeter) which corresponds to a PT force of ~11100 N, (when the

momentarm is 4.5 cm (51)) at landing after a volleyball jump (45;46). These reported tendon forces reach or even exceed critical values if the average ultimate tensile stress is 100 MPa (52). Therefore the tendon CSA becomes paramount to reduce stress and increase the safety factor in many sport activities. Furthermore, the ultimate stress may also be questioned, since they were performed under *in vitro* conditions. However it remains unknown if the variation in tendon CSA is a contributing factor to the development of tendon injuries. Controlled lunged exercises have revealed knee extension moment of 116 Nm ~2500 N within PT (53). Lunge performance has been investigated in fencing in terms of kinematics and EMG activity associated with the movement (54-56). It was found that the most experienced fencers achieved the greatest lunge velocities suggesting that the forces in PT are considerable higher than in controlled exercises.

### **Physical activity and tendon properties**

It is well known that tissues like muscle (57;58) and bone (59;60) adapt to increased loading by hypertrophy and also by improving the material properties. The changed muscle properties will increase the forces from muscle through the tendons. Consequently, this could lead to a situation where the forces on the tendons exceed the strength of the tissue with the risk of sustaining an injury. The knowledge, whether and how tendons adapt to increased loading, has until recently been quite limited. Although animal and human data indicates that muscle CSA is related to tendon CSA (61;62), and that tendons are stiffer in subjects with greater muscle strength (63). However, anthropological evidence suggests that collagen tissue adapts to increased loading with hypertrophy in order to lower stress and thereby avoid injury (28). During evolution, the adoption of bipedality resulted in fundamental anatomical restructuring of bones such as larger CSA of the calcaneal tuber relative to body mass (64). Many studies have found that compared to the early hominids, humans have substantially larger articular surface areas relative to body mass in most joint in the lower extremity (28).

Recent data in human models suggest that tendon tissue is quite metabolically responsive to tensile loading (65-67). In fact, it has been shown that both a single loading bout as well as long term habitual loading produces a markedly elevated collagen synthesis response (68-70). However, to what extent this elevated synthesis yields incorporation of collagen into the load bearing structure of tendon, and therefore either an increase in tendon size (hypertrophy) or an altered composition and a change in mechanical function remains ambiguous.

Studies that address the influence of exercise on the mechanical properties of tendon have until recently largely been conducted in animal models. These data show that endurance like exercise is associated with an increase (71-73), decrease (72), or unchanged (27;74) tendon size, and thus do not provide a coherent picture. It appears that tendon of immature animals are more likely to hypertrophy than tendons of mature animals (61;75). It has been suggested that endurance training results in qualitative but not quantitative changes (74;76), whereas other researches have concluded that tendon reacts to endurance training by both quantitative (hypertrophy) and qualitative ( stress-strain properties) adaptation(77). In humans, cross-sectional data suggests that endurance training is associated with a larger Achilles tendon CSA (62;78;79), which appears to be site specific (62;79). However, in a recent intervention study it was shown that 9 months of endurance training in untrained persons left the Achilles tendon CSA unchanged (80). On the other hand, animal data have shown that muscle strength/size is related to the tendon size (61), suggesting that perhaps the magnitude of loading (81) influences tendon size. Several human studies have shown that resistance training over 12-14 weeks that produces increases in muscle strength of up to 21 % does not result in an accompanying increase in tendon CSA (82-86), but rather a markedly altered modulus, which implies that there is a change in the composition of the structure rather than the size. However, it was recently shown in humans that resistance training for 12 weeks yielded regions specific increases in PT CSA, without a change in modulus (87). Region specific hypertrophy ~5% and increased stiffness in Achilles tendon after 14 weeks of high strain loads (90%MVC), has also recently been confirmed. In this study no change was observed in paired tendon that trained with low strain magnitude (55% MVC) (88). Seynnes et al.(89) recently observed a 5-6% hypertrophy response along the PT together with increased tendon stiffness and modulus followed by 9 weeks of knee extension resistance training.

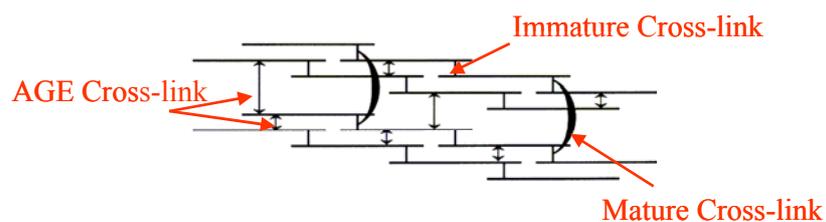
Conclusions from studies of cross-sectional design are inevitably hampered by issues of training history, selection bias and inter-subject variations. Additionally, longitudinal training studies may have been of insufficient duration to produce a robust tendon hypertrophy response. Finally, existing training studies (82-86) have examined tendon size in a region that appears unresponsive to training associated adaptation. However, some of these limitations may be partially circumscribed by examination of region specific PT properties in persons who engage in sport where one lower extremity is habitually subjected to more loading than the contralateral ('control') side, such as in fencing or badminton.

Reference	Design	Subject	Age (yrs)	Training mode	duration	CSA (%)	Stiffness (%)	Modulus (%)
Reeves 2003 (82)	Prospective RCT	18 women & men Healthy	74	Knee ext. + leg press 70% 1RM	14 weeks (3/week)	ns **	65	69
Kubo 2006 (84)	Prospective	14 men Healthy	20 24	Isometric leg press 70% 1RM	12 weeks (4/week)	ns	ns	ns
Kongsgaard 2007 (87)	Prospective	12 men Healthy	25	Knee ext. 70% 1RM vs. 15% 1RM	12 weeks (3/week)	4-6	13	ns
Couppé 2008 (90)	Cross-sectional	7 elite athletes men	23	Habitual sports specific loading	>5 years	20-28	36	ns
Seynnes 2009 (89)	Prospective	15 men Healthy	20	Knee ext. 80% 1RM	9 weeks (3/week)	5-6	20	24

**Table 1.** Patellar tendon and physical activity, human studies, *in vivo*. \*\*CSA analysed with ultrasonography (US)

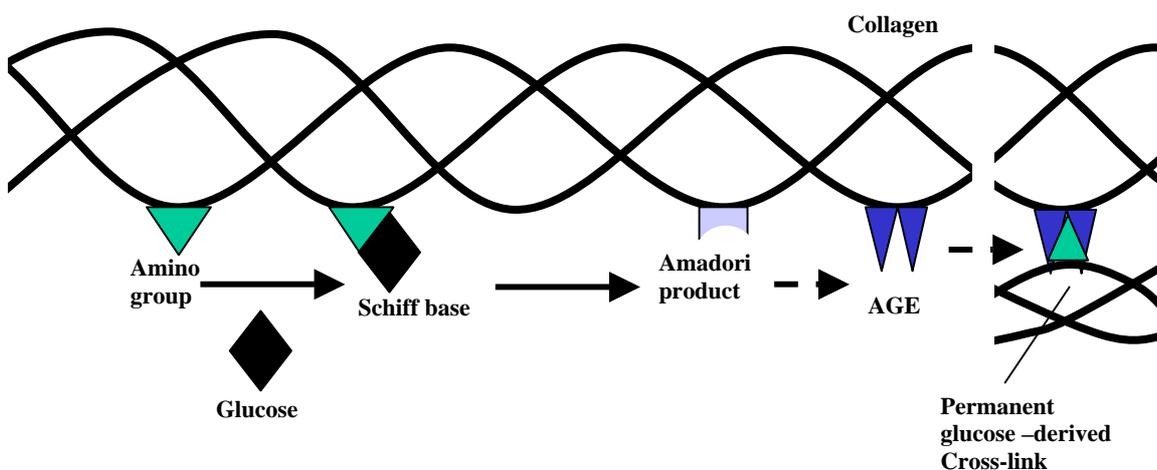
### Aging and tendon properties

Aging is associated with a decline in muscle mass, strength and physical function (91), and although tendon properties influence the overall function of the muscle–tendon complex (92;93), there is a relative lack of human data that describe possible age-associated changes in mechanical properties of tendon. Animal data show that aging yields a stronger and stiffer tendon (23;94-97), a weaker and more compliant tendon (98-100), or leaves the tendon unchanged (101) and are thus inconclusive. In contrast, data on isolated human cadaver tendon suggest that the aging process largely leaves the mechanical properties unaltered (102-104). Despite the development of the US-based methods to evaluate human tendon properties, *in vivo*, (105-107) the effect of aging on the mechanical properties of human tendon *in vivo* remains elusive. Tendon strain in humans has been shown to decrease (108-110) or increase (111-115) with aging. Yet others have shown that aging leaves the mechanical properties of the PT unchanged (116). Thus, the picture is incoherent, which may partly be related to methodological and design differences, the physical activity level of the sample population, and the type of tendon tested, i.e. the tendon-aponeurosis complex or free tendon alone.



**Figure 3.** Location of cross-links. Immature, mature and AGE (Advanced Glycation Endproduct) cross-links during aging of the collagen fibril. Adapted from Bailey *et al.*, 1998 (1).

In tendon, the trivalent intermolecular pyridinoline cross-links (primarily hydroxylysyl-pyridinoline and lysyl-pyridinoline) stabilize the fibrillar structure of collagen and thus contribute to the mechanical properties of the tendon (10;12;117). These cross-links are formed from enzymatically derived covalent immature cross-links (Figure 3), which undergo a spontaneous conversion into more mature trivalent cross-links with collagen maturation (Figure 3) (10). The slow turnover of mature collagen allows further cross-linking via the adventitious non-enzymic reactions of glucose with the lysyl and arginine amino acid residues in the collagen triple helix as a true aging process (10;15;118)(Figure 4). This non-enzymatic process results in the accumulation of advanced glycation end products (AGE) in tendon tissue. The most widely studied AGE is pentosidine (119;120). AGE accumulation is known to accelerate with aging and diabetes (15;121;122) and is believed to yield a stiffer and more load-resistant tendon (123;124). It has been shown that AGE cross-link density in collagenous tissues (125-128) and in human tendons, (129;130) is markedly higher in older compared to younger individuals, and there is only sparse human data on mature cross-link density of tendon and how these are influenced by aging (130). However, recent reports of an age-associated reduction in human tendon stiffness, *in vivo* (111-115), in conjunction with the notion of simultaneously elevated cross-link density and AGE accumulation is difficult to reconcile. To the best of our knowledge there are currently no human studies examining collagen cross-linking and mechanical properties of the PT, *in vivo*, in old and young men.



**Figure 4.** Formation of glucose-derived cross-links, shown highly schematically, begins when glucose attaches to an amino group ( $\text{NH}^2$ ) of a protein such as collagen. The initial product, known as a Schiff base, soon transforms itself into a Amadori product, which can eventually pass through several incompletely understood steps (*broken arrow*) to become an AGE. In many instances AGE's are like unsprung traps, poised to snap shut on free amino groups of any nearby protein and to form cross-links. *From Cerami et al.1987(2).*

## Immobilization and tendon properties

It is well known that immobilization results in a rapid loss of muscle volume and strength, which will negatively influence muscle function (93;131;132). The effects of immobilization on the mechanical properties of the tendon have been studied extensively in animal models (133;134), and several studies show a decline of the mechanical properties of the tendon (72;135-140), while some demonstrate the opposite (141;142). Human studies also suggest that the tendon undergoes a substantial reduction in the mechanical properties with immobilization (143-149), which has been principally attributed to changes in material rather than size (143-149). However, the aforementioned effects of immobilization on the mechanical properties of tendon have predominantly been investigated in young persons (Table 3), despite the fact that old persons are frequently exposed to periods of disuse caused by disease or injury (150;151). It remains unknown if similar magnitudes of changes occur in older persons. However, evidence suggest that quadriceps muscle function is more affected in old than young persons by unloading (132;152), indicating that the magnitude of change in tendon properties is perhaps even greater in the elderly.

Reference	Design	Subject (age)	N	Tendon	Duration (Days)	CSA (%)	Stiffness (%)	Modulus (%)	MVC (%)	RFD (%)	EMD (%)
<i>Kubo (146)</i> 2000	Prospective	Healthy (24)	6	*VL	20		-33		-20	-47	21
<i>Kubo (153)</i> 2004	Prospective	Healthy (24)	6	*VL *MG	20	NS NS	-28 -14, ns		-23		
<i>Kubo (147)</i> 2004	Prospective **	Healthy (24)	6 6	*VL	20	NS	-29 ns				22
<i>Reeves (143)</i> 2005	Prospective **	Healthy (33)	9 9	*MG	90	NS	-58 -37	-57 -38			
<i>Maganaris (154)</i> 2006	Cross-sectional	Spinal-cord injured (36)	6	Patellar	(1.5-24) yrs	↓17	-77	-59			
<i>De Boer (145)</i> 2007	Prospective	Healthy (19)	9	Patellar	14/23	NS	-10/-30	-10/-30	-15 /-21	-42/	/16
<i>Zhao (148)</i> 2008	Cross-sectional	Hemiparetic +women (60)	10	*MG	( >1 yr)	NS	-43	-38			
<i>Shin (144)</i> 2008	Prospective	Healthy +1women (24)	5	Achilles Tendon	44	NS		-17	-54		
<i>Seynnes (149)</i> 2008	Prospective	Healthy (19)	8	*MG	23		-36		-10		

**Table 2** Human studies on tendon and immobilization. All subjects non-active men unless noted, \*\* 1 training group, \* Tendon-aponeurosis, Medial Gastrocnemius = MG, Vastus Lateralis = VL

## **Hypothesis**

*Hypothesis for study 1:* A side specific increased loading will yield a increased tendon hypertrophy and stiffness, but will leave the material properties unchanged.

*Hypothesis for study 2:* Tendon of older persons is more compliant compared to young persons.

*Hypothesis for study 3:* Immobilization will produce a marked decline in muscle function and tendon properties in older persons.

## **Objectives/purpose**

The overall purpose of this thesis was to study the effects of habitual loading, aging and immobilization on the human PT structural and mechanical properties, *in vivo*.

### *Study 1*

The specific purpose of this study was to examine patellar tendon (PT) size and mechanical properties in subjects (elite badminton players and fencers) with a side-to-side strength difference of  $\geq 15\%$  due to sport induced loading.

### *Study 2*

The purpose of this study was to examine the mechanical properties of the patellar tendon (PT), *in vivo* and collagen cross-link composition in old (OM) and young men (YM) subjects.

### *Study 3*

The purpose of this study was to examine the effects of short-term unilateral immobilization on the human patellar tendon (PT) structural and mechanical properties in old men (OM), *in vivo*.

## Material and Methods

In the following section the material and methods are presented along with methodological consideration. Detailed information about material and methods can be obtained in each paper enclosed in the thesis.

### Material

Rationale for design and material in:

#### *Study I (Cross-sectional study):*

We included 7 elite fencers (n = 4) and badminton (n = 3) players that met the criteria of a side-to-side isometric knee extensor strength difference of  $\geq 15\%$ . They had participated in their respective sport for more than five years at an elite level.

The main reason for choosing this design was because that there are several shortcomings in the human studies. The majority of studies are cross-sectional in design, and because there is considerable inter-subject variation (155), it may be difficult to detect difference in limited samples. Also, the longitudinal training studies are typically of relatively short duration, which may simply be insufficient for tissue hypertrophy to take place. One way to particularly avoid these issues was to examine PT properties in subjects who have trained one leg unilaterally for a long time, since the 'untrained' leg then could serve as a within subject 'control'. Both badminton and fencing involves repeatedly performing rapid forward lunges with a preferred and thus more loaded lead extremity compared to the non-lead extremity, which served as a within-subject control. Sapega (156) and Nystrom (157) found more than a 14% strength difference between the legs in elite fencers reflecting a sport specific loading (Figure 5). The sport specific loading in badminton and fencing creates a higher impact on the leg and therefore also on the PT compared with other sports without jumping and unilateral rapid lunges (158). In addition, seven healthy recreational athletes, who did not engage in fencing or badminton were examined with respect to thigh strength and tendon morphology, but not mechanical properties, was also included to ensure that there was no side to side difference between the lead extremity and non-lead extremity side as determined by preferred kicking leg. All players underwent clinical and US assessments on both the lead and non-lead extremity in order to exclude players with knee pathology or patellar tendinopathy (159;160).



**Figure 5.** Typical forward lunge in badminton and fencing, where one side (lead-extremity) is used repetitively compared to the other (non-lead extremity).

### *Study II (Cross-sectional study):*

In study II, 7 old (OM) and 10 young men (YM) with similar activity level (OM  $5\pm 6$  hrs/weeks, YM  $5\pm 2$  hrs/weeks) were included. In order to ensure health status and to determine levels of physical activity during work and leisure time, participants were asked questions (e.g.: hours of exercise/ per week) based on previous questionnaires (161). It has been shown that physical activity such as strength training and habitual loading pattern may result in tendon hypertrophy (87;88;90), which would influence the mechanical properties of the tendon (82). In contrast to previous *in vivo* studies (108;111-116), we took this aspect into account by comparing age groups with similar activity level.

### *Study III (Interventional immobilization study):*

Eight old men with a moderate physical activity level volunteered to participate in the study. They had one lower limb randomly assigned to an immobilization intervention for 2 weeks. The contra lateral limb served as control. The within –subject (paired) design thereby excluded potentially bias of individual intrinsic and extrinsic factors.

## **Methods**

### *Muscle and tendon dimensions*

In all of the included studies Magnetic Resonance Imaging (MRI) was used for determining tendon CSA and length. MRI is a widespread technique for non-invasive visualization of internal tissue

structures. MRI provides images at a relatively high resolution that can be intuitively interpreted. With respect to tendons, the technique has been commonly applied to determine CSA and length either to measure structural changes or to normalize mechanical measurements to stress and strain. The reason for choosing MRI, rather than to other imaging modalities such as US, was that with the MRI-technique tendon hypertrophy has been observed following resistance training (87). US failed to show differences in CSA along the length of the PT (82;154), perhaps due to inaccuracy (162). Subject were asked to abstain from sport activities for 24 hours before the MRI procedure (163;164), since it has been demonstrated that the tendon increases its volume by fluid accumulation.

The anatomical cross sectional area (CSA) of the quadriceps femoris muscle was measured 20 cm proximal to the tibia plateau (mid-thigh level) by magnetic resonance imaging (MRI) (General Electric, Sigma Horizon LX 1.5 Tesla, T1 weighted SE) using a lower extremity coil. The images were obtained using the following parameters: TR/TE = 500/14 ms, FOV 18, matrix 512x512 and slice thickness = 6 mm (87;90). Subsequently, the lean muscle mass of the quadriceps muscle (subcutaneous and intermuscular non-contractile tissue were excluded in the measurement) was manually outlined using the software program Osiris 4.19 (<http://www.sim.hcuge.ch/osiris/>). The mean value of three measurements of the same image was used for analysis. PT CSA and length were determined with the use of MRI (General Electric, Sigma Horizon LX 1.5 Tesla, T1 weighted SE) (87;90). PT CSA was determined by axial plane MR using the following parameters: TR/TE 400/14 ms, FOV 20, matrix 256x256, slice thickness 5.0 mm and spacing 0 mm. The axial scans were performed perpendicular to the PT. As described in detail elsewhere, the tendon CSA was measured I) just distal to the patellar insertion, II) just proximal to the tibia insertion and III) midway between these two sites (87;90) . The PT length was determined from sagittal plane MRI using the following parameters: TR 500, ET: 3 x (TE: 12.4 ms), FOV 16, matrix 256x192, slice thickness 4.0 mm and no spacing. The PT length was obtained by measuring the distance from the dorsal insertion at the patella apex to the dorsal insertion on the tibia.

The PT CSA and length was manually outlined using the software program Osiris 4.19 (<http://www.sim.hcuge.ch/osiris/>). This software was used due to its ability adjust image intensity levels. The color intensity of each image was adjusted using the National Institute Healthcare color scale mode of the software. Tendon CSA and length was measured using the gray scale image display.

In study I, the tendon CSA was measured I) distal to the patellar insertion, II) proximal to the tibia insertion and III) midway between these two sites in order to examine possible site-specific changes. It has been previously observed by Kongsgaard et al. (87) that mechanical loading result in hypertrophy at the distal and proximal portion of the PT. The increased tendon CSA will reduce tendon stress for given force level, which have implications of PT injury that usually occurs at the proximal and distal region (165;166).

In study II, tendon CSA data was normalized to body weight and raised to the power of  $2/3$  (167) in order to compare tendon dimensions between the subjects of various body size. In study II and III, possible site specific differences in tendon CSA were not reported, but tendon CSA was calculated as an average CSA along the tendon from the 3 levels (proximal, mid and distal CSA).

The ability to reproduce CSA measurements on MRI images were investigated in study I, where the mean value of three measurements of the same image was used for analysis. The reproducibility data showed that the typical error percent of repeated measures of tendon CSA was 2.5 % for repeated measurements at the proximal tendon level, 2.5 % for repeated measurements at the mid-tendon tendon level and 2.0 % for repeated measures at the most distal tendon level. In all the studies the MRI examiner was blinded to the subjects including the lead (study 1) or unloaded extremity (study III) in order to avoid investigator bias.

#### *Maximal voluntary contraction (MVC)*

In study I, II, and III, maximal voluntary strength was determined by an isometric knee extension on both legs. In study I, the subjects performed a 10-min warm-up on a stationary bike (5 min at 150 W and 5 min at 200 W) prior to testing. This was done in order to select players with a side-to-side difference in strength of  $\geq 15\%$  was defined as:  $[\text{lead extremity} - \text{non-lead extremity}] / \text{non-lead extremity} * 100 \geq 15\%$ .

In all studies, the subjects were seated in a custom made rigid chair with both hips and knees flexed to an angle of  $90^\circ$ . The reproducibility data showed that the typical error percent of repeated measures of MVC was 4.5% for repeated measurements. In study II and III the MVC was determined after testing of the mechanical properties of the tendon.

### *Mechanical properties of tendon*

The subjects were seated in a custom made rigid chair with both hips and knees flexed to an angle of 90°. A leg cuff, which was connected to a strain gauge (Bofors KRG-4, Bofors, Sweden) through a rigid steel rod perpendicular to the lower leg, was mounted on the leg just above the medial malleolus. An ultrasound probe (7.5 MHz, linear array B-mode, Sonoline Sienna, Siemens, Erlangen, Germany) was fitted into a custom made rigid cast that was secured to the skin above the PT in the sagittal plane. The ultrasound probe and cast was positioned so that the patella, the PT and the tibia were all visible within the viewing field throughout the ramped contractions (Figure6).

The ultrasound S-VHS video images obtained during the ramp trials were sampled at 50 Hz on a PC using frame-by-frame capturing software (Matrox Marvel G400-TV, Dorval, Canada). Force was sampled on two separate PC's at 50 Hz via a 12-bit A/D converter (dt 2810A, Data Translation, MA, USA). The two computers were inter-connected to permit synchronous sampling of all data using a custom-built trigger device (168). The subjects performed 4-5 slow isometric knee extensions ramps by applying gradually increasing force until maximum over a 10 s. period during which PT displacement and knee extension force were measured simultaneously. Each ramp was separated by a 2 minutes rest. All measurements were performed on one side, randomized to either the right or left knee. During the ramp contractions, force was sampled at 50 Hz and lowpass filtered at a 1.0 Hz cutoff frequency using a 4<sup>th</sup> order zero-lag Butterworth filter.

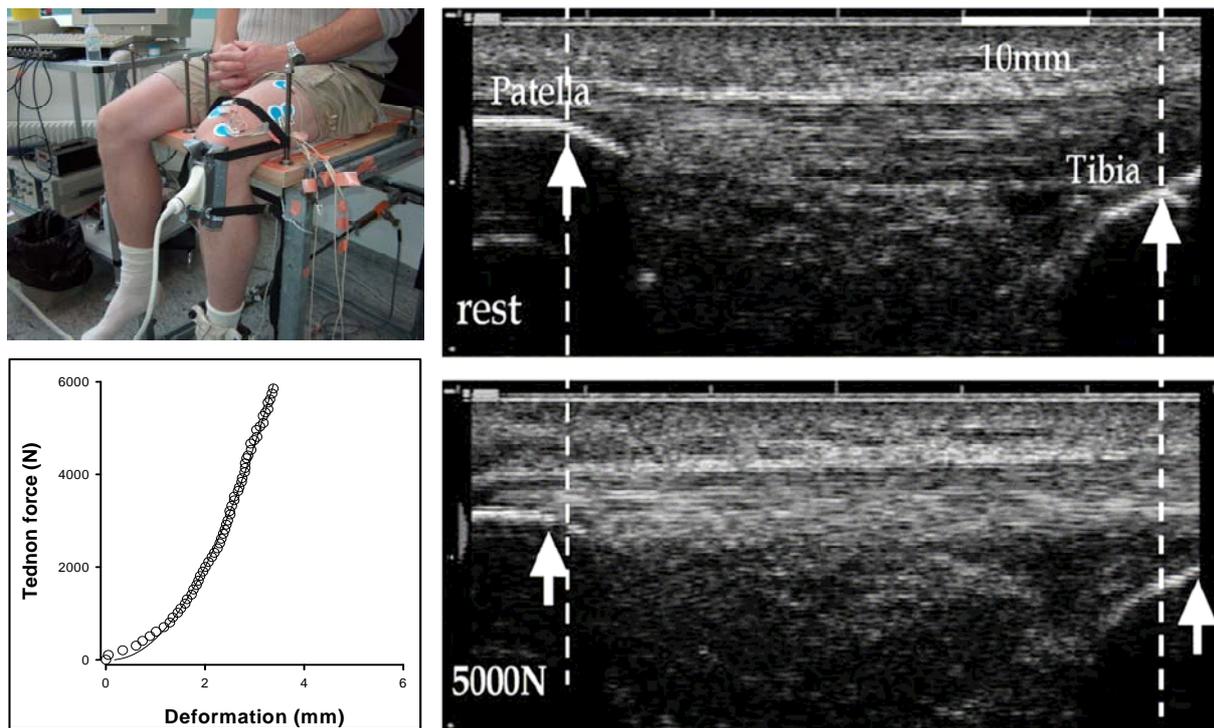
Tendon force was calculated by dividing the estimated total knee extension moment by the internal moment arm, which was estimated from individually measured femur lengths (169). Tendon stress was calculated by dividing tendon force with the average of the 3 levels (proximal, mid and distal) of the PT CSA determined from MRI. Tendon deformation was defined as the change in distance between the patellar apex and the tibia (105;170). Tendon strain was calculated as the change in length related to the initial tendon length ( $\Delta L/L_0$ ). Each single force-deformation curve was fitted to a 2<sup>nd</sup> or 3<sup>rd</sup> order polynomial fit, which yielded  $R^2 > 0.98$ . Tendon stiffness ( $\Delta \text{force} / \Delta \text{deformation}$ ) and Young's modulus ( $\Delta \text{stress} / \Delta \text{strain}$ ) based on common force were calculated in the final 10% or 20% of the force-deformation and stress-strain curves respectively (170).

The use of ultrasonography (B-mode) to determine mechanical properties of the tendon, *in vivo* is widely used. Fukashiro pioneered the use of ultrasonography (B-mode) for measurements of the whole tendon-aponeurosis complex in the mid 90s (106) and later it was developed to measure the isolated tendon (107). The method is easy to apply and its limitations are

widely acknowledged. From the beginning the technique was used to evaluate deformation of the intramuscular fascicular structures.

Several research groups have tried to investigate the free tendon such as the PT using the US (B-mode) methodology by recording the movement of the patella. This is however a proximal displacement of the patella relative to the skin. Structural deformation in the sagittal plane is quantified in sequential US images and then manually analyzed (82;107;145;171;172). It has been shown that the US-based method of obtaining PT deformation requires that the movement of the tibia also has to be accounted for (105;173), which has not been achieved in other studies (82;107;145;171;172). Hansen et al (105) demonstrated that the tibia-movement account for 45% of the total PT deformation under the ramp contractions. Therefore, the measurement of the total PT elongation will be underestimated, if the tibia movement is omitted. For that reason we employed the method of Hansen et al in the 3 studies (I+ II +III). This method has been shown to be accurate and reliable (105). The within-day correlation coefficients and typical error percent of repeated measures were 0.95 and 9.9% for tendon stiffness, 0.97 and 5.5% for tendon strain, and 0.94 and 9.4% for Youngs Modulus.

Importantly, the method can only account for changes in 2 dimensions, when measuring structural deformation. It has been demonstrated that small changes in tendon length during sampling can occur due to the scanning-angle, e.g. medial-lateral movement of the transducer (168). In the three studies this was avoided using an external cast that fixates the transducer to the skin.



**Figure 6.** Measurements of mechanical properties of the patellar tendon. The subjects were seated in a custom made rigid chair with both hips and knees flexed 90°. A leg cuff, which was connected to a strain gauge (Bofors KRG-4, Bofors, Sweden) through a rigid steel rod perpendicular to the lower leg, was mounted on the leg just above the medial malleolus. An ultrasound probe (7.5 MHz, linear array B-mode, Sonoline Sienna, Siemens, Erlangen, Germany) was fitted into a custom made rigid cast that was secured to the skin above the PT in the sagittal plane. The ultrasound probe and cast was positioned so that the patella, the PT and the tibia were all visible within the viewing field throughout the ramped contractions. Synchronized values of PT force and tendon deformation were used to construct force-deformation curves from which the mechanical properties were subsequently calculated based on polynomial fitting.

An automated frame-by-frame tracking software based upon the Lucas Kanade/optical flow estimation method was developed by our lab, that reduces labor time and eliminates the potential examiner bias (105;168;170). Magnusson et al. showed that the software is reproducible and accurate (170). One of the limiting factors includes the sampling frequency of US and the video recording equipment when it comes to analysis of faster contraction velocities. Furthermore, the method lacks an accurate measurement of tendon force.

Tendon is highly viscoelastic tissue. The viscoelastic properties of the tendon will influence the overall function of the muscle–tendon complex (22;25;52;61;93). Therefore, the loading rate will influence the curve or slope due to creep. By definition, a slower muscle contraction produces less stiffness than a fast contraction within the same tendon. This has recently been shown by Pearson (172). In our lab and in the 3 studies (I+II+III), we used 10 s. ramps for 2

two reasons 1) it reduces the rate of force development 2) it ensures more proper tracking of tendon deformation. Subjects in all studies were tested at the same time of the day (171).

It is also well accepted that in addition to the magnitude and rate of tensile loading, the history of loading will affect the tendon properties. This history dependency is readily observed during repeated tensile loading of isolated tendon as the stress-strain curve is shifted to the right, in particular with the initial loading cycles (20;174). Such an acute response of tendon material may have significant physiological implications since it would influence the overall length-tension curve of the muscle-tendon complex. With respect to laboratory measurement procedures, such a rightward shift of the load-deformation curve demands so-called pre-conditioning to obtain reproducible data (20;174). Therefore, before each test of mechanical properties of the PT a preconditioning test were performed following a 5 min. warm-up on a stationary bike.

To avoid an underestimation of real PT force (105;155), the co-activation of the hamstrings have to be taken into account. EMG (Electromyographic) activity from the biceps femoris was therefore obtained during the ramped isometric extension. Due to an unknown noise-source during the first study, which we were not able to filter, we chose to omit the biceps femoris EMG activity in the calculations. From our previous studies we calculated that this error would underestimate the tendon force and tendon stress by 5-20%. However, studies have demonstrated that co-contraction does not change during maximal isometric contractions after 12 weeks of strength training (82;175).

#### *Patellar tendon biopsies*

A Bard MAGNUM® Biopsy Instrument (C.R. Bard, Inc., Covington, USA), with a disposable core biopsy needle (14G) was used. Following sterilization the skin was injected with local anaesthetic (lidocaine, 1%) and a 3-5 mm long incision was created just distal to the patella apex. The biopsy needle was inserted onto the tendon surface at a ~30° angle and fired securing a tissue sample of approximately 8 mg. Samples were snap-frozen in liquid nitrogen and stored at -80°C. Tendon biopsies were taken from the same side as tendon mechanical properties assessments were performed. No previous biopsy had been taken from that site in all subjects. All biopsy samples were analyzed in an investigator-blinded fashion.

### *Biochemical analysis*

Freeze dried tendon samples were hydrolyzed in 6 M HCl, (+108 C, 24 h) and evaporated into dryness and dissolved into H<sub>2</sub>O. Hydroxyproline, the collagen specific amino acid, was measured spectrophotometrically (176) to quantify collagen protein (176). Hydroxylysyl pyridinoline (HP), lysyl pyridinoline (LP) and pentosidine (PENT) were analyzed via a single reversed phase HPLC (high performance liquid chromatography) run and detected on the basis of their natural fluorescence (177). At 0-16 minutes the wavelength for HP and LP fluorescence was 400 nm for emission and 295 nm for excitation. The wavelengths were changed at 16-60 minutes to 328/378 nm to measure the PENT. For the elution of the cross-links, a gradient was built up to contain 17% eluent B (75% acetonitrile with 0.13% HFBA) at 0 min and 25% eluent B at 30 min. Eluent A was 0.13% HFBA. Flow rate was 1ml/min. HP was eluted at 12 min, LP at 13,5 min and PENT at 23 min. The HPLC system used included Quaternary Gradient Pump unit, PU-2089 Plus, Intelligent Autosampler AS-2057 Plus, and Intelligent Fluorescence Detector, FP-2020 by Jasco. Data processing software was Jasco Chrompass. The LiChroCART® 125-4 column was from Merck Hitachi. The results of HP, LP and PENT are given in comparison with the standards injected at four different concentrations in each HPLC run. The intra-assay CVs based on duplicates within a run were 2.6 %, 3.7 %, and 3.9 % for HP, LP, and PENT, respectively. The detection limit for HP and LP is 0.4 pmol and 0.05 pmol for PENT.

## **Statistics**

The statistical analyses of all three studies (I+II+III) were performed with GraphPad Prism® version 4.01. Non-parametric statistics were applied in all instances. An alpha level of P<0.05 was considered significant and all tests were two-tailed.

### *Study I*

Wilcoxon matched-pairs signed-ranks tests were used to examine if there was a side-to-side difference in measured variables. Friedman's analyses of variance with subsequent Dunn's Multiple Comparison tests were used to detect possible tendon region-specific differences along its length. Typical error percent for duplicates measures were used to analyze reproducibility. The typical error percent is calculated as  $(SD_{diff}\sqrt{2} / \text{Mean overall}) \times 100$  and provides a measure of the relative measurement error.

### *Study II*

Mann-Whitney U Tests were used to examine if there were differences between the groups in the measured variables. Spearman's rank-order correlation was used to analyze the strength of relationships between variables.

### *Study III*

Wilcoxon signed rank test were used to examine if there were differences between pre and post immobilization values for the measured variables.

## **Results and Discussion**

In this part, the most important findings from the 3 studies will be presented and discussed. Further detailed information about the study results can be obtained in the articles later in the thesis

### **Study I**

#### *Results*

In the recreational athletes there was no side to side difference in MVC (lead extremity  $217 \pm 14$  Nm, non-lead extremity  $203 \pm 16$  Nm) ( $P > 0.05$ ). In the elite athletes MVC was significantly greater in the lead extremity ( $239 \pm 26$  Nm) compared to the non-lead extremity ( $197 \pm 23$  Nm) ( $P < 0.01$ ).

In the recreational athletes there was no side to side differences in PT CSA for the distal (lead extremity  $124 \pm 7$  mm<sup>2</sup>, non-lead extremity  $120 \pm 10$  mm<sup>2</sup>), mid (lead extremity  $75 \pm 8$  mm<sup>2</sup>, non-lead extremity  $74 \pm 9$  mm<sup>2</sup>) or proximal (lead extremity  $84 \pm 8$  mm<sup>2</sup>, non-lead extremity  $89 \pm 7$  mm<sup>2</sup>) tendon. The CSA of the tendon for the elite athletes are shown in Figure 7. There was a significant side to side difference in CSA at the distal and proximal tendon ( $P < 0.05$ ), but not at the mid-tendon section ( $P = 0.218$ ). The PT CSA was greater at the distal tendon compared to mid and proximal tendon on both the lead extremity and non-lead extremity ( $P < 0.05$ ).

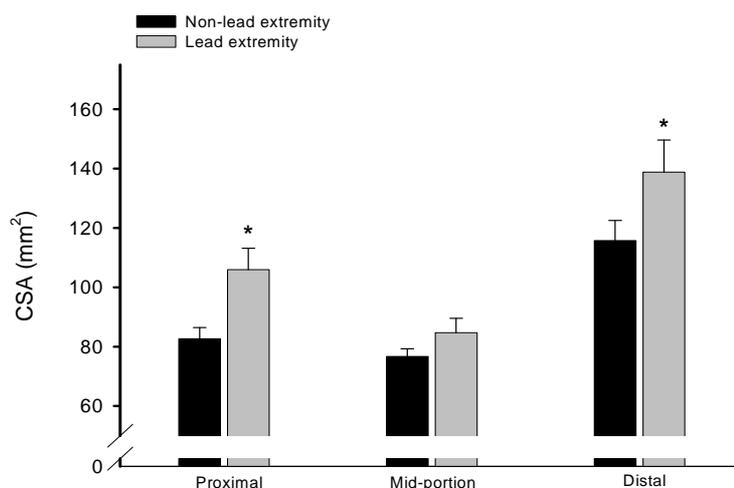
Mechanical properties at a common force are shown in Table 3. Tendon stiffness was higher on the lead extremity compared to the non-lead extremity ( $P < 0.05$ ) (Figure 8). Stress was lower on the lead extremity compared to the non-lead extremity at the proximal (Figure 9) and distal tendon level ( $P < 0.05$ ).

	Lead extremity	Non-lead extremity
Deformation (mm)	2.5 ± 0.3	3.0 ± 0.4
Stiffness (N mm <sup>-1</sup> )	4766 ± 716 <sup>*</sup>	3494 ± 446
Stress (MPa)		
Proximal tendon	52.9 ± 4.8 <sup>**</sup>	66.0 ± 8.0
Mid tendon	65.6 ± 5.6	71.2 ± 8.2
Distal tendon	40.9 ± 4.4 <sup>#</sup>	46.6 ± 4.4 <sup>#</sup>
Strain (%)	5.0 ± 0.5	5.9 ± 0.6
Modulus (GPa)		
Proximal tendon	2.27 ± 0.27	2.16 ± 0.28
Mid tendon	2.87 ± 0.39	2.26 ± 0.25
Distal tendon	1.79 ± 0.25 <sup>#</sup>	1.54 ± 0.19 <sup>#</sup>

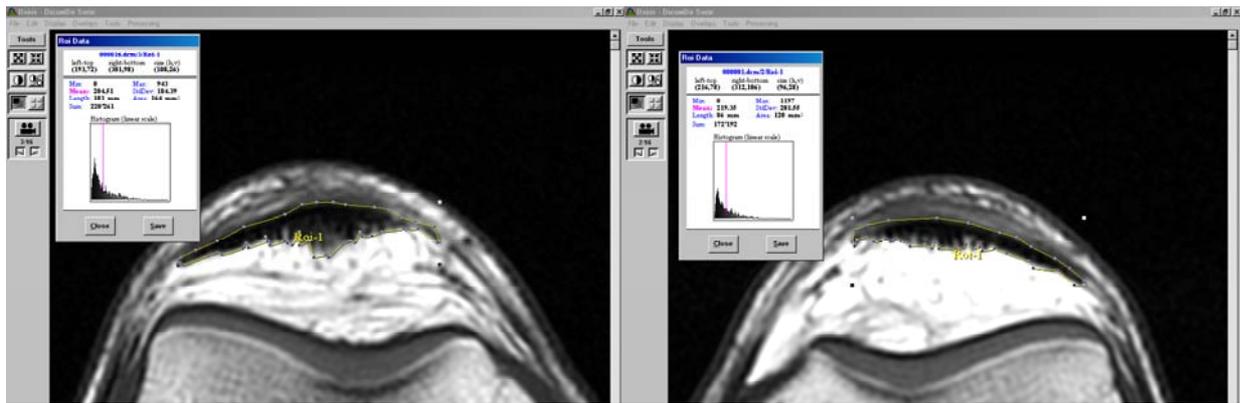
**Table 3.** Patella tendon mechanical properties based on common force. Mean±SEM. Significantly different from non-lead extremity <sup>\*</sup>P<0.05, <sup>\*\*</sup>P<0.01. Significantly different from mid- and proximal tendon <sup>#</sup>P<0.01.

### Discussion

Study 1 examined PT size and mechanical properties in subjects that had a side to side difference in knee extensor strength as a result of habitual sport specific loading. The main findings were that the lead extremity, which was on average 22 % stronger than the non-lead extremity, had a greater distal, proximal and average (24%) PT CSA, which was not evident at the mid-tendon tendon level, indicating a region specific tendon hypertrophy. Further, the lead extremity displayed greater tendon stiffness without a significant difference in modulus, suggesting that the change in mechanical properties was largely the result of a change in size.

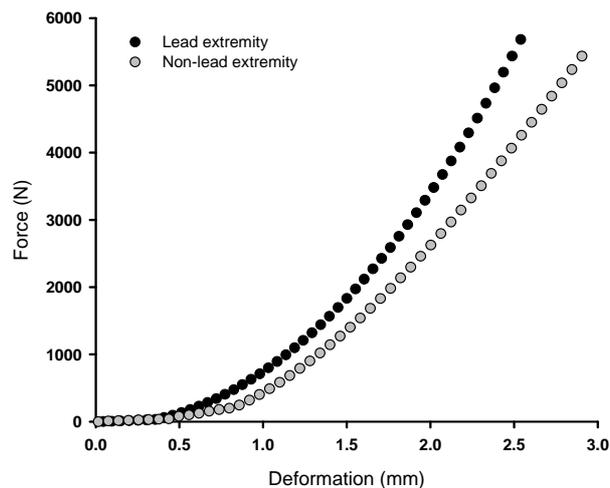


**Figure 7.** Mean ± SEM. The PT CSA distribution. There was a significant side to side difference in CSA at the distal and proximal level but not at the mid-tendon section (<sup>\*</sup>P<0.05). CSA was greater at the distal level compared to mid- and proximal tendon on both the lead extremity and non-lead extremity (<sup>\*</sup>P<0.05).



**Figure 8.** Patellar tendon CSA (just distal to the patellar insertion) on the lead (left image) and nonlead extremity in one elite badminton player determined by axial plane MRI.

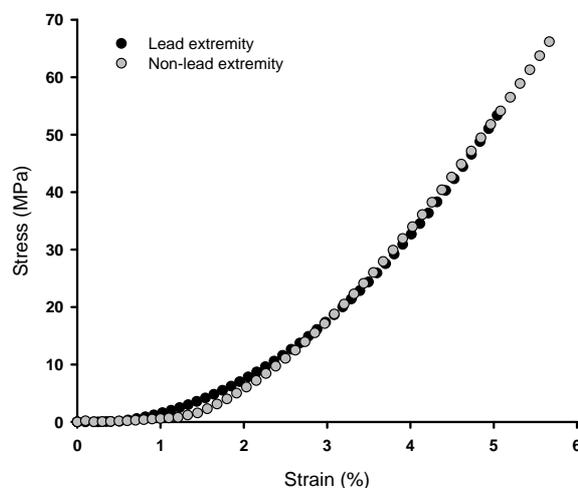
It was demonstrated that the lead extremity had a greater PT CSA in the distal (20 %) proximal region (28%), but not in the mid-section of the tendon. Further, there were no PT CSA side to side differences in recreational athletes. The magnitude of this region specific human tendon hypertrophy confirms and reinforces previous cross-sectional data (62;78;79) and more recent training data (87). Furthermore, the fact that the lead extremity had a 24% greater PT CSA and was 22% stronger than the non-lead extremity, indicates that tendon size follows muscle strength, confirming previous animal data (43;61). However, it was recently shown in humans that 9 weeks of knee extension resistance training yielded a 5-6% region specific hypertrophy response along the PT together with increased tendon stiffness and modulus (89).



**Figure 9.** The PT force-deformation relationship to a common force. Stiffness was higher on the lead extremity compared to the non-lead extremity ( $P < 0.05$ ). Values are means of all subjects.

Like previous studies (87;90), Seynnes et al. have confirmed a substantial increase in tendon CSA at the proximal and distal region, and interestingly also finds a small increase in tendon CSA at the mid-portion of the PT. In the present study, we did not observe an increase at the mid-tendon, maybe due to the sample size was limited. However, Kongsgaard et al.(87) was not able demonstrate an increased tendon CSA at the mid-portion, indicating a limited hypertrophy response in this part of the tendon (178-180). Moreover, region specific hypertrophy (~5%) and increased stiffness in Achilles tendon after 14 weeks of high strain loads (90%MVC), has also recently been confirmed. In this study, no change was observed in paired tendon that trained with low strain magnitude (55% MVC) (88). This indicates that a firm strain or force threshold may exist before tendon adaptations occur such as increase in tendon CSA and tendon stiffness. More, recently, it has been suggested that the applied strain on the connective tissues regulates the catabolic and anabolic responses of the cells (181). Further, an external mechanical loading of the tissue above the upper limit at which the endogenous contraction of the fibroblast may maintain their tensional homeostasis should stimulate cells for remodeling, whereas a reduction of the mechanical threshold below the lower limit will lead to tissue destruction (182).

Altogether, it seem that tendon primarily adapts to increase loading by tendon hypertrophy that leads to greater tendon stiffness. In addition, the increased tendon CSA and tendon stiffness have several clinical implications; it will decrease tendon stress and strain for any given magnitude of tensile loading, and may therefore reduce the risk of overload injuries, which may be protective mechanism. Further, the greater stiffness may also improve the load transfer (e.g. RFD) and performance (e.g. running economy)(183).



**Figure 10.** The PT stress-strain relationship based on common force. Stress was lower on the lead extremity compared to the non-lead extremity at the proximal tendon level ( $P < 0.05$ ). Values are means of all subjects.

## Study II

### Results

Tendon dimensions are shown in Table 4. Tendon length did not differ between YM and OM.

Absolute average tendon CSA did not differ between OM and YM. Similarly, there was no between group difference in average tendon CSA normalized for body weight.

	OM (n = 7)	YM (n = 10)
Tendon length (mm)	43±4	43±5
Tendon CSA (mm <sup>2</sup> )	101±17	103±8
Tendon CSA (mm <sup>2</sup> /BW <sup>2/3</sup> )	5.34±0.48	5.47±0.25
Q-ACSA (mm <sup>2</sup> )	6214±705**	8431±522

**Table 4.** Patellar tendon dimensions and quadriceps muscle cross-sectional area (Q-ACSA) for old (OM) and young men (YM). Values are mean±SD. \*\* Significantly different from YM, P<0.01.

Mechanical properties determined at maximal force are shown in Table 5. Maximal tendon force was lower in OM than YM ( $P<0.05$ ). There were no differences between OM and YM with respect to tendon deformation, stiffness, strain, stress or Young's modulus based on average

Mechanical properties at a common force are shown in Table 6. Again, there were no differences between OM and YM for any of the variables (Figure 11 & 12).

Collagen concentration and cross-link density data are shown in Table 7. Collagen concentration was lower in OM than YM ( $P<0.05$ ). Both HP and LP ( $P<0.05$ ) as well as PENT ( $P<0.01$ ) concentrations in collagen were higher in OM than YM. PENT was positively related to age in YM ( $r = 0.74$ ,  $P<0.01$ , Figure 13), but not in OM ( $r = 0.65$ ,  $P = 0.11$ ). There were no significant correlations between the mechanical and biochemical variables.

	OM (n = 7)	YM (n = 10)
Force (N)	5161±737*	7415±2184
Deformation (mm)	2.6±0.4	2.9±0.9
Stiffness (N/mm)	3926±1091	5546±1871
Stress (MPa)	51±8	65±24
Strain (%)	6.1±0.9	6.9±2.3
Modulus (GPa)	1.7±0.3	2.2±0.7

**Table 5.** Patellar tendon mechanical properties for old (OM) and young men (YM) based maximum force. Values are mean±SD. \* Significantly different from YM, P<0.05.

## Discussion

To the best of our knowledge this is the first study that has examined both the mechanical properties of the human PT, *in vivo*, and collagen cross-links densities in YM and OM persons with a similar physical activity level. The main findings were that the collagen concentration was lower in OM than YM, while the enzymatically derived cross-links (HP and LP) were greater in OM compared to YM. At the same time, the non-enzymatically derived AGE marker (PENT) was markedly more abundant in OM compared to YM. However, despite these apparent age-related differences in the tendon collagen properties, the tendon mechanical properties in the two age separated groups did not diverge appreciably.

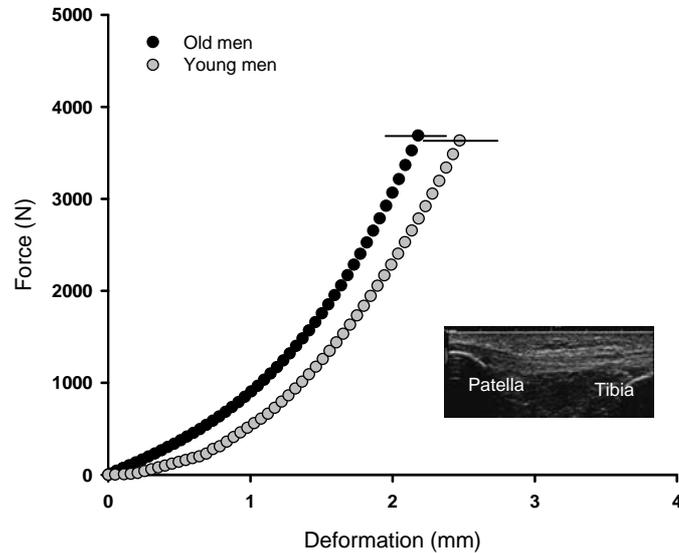
	OM (n = 7)	YM (n = 10)
Deformation (mm)	2.3±0.4	2.4±0.6
Stiffness (N/mm)	3511±837	3290±869
Stress (MPa)	41±7	37±5
Strain (%)	5.3±0.9	5.9±1.7
Modulus (GPa)	1.5±0.4	1.4±0.4

**Table 6.** Patellar tendon mechanical properties for old (OM) and young men (YM) based on common force. Values are mean±SD.

The present mechanical data diverges from that of others based on human *in vivo* model with one exception (116), and this may be related to methodological differences and study design. It has been shown that strength training and habitual loading pattern may result in tendon hypertrophy (87;88;90), which would influence the mechanical properties of the tendon (82). In contrast to previous *in vivo* studies (108;111-116), we have taken this aspect into account by comparing age groups with similar activity level. However, it cannot be ruled out that prior life long training history, including exercise mode and intensity, which was unaccounted for in the study may have influenced the data. It should also be recognized that the present data and that of others are based on cross-sectional designs with inherent limitations, including the striking variations in tendon mechanical properties between subjects (155), and therefore a type II error cannot be ruled out.

	OM (n = 7)	YM (n = 10)
Collagen (mg/mg dry weight)	0.49±0.27*	0.73±0.14
HP (mmol/mol collagen)	898±172*	645±183
LP (mmol/mol collagen)	49±38**	16±8
PENT (mmol/mol collagen)	73±13**	11±2

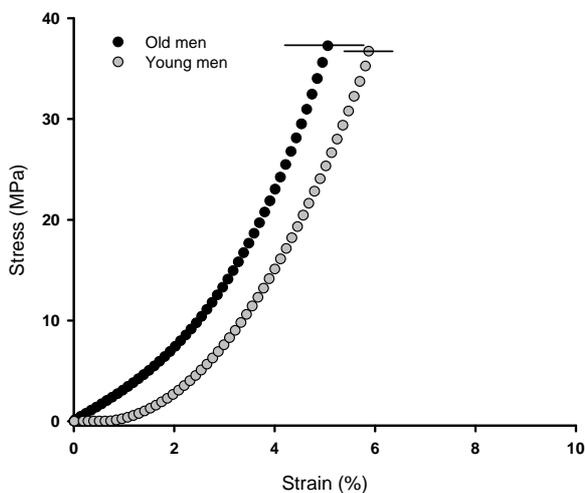
**Table 7.** Concentration of collagen and hydroxylysyl pyridinoline (HP), lysyl pyridinoline (LP) and pentosidine (PENT) cross-links in the patellar tendons of old (OM) and young men (YM). Values are mean±SD. \*, \*\* Significantly different from the values of YM, (P<0.05 and P<0.01, respectively).



**Figure 11.** The patellar tendon force-deformation relationship based on common force. Values are mean $\pm$ SD of all subjects. There were no differences between old (OM) and young men (YM) with respect to tendon deformation or stiffness ( $P>0.05$ ).

The density of mature lysyl oxidase-derived intermolecular covalent cross-links, such as the HP and LP, gradually increase during tendon tissue maturation, and it is commonly believed that these cross-links are the chief contributors to the function and mechanical properties of the tendon (10;12;117;184). In animal models it has been shown that there is a positive relationship between HP cross-link density and mechanical strength of the healing medial collateral ligament tissue and ACL graft (185;186), and furthermore that changes in HP and LP density will result in altered mechanical properties in collagenous tissue (10;187). However, there is only sparse human data on mature cross-link density in tendon and how these are influenced by aging (130). A small but significant age-related increase of LP has been demonstrated in the supraspinatus tendon, *in vitro* (130). The present data on LP and HP density on YM corresponds well with that previously reported in a similar population (159). However, the present data also demonstrate that HP and LP density of the human PT was  $\sim 40\%$  and 3-fold higher, respectively, in OM compared to YM, which suggests a rather marked age associated elevation in these enzymatic cross-links. It is noteworthy that despite the rather robust difference in both HP and LP there was no difference in the mechanical properties of the tendon. However, it should be noted that fibril length (188;189), fibril diameter (190), and proteoglycans and glycosaminoglycans (6;191;192) have all been

implicated in contributing to the tendon mechanical properties, and the relative contribution of these factors and that of mature cross-links remains elusive.



**Figure 12.** The patellar tendon stress-strain relationship based on common force. Values are mean $\pm$ SD of all subjects. There were no differences between old (OM) and young men (YM) with respect to tendon strain, stress or Young's modulus based on average tendon CSA ( $P>0.05$ ).

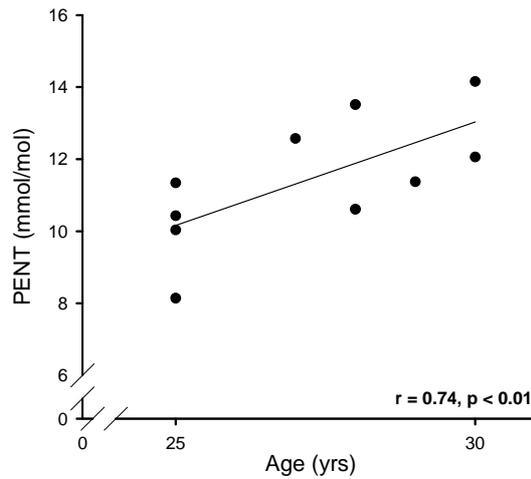
To our knowledge these are the first data showing that AGE is markedly increased in OM compared to YM in the human PT. AGE cross-links, including PENT, are formed when lysine amino acid residues in the collagen triple helix come into contact with glucose (10;15), and the accumulation of these cross-links are known to accelerate with aging (6;10) and disease processes such as diabetes (15;121;122), atherosclerosis (193), Alzheimer (194;195) and renal failure (196). AGE products are also used as markers of tissue turnover (130). It has been shown that the difference between young and older individuals in AGE cross-link density in collagen is approximately 2-fold in human skeletal muscle (127), 5-fold in human bone (128), 7-fold in human tendon (present data), 9-fold in human ligaments (126), and 33-fold in human cadaver cartilage (125), demonstrating the tissue specific turnover. The fact that PENT density is 7-fold greater in tendon of OM compared with YM (Table 6) coupled with the fact that PENT appears to be related to age in a narrow age span (Figure 13) firmly demonstrates the positive relationship between AGE cross-linking of human PT collagen and aging, *ex vivo*. These data corroborate and extend those previously reported on cadaver tissue (129;130). From a functional standpoint, an elevated AGE cross-link density has been suggested to result in increased tensile stress and tendon stiffness in animal models (97;123;124;197-200). However, in the present study both the stiffness (Figure 11) and the Young's modulus (Figure 12) of the tendon did not differ between OM and YM despite the

7-fold difference in PENT, which suggest that factors other than AGE may also play a major role in determining the mechanical properties of human tendon.

In the present study, the total collagen concentration of the PT was ~ 34 % lower in OM compared to YM (Table 6), and this age linked reduction is in accordance with that found in the canine PT (101) and rat tail tendon (99). It is possible that the lower collagen concentration with aging may represent the reduced size and/or density of collagen fibrils that is known to occur with aging (178;184;190;201;202). It was recently reported that MRI signal intensity of the PT was reduced with aging (116), which may be a function of the reduction in collagen concentration observed in the present study. Unfortunately, the size of the obtained biopsy in the present study precluded transmission electron microscopy analysis for fibril size and density. The lower collagen concentration in OM may be an age-related change in the tendon per se, and/or a function of reduced tendon loading due to an age-related loss of muscle mass/strength. Interestingly, the average whole tendon CSA did not differ between OM and YM (Table 3), which in light of the reduced collagen concentration may be related to increased amounts of other extracellular matrix components, such a proteoglycans and glycosaminoglycans. Alternatively, the retained tendon CSA in OM may result from tendon intrafibrillar fat that can accumulate with aging (203-205).

Albeit speculative, it is possible that the elevated enzymatic and/or non-enzymatic cross-link density in OM compared to YM served to maintain tendon stiffness and Young's modulus despite the diminished collagen concentration. Such maintenance of tendon stiffness would serve to maintain effective transfer of muscle force despite a lower absolute muscle size (Table 3) and strength (Table 4). In this context it should be noted that the present data were obtained in moderately physically active person and future studies will need to address the effect of training per se in elderly.

In conclusion, the data of present study raise the possibility that the dimensions and mechanical properties of the human PT, *in vivo*, may not differ between old and young men. On the other hand, old men displayed lower collagen concentration, but greater enzymatic (HP & LP) and non-enzymatic (PENT) collagen cross-links compared to young men. This age related increase both in enzymatic and non-enzymatic cross-linking compounds, may serve to maintain the mechanical properties of tendon with aging.



**Figure 13.** PENT was positively related to age in young men (YM) ( $r = 0.74$ ,  $P < 0.01$ ).

### Study III

#### Results

There were no side-to-side differences for any of the measured variables prior to immobilization. Peak knee extensor moment decreased on the immobilized side ( $143 \pm 25$  vs.  $119 \pm 26$  Nm,  $P < 0.05$ ). A statistical trend was observed toward a decrease in peak knee extensor moment ( $152 \pm 32$  vs.  $139 \pm 39$  Nm,  $P = 0.078$ ) on the control side.

(n = 8)	Immobilized side		Control side	
	Pre	Post	Pre	Post
Tendon length (mm)	43±3	43±3	43±6	43±3
Tendon CSA (mm <sup>2</sup> )	101±11	101±11	99±15	102±16

**Table 8.** Patellar tendon dimensions for the immobilized side and control side. Values are mean ± SD. There were no differences in patellar tendon dimensions between Pre and Post immobilization

Tendon dimensions are shown in Table 8. There were no changes with respect to tendon length or absolute tendon CSA on the immobilized or the control side. Mechanical properties assessed at maximum common force are shown in Table 9. Tendon stiffness decreased both on the immobilized side ( $P < 0.01$ ) (Figure 14) and on the control side ( $P < 0.01$ ) (Figure 15). Likewise, Young's Modulus decreased on the immobilized side ( $P < 0.05$ ) (Figure 16) and on the

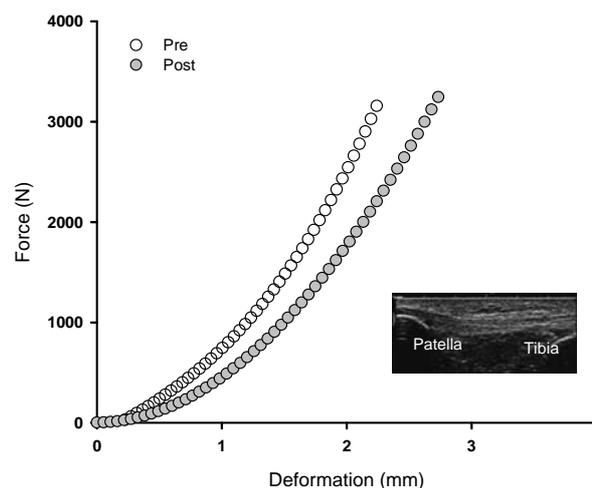
control side ( $P<0.05$ ) (Figure 17). There were no other differences with respect to tendon deformation, strain or stress on either the immobilized side or the control side.

(n = 8)	Immobilized side		Control side	
	Pre	Post	Pre	Post
Deformation (mm)	2.3±0.5	2.7±0.8	2.3±0.4	2.8±0.6
Stiffness (N mm <sup>-1</sup> )	2949±799	2366±774**	3330±839	2365±436**
Stress (MPa)	32±7	33±10	34±8	32±8
Strain (%)	5.5±1.6	6.6±2.2	5.3±0.7	6.5±1.7
Modulus (GPa)	1.2±0.3	1.0±0.3*	1.5±0.4	1.0±0.3*

**Table 9.** Patellar tendon mechanical properties for the immobilized side and control side based on common force. Values are mean ± SD. Significantly different from Pre, \* $P<0.05$ , \*\* $P<0.01$ .

### Discussion

To the best of our knowledge this is the first study that has examined the effects of short-term unilateral immobilization on the structural and mechanical properties of the human PT, *in vivo*, in old healthy individuals. The main findings were that the mechanical properties of the PT decreased on the immobilized side. A decline in stiffness and Young's modulus took place without any measurable change in the size of the tendon, suggesting that the material properties of the tendon were primarily affected by the unloading.



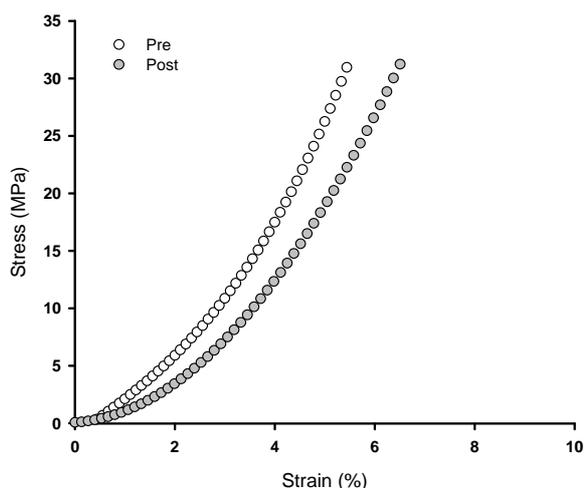
**Figure 14.** The patellar tendon force-deformation relationship based on common force for the immobilized side. Values are mean; SD values are stated in Table 2. Stiffness decreased after 14 days of immobilization (Pre vs. Post,  $P<0.01$ ).

It is well known that the immobilization associated drop in muscle strength is greater than that of muscle mass, and that both contribute to a reduced muscle function (92;93;132). The larger drop in muscle strength has mainly been attributed to rapid changes in the nervous system that occur with short term unloading (92;206-208). Interestingly, recent studies report that unloading affect tendon properties to a greater extent than muscle loss, which in turn may contribute to a decline in electromechanical delay (the time lag between muscle activation and muscle force production) and reduced rate of force development (92;145;146;153;209). However, evidence suggests that quadriceps muscle function is more affected in old than in young persons by unloading (132;152), indicating that the magnitude of change in tendon properties is perhaps even greater in the elderly. Nevertheless, the aforementioned effects of immobilization on the mechanical properties of tendon have mainly been investigated in young persons, despite the fact that old persons are frequently exposed to periods of disuse caused by disease or injury (150;151), and if similar or greater magnitudes of changes occur in older persons remains unknown. However, in the present study, it was demonstrated for the first time *in vivo* for aging human individuals, that 14 days of immobilization led to decreased tendon stiffness on the immobilized side, which was accompanied by a decrease in Young's Modulus. Furthermore, a similar effect was observed on the control side. This decline occurred without changes in tendon dimensions, indicating that the material properties of the patella tendon were altered for our old subjects, which is in line with previous reports in young persons (144-146;149;153). In the present study, we extended these findings by showing that muscle tendon mechanics are also affected by acute disuse in the elderly, and that the magnitude of change is perhaps even greater. Collectively, these data demonstrate that short-term limb unloading has considerable effects on the mechanical properties of the tendon that to some extent mirrors that of the muscle, and that these changes can be expected to occur in the early phase of unloading.

It has been proposed that the degree of tension on the PT during immobilization could influence the tendon biomechanical properties quite dramatically within one week (210). The present study did not investigate possible biochemical changes, and it should be kept in mind that the immobilized knee was positioned in a relatively shorten position ( $\sim 30^\circ$  knee flexion), which likely is associated with no or only very minor passive tension in the patella tendon (170). The acute lack of tensile tendon input during the immobilization period might have influenced the material properties of the tendon, since tendon stiffness and modulus decreased without tendon atrophy. Unloading tenocytes have been shown to have severe effects on collagen expression, proteoglycan

expression, growth factor secretion and metalloproteinases (182;211). Furthermore, it has recently been demonstrated that complete tension deprivation in the rabbit PT induced fibroblast apoptosis within 24 hrs (212). Thus, it is possible that the removed tension development in the PT was the initiating event for the present findings of reduced tendon stiffness and Young's Modulus.

A number of limitations may be observed with the present study. Unfortunately we were unable to obtain tendon biopsies in the present study, which would have allowed us to address possible changes in tendon composition. Furthermore, to our surprise changes occurred in the non-immobilized limb, which rendered it inadequate as a control. Albeit non-significant, muscle strength tended ( $P=0.078$ ) to decrease on the non-immobilized side. Furthermore, both stiffness and modulus also decreased on the non-immobilized side. Thus, although the data show that the non-immobilized side was not an ideal control in this sample, the data suggests that muscle strength changes are accompanied by changes in tendon properties. A possible explanation for the findings in the control limb is that unilateral casting and crutch walking in old persons may influence the



**Figure 15.** The patellar tendon stress-strain relationship based on common force for the immobilized side. Values are mean; SD values are stated in Table 2. Stiffness decreased after 14 days of immobilization (Pre vs. Post,  $P<0.05$ ).

overall activity level. It has been reported that overall amount of weight-bearing support per day is decreased during unilateral limb suspension monitored by accelerometry (213;214).

In conclusion, a decline in stiffness and Young's modulus was observed following short-term (14 days) unilateral limb immobilization in old men without any changes in tendon size, suggesting that the material properties of the tendon were affected by unloading. These findings indicate that short-term inactivity has a major negative influence on the mechanical properties of the human PT in the elderly.

## Conclusions

The main focus in this thesis was to investigate the effects of long-term habitual sport specific loading, aging and short-term immobilization on the human PT structural and mechanical properties, *in vivo*.

It was found that a habitually more loaded PT of a stronger lower extremity had a greater PT CSA compared to a less loaded PT of a weaker lower extremity. However, the greater tendon CSA was evident at the distal and proximal level and not at the mid-tendon, indicating a region specific tendon hypertrophy. Further, the habitually more loaded PT displayed greater tendon stiffness without a significant difference in modulus, suggesting that the change in mechanical properties was largely the result of a change in size. In study II, investigating the age-related changes in tendon structure and function, it was observed that the collagen concentration was lower in OM than YM with a similar physical activity level, while the enzymatically derived cross-links (HP and LP) were greater in OM compared to YM. At the same time, the non-enzymatically derived AGE marker (pentosidine) was markedly more abundant in OM compared to YM. Despite these apparent age-related differences in the tendon collagen properties, the tendon mechanical properties in the two age separated groups did not diverge appreciably. It is possible that the elevated enzymatic and/or non-enzymatic cross-link density in OM compared to YM served to maintain tendon stiffness and Young's modulus despite the diminished collagen concentration. Such maintenance of tendon stiffness would serve to maintain effective transfer of muscle force despite a lower absolute muscle size and strength. In the third study, short-term (14 days) unilateral limb immobilization in old men resulted in a decrease in the mechanical properties of the PT on both sides. A decline in stiffness and Young's modulus without any changes in tendon size was observed, suggesting that the material properties of the tendon were affected by unloading. These findings indicate that short-term inactivity has a major negative influence on the mechanical properties of the human PT in the elderly.

## Perspective

The completed studies answered some questions, but also prompted several additional questions that may be addressed in future studies.

Based on Study I, it may be of interest to better understand the hypertrophy response of chronic habitual loading by investigating the composition of tendon through tendon biopsy (e.g., collagen cross-links, fibrils morphology, and density). Furthermore, it is still largely unknown how the tendon adapts to different types of loading. It could, therefore, be interesting to look at the effect of different loading regimes such as resistance versus endurance training. Additionally, the possible combined effects of these two, and how they influence the mechanical properties and the composition of the tendon is not understood.

From bone and muscle research, it is well established that mechanical loading and physical activity in general improves tissue properties, physical function, and prevents disease. It seems that age of initiation of physical activity affects disease. Therefore, the importance and effect of activity (training) during the growing years and how it influences the mechanical properties of the tendon can be studied in several sport populations. Furthermore, the effect of early physical training on the tendon properties, not only needs to be studied in younger persons but also in the elderly.

In line with the findings in Study II, it may also be of interest to investigate the importance of different levels of physical activity on aging tendon. For instance, it may be of interest to study whether the mechanical properties and the composition of the tendon, *in vivo*, are changed in master athletes (older persons that have trained all their life), and especially the role of AGE cross-links and how they are influenced by physical activity.

From Study III and from a rehabilitation point of view, it would be essential to investigate if the mechanical and the material properties can be restored after a period of immobilization in the elderly. Lastly, it is of interest to study if the effects of immobilization on the mechanical properties of the tendon can be prevented by passive range of motion or by putting the tendon into elongated position during immobilization.

## Abstract

The overall purpose of this thesis was to study the effects of habitual loading, aging and immobilization on the human PT structural and mechanical properties, *in vivo*.

### *Study I*

The specific purpose of this study was to examine PT (PT) size and mechanical properties in subjects (elite badminton players and fencers) with a side-to-side strength difference of  $\geq 15\%$  due to sport induced loading.

The main findings were that the lead extremity, which was on average 22% stronger than the non-lead extremity, had a greater distal and proximal PT CSA, which was not evident at the mid-tendon tendon level, indicating a region specific tendon hypertrophy. Further, the lead extremity displayed greater tendon stiffness without a significant difference in modulus, suggesting that the change in mechanical properties was largely the result of a change in size.

### *Study II*

The purpose of this study was to examine the mechanical properties and pyridinoline and pentosidine cross-link, and collagen concentrations of the PT, *in vivo*, in old (OM) and young men (YM) with a similar physical activity level.

The main findings were that the collagen concentration was lower in OM than YM, while the enzymatically derived cross-links (HP and LP) were greater in OM compared to YM. At the same time, the non-enzymatically derived AGE marker (PENT) was markedly more abundant in OM compared to YM. However, despite these apparent age-related differences in the tendon collagen properties, the tendon mechanical properties in the two age separated groups did not diverge appreciably.

### *Study III*

The purpose of this study was to examine the effects of short-term (14 days) unilateral immobilization on the human PT structural and mechanical properties in old men (OM), *in vivo*.

The main findings were that the mechanical properties of the PT decreased on the immobilized side. Furthermore, a similar effect was observed on the control side. A decline in stiffness and Young's modulus took place without any measurable change in the size of the tendon, suggesting that the material properties of the tendon were primarily affected by the unloading.

## Dansk abstract

Hovedformålet med denne afhandling var at undersøge effekterne af langtidstræning, aldring og immobilisering på den humane patellasenes strukturelle og mekaniske egenskaber, *in vivo*. I afhandlingen indgår tre separate studier.

### *Studie I*

Formålet med dette studie var at undersøge patellasens størrelse og mekaniske egenskaber hos personer (elite badmintonspillere og fægtere) med en side til side styrke  $\geq 15\%$  som følge af langvarig idrætsspecifik belastning.

Hovedfundene i studiet var, at patellaseen på fremfaldsbenet, som gennemsnitligt var 22% stærkere end ikke-fremfaldsbenet, havde et større tværsnitsareal i den distale og proximale del men ikke i midt-sene delen, hvilket tyder på en regionsspecifik hypertrofi. Yderligere havde fremfaldsbenet en stivere patellase, uden at der var forskel i Modulus, hvilket indikerer at ændringer i de mekaniske egenskaber var et resultat af ændret sene størrelse.

### *Studie II*

Formålet med dette studie var at undersøge de mekaniske egenskaber, pyridinolin og pentosidine tværbindinger samt kollagen koncentrationen i patellaseen, *in vivo*, hos unge og ældre mænd med samme aktivitetsniveau.

Hovedfundene i studiet var, at kollagen koncentrationen var lavere hos ældre mænd sammenlignet med unge mænd, mens enzymatisk-dannede tværbindinger var højere hos ældre mænd end unge mænd. Ligeledes var den non-enzymatiske AGE tværbinding markør (pentosidine) væsentligt forhøjet hos ældre sammenlignet med unge mænd. På trods af de markante aldersrelaterede forskelle i senens koncentration af kollagen og tværbindinger, var der ingen forskel i de mekaniske egenskaber mellem de to grupper.

### *Studie III*

Formålet med dette studie var at undersøge effekterne af kort-tids (14 dage) unilateral immobilisering på den humane patellasenes strukturelle og mekaniske egenskaber hos ældre mænd, *in vivo*.

Hovedfundene i studiet var en nedgang i sene stivhed og Youngs Modulus efter immobilisering uden en samtidig ændring af patellasens dimensioner, hvilket tyder på, at patellasens materiale egenskaber var ændret efter immobiliseringen, som følge af mindre aktivitet.

## Reference List

- (1) Bailey AJ, Paul RG, Knott L. Mechanisms of maturation and ageing of collagen. *Mech Ageing Dev* 1998 Dec 1;106(1-2):1-56.
- (2) Cerami A, Vlassara H, Brownlee M. Glucose and aging. *Sci Am* 1987 May;256(5):90-6.
- (3) Khan KM, Maffulli N, Coleman BD, Cook JL, Taunton JE. Patellar tendinopathy: some aspects of basic science and clinical management. *Br J Sports Med* 1998 Dec;32(4):346-55.
- (4) Jozsa L, Lehto M, Kvist M, Balint JB, Reffy A. Alterations in dry mass content of collagen fibers in degenerative tendinopathy and tendon-rupture. *Matrix* 1989 Mar;9(2):140-6.
- (5) Prockop DJ, Kivirikko KI. Collagens: molecular biology, diseases, and potentials for therapy. *Annu Rev Biochem* 1995;64:403-34.:403-34.
- (6) Kjaer M. Role of extracellular matrix in adaptation of tendon and skeletal muscle to mechanical loading. *Physiol Rev* 2004 Apr;84(2):649-98.
- (7) Eyre DR. Collagens and cartilage matrix homeostasis. *Clin Orthop Relat Res* 2004 Oct;(427 Suppl):S118-S122.
- (8) Parry DA, Squire JM. Fibrous proteins: new structural and functional aspects revealed. *Adv Protein Chem* 2005;70:1-10.
- (9) Amiel D, Frank C, Harwood F, Fronck J, Akeson W. Tendons and ligaments: a morphological and biochemical comparison. *J Orthop Res* 1984;1(3):257-65.
- (10) Avery NC, Bailey AJ. Enzymic and non-enzymic cross-linking mechanisms in relation to turnover of collagen: relevance to aging and exercise. *Scand J Med Sci Sports* 2005 Aug;15(4):231-40.
- (11) Bailey AJ, Robins SP, Balian G. Biological significance of the intermolecular crosslinks of collagen. *Nature* 1974 Sep 13;251(5471):105-9.
- (12) Bailey AJ. Molecular mechanisms of ageing in connective tissues. *Mech Ageing Dev* 2001 May 31;122(7):735-55.
- (13) Eyre DR, Paz MA, Gallop PM. Cross-linking in collagen and elastin. *Annu Rev Biochem* 1984;53:717-48.
- (14) Fujimoto D, Moriguchi T, Ishida T, Hayashi H. The structure of pyridinoline, a collagen crosslink. *Biochem Biophys Res Commun* 1978 Sep 14;84(1):52-7.
- (15) Monnier VM. Toward a Maillard reaction theory of aging. *Prog Clin Biol Res* 1989;304:1-22.
- (16) Myllyharju J, Kivirikko KI. Collagens and collagen-related diseases. *Ann Med* 2001 Feb;33(1):7-21.

- (17) Kastelic J, Galeski A, Baer E. The multicomposite structure of tendon. *Connect Tissue Res* 1978;6(1):11-23.
- (18) Diamant J, Keller A, Baer E, Litt M, Arridge RG. Collagen; ultrastructure and its relation to mechanical properties as a function of ageing. *Proc R Soc Lond B Biol Sci* 1972 Mar 14;180(60):293-315.
- (19) Dale WC, Baer E, Keller A, Kohn RR. On the ultrastructure of mammalian tendon. *Experientia* 1972 Nov 15;28(11):1293-5.
- (20) Viidik A. Functional properties of collagenous tissues. *Int Rev Connect Tissue Res* 1973;6:127-215.
- (21) Kannus P. Structure of the tendon connective tissue. *Scand J Med Sci Sports* 2000 Dec;10(6):312-20.
- (22) Alexander RM, net-Clark HC. Storage of elastic strain energy in muscle and other tissues. *Nature* 1977 Jan 13;265(5590):114-7.
- (23) Shadwick RE. Elastic energy storage in tendons: mechanical differences related to function and age. *J Appl Physiol* 1990 Mar;68(3):1033-40.
- (24) Cavagna GA. Storage and utilization of elastic energy in skeletal muscle. *Exerc Sport Sci Rev* 1977;5:89-129.
- (25) Biewener A, Baudinette R. In vivo muscle force and elastic energy storage during steady-speed hopping of tammar wallabies (*Macropus eugenii*). *J Exp Biol* 1995;198(Pt 9):1829-41.
- (26) Scott SH, Loeb GE. Mechanical properties of aponeurosis and tendon of the cat soleus muscle during whole-muscle isometric contractions. *J Morphol* 1995 Apr;224(1):73-86.
- (27) Buchanan CI, Marsh RL. Effects of long-term exercise on the biomechanical properties of the Achilles tendon of guinea fowl. *J Appl Physiol* 2001 Jan;90(1):164-71.
- (28) Bramble DM, Lieberman DE. Endurance running and the evolution of Homo. *Nature* 2004 Nov 18;432(7015):345-52.
- (29) Thorpe SK, Crompton RH, Gunther MM, Ker RF, McNeill AR. Dimensions and moment arms of the hind- and forelimb muscles of common chimpanzees (*Pan troglodytes*). *Am J Phys Anthropol* 1999 Oct;110(2):179-99.
- (30) Fukunaga T, Kubo K, Kawakami Y, Fukashiro S, Kanehisa H, Maganaris CN. In vivo behaviour of human muscle tendon during walking. *Proc Biol Sci* 2001 Feb 7;268(1464):229-33.
- (31) Kawakami Y, Muraoka T, Ito S, Kanehisa H, Fukunaga T. In vivo muscle fibre behaviour during counter-movement exercise in humans reveals a significant role for tendon elasticity. *J Physiol (Lond)* 2002 Apr 15;540(2):635-46.

- (32) Kurokawa S, Fukunaga T, Nagano A, Fukashiro S. Interaction between fascicles and tendinous structures during counter movement jumping investigated in vivo. *J Appl Physiol* 2003 Dec 1;95(6):2306-14.
- (33) Voigt M, Bojsen-Moller F, Simonsen EB, Dyhre-Poulsen P. The influence of tendon Young's modulus, dimensions and instantaneous moment arms on the efficiency of human movement. *J Biomech* 1995 Mar;28(3):281-91.
- (34) Maganaris CN, Paul JP. Tensile properties of the in vivo human gastrocnemius tendon. *J Biomech* 2002 Dec;35(12):1639-46.
- (35) Lichtwark GA, Wilson AM. In vivo mechanical properties of the human Achilles tendon during one-legged hopping. *J Exp Biol* 2005 Dec;208(Pt 24):4715-25.
- (36) Ishikawa M, Komi PV. Effects of different dropping intensities on fascicle and tendinous tissue behavior during stretch-shortening cycle exercise. *J Appl Physiol* 2004 Mar 1;96(3):848-52.
- (37) McHugh MP, Connolly DA, Eston RG, Kremenic IJ, Nicholas SJ, Gleim GW. The role of passive muscle stiffness in symptoms of exercise-induced muscle damage. *Am J Sports Med* 1999 Sep;27(5):594-9.
- (38) Griffiths RI. Shortening of muscle fibres during stretch of the active cat medial gastrocnemius muscle: the role of tendon compliance. *J Physiol* 1991 May;436:219-36.
- (39) Lieber RL, Leonard ME, Brown-Maupin CG. Effects of muscle contraction on the load-strain properties of frog aponeurosis and tendon. *Cells Tissues Organs* 2000;166(1):48-54.
- (40) Butler DL, Grood ES, Noyes FR, Zernicke RF, Brackett K. Effects of structure and strain measurement technique on the material properties of young human tendons and fascia. *J Biomech* 1984;17(8):579-96.
- (41) Oxlund H. Relationships between the biomechanical properties, composition and molecular structure of connective tissues. *Connect Tissue Res* 1986;15(1-2):65-72.
- (42) Wren TA, Beaupre GS, Carter DR. Tendon and ligament adaptation to exercise, immobilization, and remobilization. *J Rehabil Res Dev* 2000 Mar;37(2):217-24.
- (43) ELLIOTT DH, CRAWFORD GN. THE THICKNESS AND COLLAGEN CONTENT OF TENDON RELATIVE TO THE CROSS-SECTIONAL AREA OF MUSCLE DURING GROWTH. *Proc R Soc Lond B Biol Sci* 1965 Apr 13;162:198-202.
- (44) Zernicke RF, Garhammer J, Jobe FW. Human patellar-tendon rupture. *J Bone Joint Surg Am* 1977 Mar;59(2):179-83.
- (45) Bobbert MF, Huijting PA, van Ingen Schenau GJ. Drop jumping. II. The influence of dropping height on the biomechanics of drop jumping. *Med Sci Sports Exerc* 1987 Aug;19(4):339-46.

- (46) Richards DP, Ajemian SV, Wiley JP, Zernicke RF. Knee joint dynamics predict patellar tendinitis in elite volleyball players. *Am J Sports Med* 1996 Sep;24(5):676-83.
- (47) Wahrenberg H, Lindbeck L, Ekholm J. Knee muscular moment, tendon tension force and EMG during a vigorous movement in man. *Scand J Rehabil Med* 1978;10(2):99-106.
- (48) Wahrenberg H, Lindbeck L, Ekholm J. Dynamic load in the human knee joint during voluntary active impact to the lower leg. *Scand J Rehabil Med* 1978;10(2):93-8.
- (49) Komi PV, Fukashiro S, Jarvinen M. Biomechanical loading of Achilles tendon during normal locomotion. *Clin Sports Med* 1992 Jul;11(3):521-31.
- (50) Ishikawa M, Komi PV, Finni T, Kuitunen S. Contribution of the tendinous tissue to force enhancement during stretch-shortening cycle exercise depends on the prestretch and concentric phase intensities. *J Electromyogr Kinesiol* 2006 Oct;16(5):423-31.
- (51) Krevolin JL, Pandy MG, Pearce JC. Moment arm of the patellar tendon in the human knee. *J Biomech* 2004 May;37(5):785-8.
- (52) Butler DL, Grood ES, Noyes FR, Zernicke RF. Biomechanics of ligaments and tendons. *Exerc Sport Sci Rev* 1978;6:125-81.
- (53) Alkjaer T, Simonsen EB, Peter Magnusson SP, Aagaard H, Dyhre-Poulsen P. Differences in the movement pattern of a forward lunge in two types of anterior cruciate ligament deficient patients: copers and non-copers. *Clin Biomech (Bristol , Avon )* 2002 Oct;17(8):586-93.
- (54) Cronin J, McNair PJ, Marshall RN. Lunge performance and its determinants. *J Sports Sci* 2003 Jan;21(1):49-57.
- (55) Hefzy MS, al KM, Harrison L. Co-activation of the hamstrings and quadriceps during the lunge exercise. *Biomed Sci Instrum* 1997;33:360-5.
- (56) Klinger A, Adrian MJ. Foil target impact forces during the fencing lunge. *Proceedings of the Eighth International Congress of Biomechanics; Champaign, IL: Human Kinetics; 2010 p. 882-8.*
- (57) Aagaard P, Andersen JL, Dyhre-Poulsen P, Leffers AM, Wagner A, Magnusson SP, et al. A mechanism for increased contractile strength of human pennate muscle in response to strength training: changes in muscle architecture. *J Physiol* 2001 Jul 15;534(Pt. 2):613-23.
- (58) Narici MV, Hoppeler H, Kayser B, Landoni L, Claassen H, Gavardi C, et al. Human quadriceps cross-sectional area, torque and neural activation during 6 months strength training. *Acta Physiol Scand* 1996 Jun;157(2):175-86.
- (59) Jones HH, Priest JD, Hayes WC, Tichenor CC, Nagel DA. Humeral hypertrophy in response to exercise. *J Bone Joint Surg Am* 1977 Mar;59(2):204-8.
- (60) Krahl H, Michaelis U, Pieper HG, Quack G, Montag M. Stimulation of bone growth through sports. A radiologic investigation of the upper extremities in professional tennis players. *Am J Sports Med* 1994 Nov;22(6):751-7.

- (61) ELLIOTT DH. STRUCTURE AND FUNCTION OF MAMMALIAN TENDON. *Biol Rev Camb Philos Soc* 1965 Aug;40:392-421.
- (62) Kongsgaard M, Aagaard P, Kjaer M, Magnusson SP. Structural Achilles tendon properties in athletes subjected to different exercise modes and in Achilles tendon rupture patients. *J Appl Physiol* 2005 Nov;99(5):1965-71.
- (63) Muraoka T, Muramatsu T, Fukunaga T, Kanehisa H. Elastic properties of human Achilles tendon are correlated to muscle strength. *J Appl Physiol* 2005 Aug;99(2):665-9.
- (64) Latimer B, Lovejoy CO. The calcaneus of *Australopithecus afarensis* and its implications for the evolution of bipedality. *Am J Phys Anthropol* 1989 Mar;78(3):369-86.
- (65) Bojsen-Moller J, Kalliokoski KK, Seppanen M, Kjaer M, Magnusson SP. Low-intensity tensile loading increases intratendinous glucose uptake in the Achilles tendon. *J Appl Physiol* 2006 Jul;101(1):196-201.
- (66) Hannukainen J, Kalliokoski KK, Nuutila P, Fujimoto T, Kemppainen J, Viljanen T, et al. In vivo measurements of glucose uptake in human Achilles tendon during different exercise intensities. *Int J Sports Med* 2005 Nov;26(9):727-31.
- (67) Kalliokoski KK, Langberg H, Ryberg AK, Scheede-Bergdahl C, Doessing S, Kjaer A, et al. The effect of dynamic knee-extension exercise on patellar tendon and quadriceps femoris muscle glucose uptake in humans studied by positron emission tomography. *J Appl Physiol* 2005 Sep;99(3):1189-92.
- (68) Langberg H, Skovgaard D, Petersen LJ, Bulow J, Kjaer M. Type I collagen synthesis and degradation in peritendinous tissue after exercise determined by microdialysis in humans. *J Physiol* 1999 Nov 15;521 Pt 1:299-306.
- (69) Miller BF, Olesen JL, Hansen M, Dossing S, Cramer RM, Welling RJ, et al. Coordinated collagen and muscle protein synthesis in human patella tendon and quadriceps muscle after exercise. *J Physiol* 2005 Sep 15;567(Pt 3):1021-33.
- (70) Langberg H, Rosendal L, Kjaer M. Training-induced changes in peritendinous type I collagen turnover determined by microdialysis in humans. *J Physiol* 2001 Jul 1;534(Pt 1):297-302.
- (71) Birch HL, McLaughlin L, Smith RK, Goodship AE. Treadmill exercise-induced tendon hypertrophy: assessment of tendons with different mechanical functions. *Equine Vet J Suppl* 1999 Jul;30:222-6.
- (72) Woo SL, Gomez MA, Woo YK, Akeson WH. Mechanical properties of tendons and ligaments. II. The relationships of immobilization and exercise on tissue remodeling. *Biorheology* 1982;19(3):397-408.
- (73) Sommer HM. The biomechanical and metabolic effects of a running regime on the Achilles tendon in the rat. *Int Orthop* 1987;11(1):71-5.

- (74) Viidik A. Tensile strength properties of Achilles tendon systems in trained and untrained rabbits. *Acta Orthop Scand* 1969;40(2):261-72.
- (75) Smith RK, Birch HL, Goodman S, Heinegard D, Goodship AE. The influence of ageing and exercise on tendon growth and degeneration--hypotheses for the initiation and prevention of strain-induced tendinopathies. *Comp Biochem Physiol A Mol Integr Physiol* 2002 Dec;133(4):1039-50.
- (76) Vailas AC, Pedrini VA, Pedrini-Mille A, Holloszy JO. Patellar tendon matrix changes associated with aging and voluntary exercise. *J Appl Physiol* 1985 May;58(5):1572-6.
- (77) Woo SL. Mechanical properties of tendons and ligaments. I. Quasi-static and nonlinear viscoelastic properties. *Biorheology* 1982;19(3):385-96.
- (78) Rosager S, Aagaard P, Dyhre-Poulsen P, Neergaard K, Kjaer M, Magnusson SP. Load-displacement properties of the human triceps surae aponeurosis and tendon in runners and non-runners. *Scand J Med Sci Sports* 2002 Apr;12(2):90-8.
- (79) Magnusson SP, Kjaer M. Region-specific differences in Achilles tendon cross-sectional area in runners and non-runners. *Eur J Appl Physiol* 2003 Nov;90(5-6):549-53.
- (80) Hansen P, Aagaard P, Kjaer M, Larsson B, Magnusson SP. Effect of habitual running on human Achilles tendon load-deformation properties and cross-sectional area. *J Appl Physiol* 2003 Dec;95(6):2375-80.
- (81) Stone MH. Implications for connective tissue and bone alterations resulting from resistance exercise training. *Med Sci Sports Exerc* 1988 Oct;20(5 Suppl):S162-S168.
- (82) Reeves ND, Maganaris CN, Narici MV. Effect of strength training on human patella tendon mechanical properties of older individuals. *J Physiol (Lond)* 2003 May 1;548(3):971-81.
- (83) Reeves ND, Narici MV, Maganaris CN. Strength training alters the viscoelastic properties of tendons in elderly humans. *Muscle Nerve* 2003 Jul;28(1):74-81.
- (84) Kubo K, Yata H, Kanehisa H, Fukunaga T. Effects of isometric squat training on the tendon stiffness and jump performance. *Eur J Appl Physiol* 2006 Feb;96(3):305-14.
- (85) Kubo K, Kanehisa H, Fukunaga T. Effects of different duration isometric contractions on tendon elasticity in human quadriceps muscles. *J Physiol (Lond)* 2001 Oct 15;536(2):649-55.
- (86) Kubo K, Kanehisa H, Fukunaga T. Effects of resistance and stretching training programmes on the viscoelastic properties of human tendon structures in vivo. *J Physiol (Lond)* 2002 Jan 1;538(1):219-26.
- (87) Kongsgaard M, Reitelseder S, Pedersen TG, Holm L, Aagaard P, Kjaer M, et al. Region specific patellar tendon hypertrophy in humans following resistance training. *Acta Physiol (Oxf)* 2007 Oct;191(2):111-21.

- (88) Arampatzis A, Karamanidis K, Albracht K. Adaptational responses of the human Achilles tendon by modulation of the applied cyclic strain magnitude. *J Exp Biol* 2007 Aug;210(Pt 15):2743-53.
- (89) Seynnes OR, Erskine RM, Maganaris CN, Longo S, Simoneau EM, Grosset JF, et al. Training-induced changes in structural and mechanical properties of the patellar tendon are related to muscle hypertrophy, but not to strength gains. *J Appl Physiol* 2009 May 28.
- (90) Coupe C, Kongsgaard M, Aagaard P, Hansen P, Bojsen-Moller J, Kjaer M, et al. Habitual loading results in tendon hypertrophy and increased stiffness of the human patellar tendon. *J Appl Physiol* 2008 Sep;105(3):805-10.
- (91) Narici MV, Maganaris C, Reeves N. Myotendinous alterations and effects of resistive loading in old age. *Scand J Med Sci Sports* 2005 Dec;15(6):392-401.
- (92) Narici MV, Maganaris CN. Plasticity of the muscle-tendon complex with disuse and aging. *Exerc Sport Sci Rev* 2007 Jul;35(3):126-34.
- (93) Magnusson SP, Narici MV, Maganaris CN, Kjaer M. Human tendon behaviour and adaptation, in vivo. *J Physiol* 2008 Jan 1;586(1):71-81.
- (94) VERZAR F. Aging of the collagen fiber. *Int Rev Connect Tissue Res* 1964;2:243-300.
- (95) Robert L. Fritz Verzar was born 120 years ago: his contribution to experimental gerontology through the collagen research as assessed after half a century. *Arch Gerontol Geriatr* 2006 Jul;43(1):13-43.
- (96) Nielsen HM, Skalicky M, Viidik A. Influence of physical exercise on aging rats. III. Life-long exercise modifies the aging changes of the mechanical properties of limb muscle tendons. *Mech Ageing Dev* 1998 Feb 16;100(3):243-60.
- (97) Galeski A, Kastelic J, Baer E, Kohn RR. Mechanical and structural changes in rat tail tendon induced by alloxan diabetes and aging. *J Biomech* 1977;10(11/12):775-82.
- (98) Dressler MR, Butler DL, Wenstrup R, Awad HA, Smith F, Boivin GP. A potential mechanism for age-related declines in patellar tendon biomechanics. *J Orthop Res* 2002 Nov;20(6):1315-22.
- (99) Vogel HG. Influence of maturation and age on mechanical and biochemical parameters of connective tissue of various organs in the rat. *Connect Tissue Res* 1978;6(3):161-6.
- (100) Vogel HG. Age dependence of mechanical properties of rat tail tendons (hysteresis experiments). *Aktuelle Gerontol* 1983 Jan;13(1):22-7.
- (101) Haut RC, Lancaster RL, DeCamp CE. Mechanical properties of the canine patellar tendon: some correlations with age and the content of collagen. *J Biomech* 1992 Feb;25(2):163-73.
- (102) Hubbard RP, Soutas-Little RW. Mechanical properties of human tendon and their age dependence. *J Biomech Eng* 1984 May;106(2):144-50.

- (103) Flahiff CM, Brooks AT, Hollis JM, Vander Schilden JL, Nicholas RW. Biomechanical analysis of patellar tendon allografts as a function of donor age. *Am J Sports Med* 1995 May;23(3):354-8.
- (104) Johnson GA, Tramaglino DM, Levine RE, Ohno K, Choi NY, Woo SL. Tensile and viscoelastic properties of human patellar tendon. *J Orthop Res* 1994 Nov;12(6):796-803.
- (105) Hansen P, Bojsen-Moller J, Aagaard P, Kjaer M, Magnusson SP. Mechanical properties of the human patellar tendon, in vivo. *Clin Biomech (Bristol , Avon)* 2005 Sep 22.
- (106) Fukashiro S, Itoh M, Ichinose Y, Kawakami Y, Fukunaga T. Ultrasonography gives directly but noninvasively elastic characteristic of human tendon in vivo. *Eur J Appl Physiol Occup Physiol* 1995;71(6):555-7.
- (107) Maganaris CN, Paul JP. In vivo human tendon mechanical properties. *J Physiol* 1999 Nov 15;521 Pt 1:307-13.
- (108) Kubo K, Ishida Y, Komuro T, Tsunoda N, Kanehisa H, Fukunaga T. Age-related differences in the force generation capabilities and tendon extensibilities of knee extensors and plantar flexors in men. *J Gerontol A Biol Sci Med Sci* 2007 Nov;62(11):1252-8.
- (109) Kubo K, Morimoto M, Komuro T, Tsunoda N, Kanehisa H, Fukunaga T. Age-related differences in the properties of the plantar flexor muscles and tendons. *Med Sci Sports Exerc* 2007 Mar;39(3):541-7.
- (110) Kubo K, Kanehisa H, Miyatani M, Tachi M, Fukunaga T. Effect of low-load resistance training on the tendon properties in middle-aged and elderly women. *Acta Physiol Scand* 2003 May;178(1):25-32.
- (111) Onambele GL, Narici MV, Maganaris CN. Calf muscle-tendon properties and postural balance in old age. *J Appl Physiol* 2006 Jun;100(6):2048-56.
- (112) Mademli L, Arampatzis A, Walsh M. Age-related effect of static and cyclic loadings on the strain-force curve of the vastus lateralis tendon and aponeurosis. *J Biomech Eng* 2008 Feb;130(1):011007.
- (113) Morse CI, Thom JM, Birch KM, Narici MV. Tendon elongation influences the amplitude of interpolated doublets in the assessment of activation in elderly men. *J Appl Physiol* 2005 Jan;98(1):221-6.
- (114) Mian OS, Thom JM, Ardigo LP, Minetti AE, Narici MV. Gastrocnemius muscle-tendon behaviour during walking in young and older adults. *Acta Physiol (Oxf)* 2007 Jan;189(1):57-65.
- (115) Karamanidis K, Arampatzis A. Mechanical and morphological properties of human quadriceps femoris and triceps surae muscle-tendon unit in relation to aging and running. *J Biomech* 2006;39(3):406-17.

- (116) Carroll CC, Dickinson JM, Haus JM, Lee GA, Hollon CJ, Aagaard P, et al. Influence of aging on the in vivo properties of human patellar tendon. *J Appl Physiol* 2008 Dec;105(6):1907-15.
- (117) Barnard K, Light ND, Sims TJ, Bailey AJ. Chemistry of the collagen cross-links. Origin and partial characterization of a putative mature cross-link of collagen. *Biochem J* 1987 Jun 1;244(2):303-9.
- (118) Maillard L. Action des acides aminés sur les sucres: formation des mélanoidines par voie méthodique. *C R Acad Sci* 1912;154:66-8.
- (119) Sell DR, Nagaraj RH, Grandhee SK, Odetti P, Lapolla A, Fogarty J, et al. Pentosidine: a molecular marker for the cumulative damage to proteins in diabetes, aging, and uremia. *Diabetes Metab Rev* 1991 Dec;7(4):239-51.
- (120) Monnier VM, Cerami A. Nonenzymatic browning in vivo: possible process for aging of long-lived proteins. *Science* 1981 Jan 30;211(4481):491-3.
- (121) Dyer DG, Dunn JA, Thorpe SR, Bailie KE, Lyons TJ, McCance DR, et al. Accumulation of Maillard reaction products in skin collagen in diabetes and aging. *J Clin Invest* 1993 Jun;91(6):2463-9.
- (122) Brownlee M. Advanced protein glycosylation in diabetes and aging. *Annu Rev Med* 1995;46:223-34.
- (123) Reddy GK, Stehno-Bittel L, Enwemeka CS. Glycation-induced matrix stability in the rabbit achilles tendon. *Arch Biochem Biophys* 2002 Mar 15;399(2):174-80.
- (124) Reddy GK. Cross-linking in collagen by nonenzymatic glycation increases the matrix stiffness in rabbit achilles tendon. *Exp Diabetes Res* 2004 Apr;5(2):143-53.
- (125) Verzijl N, DeGroot J, Oldehinkel E, Bank RA, Thorpe SR, Baynes JW, et al. Age-related accumulation of Maillard reaction products in human articular cartilage collagen. *Biochem J* 2000 Sep 1;350 Pt 2:381-7.
- (126) Chen JR, Takahashi M, Kushida K, Suzuki M, Suzuki K, Horiuchi K, et al. Direct detection of crosslinks of collagen and elastin in the hydrolysates of human yellow ligament using single-column high performance liquid chromatography. *Anal Biochem* 2000 Feb 15;278(2):99-105.
- (127) Haus JM, Carrithers JA, Trappe SW, Trappe TA. Collagen, cross-linking, and advanced glycation end products in aging human skeletal muscle. *J Appl Physiol* 2007 Dec;103(6):2068-76.
- (128) Saito M, Marumo K, Fujii K, Ishioka N. Single-column high-performance liquid chromatographic-fluorescence detection of immature, mature, and senescent cross-links of collagen. *Anal Biochem* 1997 Nov 1;253(1):26-32.

- (129) Suzuki D, Takahashi M, Abe M, Nagano A. Biochemical study of collagen and its crosslinks in the anterior cruciate ligament and the tissues used as a graft for reconstruction of the anterior cruciate ligament. *Connect Tissue Res* 2008;49(1):42-7.
- (130) Bank RA, TeKoppele JM, Oostingh G, Hazleman BL, Riley GP. Lysylhydroxylation and non-reducible crosslinking of human supraspinatus tendon collagen: changes with age and in chronic rotator cuff tendinitis. *Ann Rheum Dis* 1999 Jan;58(1):35-41.
- (131) Narici MV, Maganaris CN. Adaptability of elderly human muscles and tendons to increased loading. *J Anat* 2006 Apr;208(4):433-43.
- (132) Suetta C, Hvid LG, Justesen L, Christensen U, Neergaard K, Simonsen L, et al. Effects of ageing on human skeletal muscle after immobilisation and re-training. *J Appl Physiol* 2009 Aug 6.
- (133) Booth FW, Gould EW. Effects of training and disuse on connective tissue. *Exerc Sport Sci Rev* 1975;3:83-112.
- (134) Tipton CM, Vailas AC, Matthes RD. Experimental studies on the influences of physical activity on ligaments, tendons and joints: a brief review. *Acta Med Scand Suppl* 1986;711:157-68.:157-68.
- (135) Yamamoto N, Ohno K, Hayashi K, Kuriyama H, Yasuda K, Kaneda K. Effects of stress shielding on the mechanical properties of rabbit patellar tendon. *J Biomech Eng* 1993 Feb;115(1):23-8.
- (136) Matsumoto F, Trudel G, Uthoff HK, Backman DS. Mechanical effects of immobilization on the Achilles' tendon. *Arch Phys Med Rehabil* 2003 May;84(5):662-7.
- (137) Rumian AP, Draper ER, Wallace AL, Goodship AE. The influence of the mechanical environment on remodelling of the patellar tendon. *J Bone Joint Surg Br* 2009 Apr;91(4):557-64.
- (138) meida-Silveira MI, Lambertz D, Perot C, Goubel F. Changes in stiffness induced by hindlimb suspension in rat Achilles tendon. *Eur J Appl Physiol* 2000 Feb;81(3):252-7.
- (139) Hannafin JA, Arnoczky SP, Hoonjan A, Torzilli PA. Effect of stress deprivation and cyclic tensile loading on the material and morphologic properties of canine flexor digitorum profundus tendon: an in vitro study. *J Orthop Res* 1995 Nov;13(6):907-14.
- (140) Loitz BJ, Zernicke RF, Vailas AC, Kody MH, Meals RA. Effects of short-term immobilization versus continuous passive motion on the biomechanical and biochemical properties of the rabbit tendon. *Clin Orthop Relat Res* 1989 Jul;(244):265-71.
- (141) Eliasson P, Fahlgren A, Pasternak B, Aspenberg P. Unloaded rat Achilles tendons continue to grow, but lose viscoelasticity. *J Appl Physiol* 2007 Aug;103(2):459-63.
- (142) Arruda EM, Calve S, Dennis RG, Mundy K, Baar K. Regional variation of tibialis anterior tendon mechanics is lost following denervation. *J Appl Physiol* 2006 Oct;101(4):1113-7.

- (143) Reeves ND, Maganaris CN, Ferretti G, Narici MV. Influence of 90-day simulated microgravity on human tendon mechanical properties and the effect of resistive countermeasures. *J Appl Physiol* 2005 Jun;98(6):2278-86.
- (144) Shin D, Finni T, Ahn S, Hodgson JA, Lee HD, Edgerton VR, et al. Effect of chronic unloading and rehabilitation on human Achilles tendon properties: a velocity-encoded phase-contrast MRI study. *J Appl Physiol* 2008 Oct;105(4):1179-86.
- (145) de B, Maganaris CN, Seynnes OR, Rennie MJ, Narici MV. Time course of muscular, neural and tendinous adaptations to 23 day unilateral lower-limb suspension in young men. *J Physiol* 2007 Sep 15;583(Pt 3):1079-91.
- (146) Kubo K, Akima H, Kouzaki M, Ito M, Kawakami Y, Kanehisa H, et al. Changes in the elastic properties of tendon structures following 20 days bed-rest in humans. *Eur J Appl Physiol* 2000 Dec;83(6):463-8.
- (147) Kubo K, Akima H, Ushiyama J, Tabata I, Fukuoka H, Kanehisa H, et al. Effects of resistance training during bed rest on the viscoelastic properties of tendon structures in the lower limb. *Scand J Med Sci Sports* 2004 Oct;14(5):296-302.
- (148) Zhao H, Ren Y, Wu YN, Liu SQ, Zhang LQ. Ultrasonic evaluations of Achilles tendon mechanical properties poststroke. *J Appl Physiol* 2009 Mar;106(3):843-9.
- (149) Seynnes OR, Maffiuletti NA, Maganaris CN, de B, Pensini M, di Prampero PE, et al. Soleus T reflex modulation in response to spinal and tendinous adaptations to unilateral lower limb suspension in humans. *Acta Physiol (Oxf)* 2008 Nov;194(3):239-51.
- (150) Creditor MC. Hazards of hospitalization of the elderly. *Ann Intern Med* 1993 Feb 1;118(3):219-23.
- (151) Covinsky KE, Palmer RM, Fortinsky RH, Counsell SR, Stewart AL, Kresevic D, et al. Loss of independence in activities of daily living in older adults hospitalized with medical illnesses: increased vulnerability with age. *J Am Geriatr Soc* 2003 Apr;51(4):451-8.
- (152) Deschenes MR, Holdren AN, McCoy RW. Adaptations to short-term muscle unloading in young and aged men. *Med Sci Sports Exerc* 2008 May;40(5):856-63.
- (153) Kubo K, Akima H, Ushiyama J, Tabata I, Fukuoka H, Kanehisa H, et al. Effects of 20 days of bed rest on the viscoelastic properties of tendon structures in lower limb muscles. *Br J Sports Med* 2004 Jun;38(3):324-30.
- (154) Maganaris CN, Reeves ND, Rittweger J, Sargeant AJ, Jones DA, Gerrits K, et al. Adaptive response of human tendon to paralysis. *Muscle Nerve* 2006 Jan;33(1):85-92.
- (155) Magnusson SP, Aagaard P, Dyhre-Poulsen P, Kjaer M. Load-displacement properties of the human triceps surae aponeurosis in vivo. *J Physiol* 2001 Feb 15;531(Pt 1):277-88.
- (156) Sapega AA, Minkoff J, Valsamis M, Nicholas JA. Musculoskeletal performance testing and profiling of elite competitive fencers. *Clin Sports Med* 1984 Jan;3(1):231-44.

- (157) Nystrom J, Lindwall O, Ceci R, Harmenberg J, Svedenhag J, Ekblom B. Physiological and morphological characteristics of world class fencers. *Int J Sports Med* 1990 Apr;11(2):136-9.
- (158) Nordstrom P, Pettersson U, Lorentzon R. Type of physical activity, muscle strength, and pubertal stage as determinants of bone mineral density and bone area in adolescent boys. *J Bone Miner Res* 1998 Jul;13(7):1141-8.
- (159) Kongsgaard M, Kovanen V, Aagaard P, Doessing S, Hansen P, Kjaer M, et al. Peritendinous Corticosteroid Injections, Eccentric Decline squat training and heavy slow resistance training in patellar tendinopathy. *Scand J Med Sci Sports*. In press 2009.
- (160) Warden SJ, Kiss ZS, Malara FA, Ooi AB, Cook JL, Crossley KM. Comparative accuracy of magnetic resonance imaging and ultrasonography in confirming clinically diagnosed patellar tendinopathy. *Am J Sports Med* 2007 Mar;35(3):427-36.
- (161) Saltin B, Grimby G. Physiological analysis of middle-aged and old former athletes. Comparison with still active athletes of the same ages. *Circulation* 1968 Dec;38(6):1104-15.
- (162) Roberts CS, King DH, Goldsmith LJ. A statistical analysis of the accuracy of sonography of the patellar tendon. *Arthroscopy* 1999 May;15(4):388-91.
- (163) Shalabi A, Kristoffersen-Wiberg M, Aspelin P, Movin T. Immediate Achilles tendon response after strength training evaluated by MRI. *Med Sci Sports Exerc* 2004 Nov;36(11):1841-6.
- (164) Boesen MI, Boesen A, Koenig MJ, Bliddal H, Torp-Pedersen S. Ultrasonographic investigation of the Achilles tendon in elite badminton players using color Doppler. *Am J Sports Med* 2006 Dec;34(12):2013-21.
- (165) Ferretti A. Epidemiology of jumper's knee. *Sports Med* 1986 Jul;3(4):289-95.
- (166) Blazina ME, Kerlan RK, Jobe FW, Carter VS, Carlson GJ. Jumper's knee. *Orthop Clin North Am* 1973 Jul;4(3):665-78.
- (167) Markovic G, Jaric S. Movement performance and body size: the relationship for different groups of tests. *Eur J Appl Physiol* 2004 Jun;92(1-2):139-49.
- (168) Bojsen-Moller J, Hansen P, Aagaard P, Kjaer M, Magnusson SP. Measuring mechanical properties of the vastus lateralis tendon-aponeurosis complex in vivo by ultrasound imaging. *Scand J Med Sci Sports* 2003 Aug;13(4):259-65.
- (169) Visser JJ, Hoogkamer JE, Bobbert MF, Huijting PA. Length and moment arm of human leg muscles as a function of knee and hip-joint angles. *Eur J Appl Physiol Occup Physiol* 1990;61(5-6):453-60.
- (170) Magnusson SP, Hansen P, Aagaard P, Brond J, Dyhre-Poulsen P, Bojsen-Moller J, et al. Differential strain patterns of the human gastrocnemius aponeurosis and free tendon, in vivo. *Acta Physiol Scand* 2003 Feb;177(2):185-95.

- (171) Onambele-Pearson NL, Pearson SJ. Time-of-day effect on patella tendon stiffness alters vastus lateralis fascicle length but not the quadriceps force-angle relationship. *J Biomech* 2007;40(5):1031-7.
- (172) Pearson SJ, Burgess K, Onambele GN. Creep and the in vivo assessment of human patellar tendon mechanical properties. *Clin Biomech (Bristol , Avon )* 2007 Jul;22(6):712-7.
- (173) Onambele GN, Burgess K, Pearson SJ. Gender-specific in vivo measurement of the structural and mechanical properties of the human patellar tendon. *J Orthop Res* 2007 Dec;25(12):1635-42.
- (174) Rigby BJ, Hirai N, Spikes JD, Eyring H. The Mechanical Properties of Rat Tail Tendon. *J Gen Physiol* 1959 Nov 1;43(2):265-83.
- (175) Kubo K, Morimoto M, Komuro T, Yata H, Tsunoda N, Kanehisa H, et al. Effects of plyometric and weight training on muscle-tendon complex and jump performance. *Med Sci Sports Exerc* 2007 Oct;39(10):1801-10.
- (176) Creemers LB, Jansen DC, van Veen-Reurings A, van den BT, Everts V. Microassay for the assessment of low levels of hydroxyproline. *Biotechniques* 1997 Apr;22(4):656-8.
- (177) Bank RA, Beekman B, Verzijl N, de Roos JA, Sakkee AN, TeKoppele JM. Sensitive fluorimetric quantitation of pyridinium and pentosidine crosslinks in biological samples in a single high-performance liquid chromatographic run. *J Chromatogr B Biomed Sci Appl* 1997 Dec 5;703(1-2):37-44.
- (178) Curwin SL, Roy RR, Vailas AC. Regional and age variations in growing tendon. *J Morphol* 1994 Sep;221(3):309-20.
- (179) Malaviya P, Butler DL, Boivin GP, Smith FN, Barry FP, Murphy JM, et al. An in vivo model for load-modulated remodeling in the rabbit flexor tendon. *J Orthop Res* 2000 Jan;18(1):116-25.
- (180) Robbins JR, Evanko SP, Vogel KG. Mechanical loading and TGF-beta regulate proteoglycan synthesis in tendon. *Arch Biochem Biophys* 1997 Jun 15;342(2):203-11.
- (181) Lavagnino M, Arnoczky SP. In vitro alterations in cytoskeletal tensional homeostasis control gene expression in tendon cells. *J Orthop Res* 2005 Sep;23(5):1211-8.
- (182) Arnoczky SP, Lavagnino M, Egerbacher M. The mechanobiological aetiopathogenesis of tendinopathy: is it the over-stimulation or the under-stimulation of tendon cells? *Int J Exp Pathol* 2007 Aug;88(4):217-26.
- (183) Arampatzis A, De MG, Karamanidis K, Morey-Klapsing G, Stafilidis S, Bruggemann GP. Influence of the muscle-tendon unit's mechanical and morphological properties on running economy. *J Exp Biol* 2006 Sep;209(Pt 17):3345-57.
- (184) Patterson-Kane JC, Parry DA, Birch HL, Goodship AE, Firth EC. An age-related study of morphology and cross-link composition of collagen fibrils in the digital flexor tendons of young thoroughbred horses. *Connect Tissue Res* 1997;36(3):253-60.

- (185) Frank C, McDonald D, Wilson J, Eyre D, Shrive N. Rabbit medial collateral ligament scar weakness is associated with decreased collagen pyridinoline crosslink density. *J Orthop Res* 1995 Mar;13(2):157-65.
- (186) Ng GY, Oakes BW, Deacon OW, McLean ID, Eyre DR. Long-term study of the biochemistry and biomechanics of anterior cruciate ligament-patellar tendon autografts in goats. *J Orthop Res* 1996 Nov;14(6):851-6.
- (187) Oxlund H, Barckman M, Ortoft G, Andreassen TT. Reduced concentrations of collagen cross-links are associated with reduced strength of bone. *Bone* 1995 Oct;17(4 Suppl):365S-71S.
- (188) Craig AS, Birtles MJ, Conway JF, Parry DA. An estimate of the mean length of collagen fibrils in rat tail-tendon as a function of age. *Connect Tissue Res* 1989;19(1):51-62.
- (189) Provenzano PP, Vanderby R, Jr. Collagen fibril morphology and organization: implications for force transmission in ligament and tendon. *Matrix Biol* 2006 Mar;25(2):71-84.
- (190) Parry DA, Barnes GR, Craig AS. A comparison of the size distribution of collagen fibrils in connective tissues as a function of age and a possible relation between fibril size distribution and mechanical properties. *Proc R Soc Lond B Biol Sci* 1978 Dec 18;203(1152):305-21.
- (191) Parry DA. The molecular and fibrillar structure of collagen and its relationship to the mechanical properties of connective tissue. *Biophys Chem* 1988 Feb;29(1-2):195-209.
- (192) PARTINGTON FR, WOOD GC. The role of non-collagen components in the mechanical behaviour of tendon fibres. *Biochim Biophys Acta* 1963 Mar 5;69:485-95.
- (193) Vlassara H. Advanced glycation end-products and atherosclerosis. *Ann Med* 1996 Oct;28(5):419-26.
- (194) Munch G, Schinzel R, Loske C, Wong A, Durany N, Li JJ, et al. Alzheimer's disease--synergistic effects of glucose deficit, oxidative stress and advanced glycation endproducts. *J Neural Transm* 1998;105(4-5):439-61.
- (195) Colaco CA, Ledesma MD, Harrington CR, Avila J. The role of the Maillard reaction in other pathologies: Alzheimer's disease. *Nephrol Dial Transplant* 1996;11 Suppl 5:7-12.
- (196) Vlassara H. Protein glycation in the kidney: role in diabetes and aging. *Kidney Int* 1996 Jun;49(6):1795-804.
- (197) Bai P, Phua K, Hardt T, Cernadas M, Brodsky B. Glycation alters collagen fibril organization. *Connect Tissue Res* 1992;28(1-2):1-12.
- (198) Andreassen TT, Oxlund H, Danielsen CC. The influence of non-enzymatic glycosylation and formation of fluorescent reaction products on the mechanical properties of rat tail tendons. *Connect Tissue Res* 1988;17(1):1-9.

- (199) Andreassen TT, Seyer-Hansen K, Bailey AJ. Thermal stability, mechanical properties and reducible cross-links of rat tail tendon in experimental diabetes. *Biochim Biophys Acta* 1981 Oct 12;677(2):313-7.
- (200) VERZAR F. The aging of collagen. *Sci Am* 1963 Apr;208:104-14.
- (201) Sargon MF, Ozlu K, Oken F. Age-related changes in human tendo calcaneus collagen fibrils. *Saudi Med J* 2005 Mar;26(3):425-8.
- (202) Nakagawa Y, Majima T, Nagashima K. Effect of ageing on ultrastructure of slow and fast skeletal muscle tendon in rabbit Achilles tendons. *Acta Physiol Scand* 1994 Nov;152(3):307-13.
- (203) Crouse JR, Grundy SM, Ahrens EH, Jr. Cholesterol distribution in the bulk tissues of man: variation with age. *J Clin Invest* 1972 May;51(5):1292-6.
- (204) Adams CW, Bayliss OB. Acid mucosubstances underlying lipid deposits in ageing tendons and atherosclerotic arteries. *Atherosclerosis* 1973 Sep;18(2):191-5.
- (205) Adams CW, Bayliss OB, Baker RW, Abdulla YH, Hunter-Craig CJ. Lipid deposits in ageing human arteries, tendons and fascia. *Atherosclerosis* 1974 May;19(3):429-40.
- (206) Deschenes MR, Giles JA, McCoy RW, Volek JS, Gomez AL, Kraemer WJ. Neural factors account for strength decrements observed after short-term muscle unloading. *Am J Physiol Regul Integr Comp Physiol* 2002 Feb;282(2):R578-R583.
- (207) Lundbye-Jensen J, Nielsen JB. Immobilization induces changes in presynaptic control of group Ia afferents in healthy humans. *J Physiol* 2008 Sep 1;586(Pt 17):4121-35.
- (208) Lundbye-Jensen J, Nielsen JB. Central nervous adaptations following 1 wk of wrist and hand immobilization. *J Appl Physiol* 2008 Jul;105(1):139-51.
- (209) Bamman MM, Clarke MS, Feeback DL, Talmadge RJ, Stevens BR, Lieberman SA, et al. Impact of resistance exercise during bed rest on skeletal muscle sarcopenia and myosin isoform distribution. *J Appl Physiol* 1998 Jan;84(1):157-63.
- (210) Majima T, Yasuda K, Fujii T, Yamamoto N, Hayashi K, Kaneda K. Biomechanical effects of stress shielding of the rabbit patellar tendon depend on the degree of stress reduction. *J Orthop Res* 1996 May;14(3):377-83.
- (211) Lavagnino M, Arnoczky SP, Tian T, Vaupel Z. Effect of amplitude and frequency of cyclic tensile strain on the inhibition of MMP-1 mRNA expression in tendon cells: an in vitro study. *Connect Tissue Res* 2003;44(3-4):181-7.
- (212) Kawabata H, Katsura T, Kondo E, Kitamura N, Miyatake S, Tanabe Y, et al. Stress deprivation from the patellar tendon induces apoptosis of fibroblasts in vivo with activation of mitogen-activated protein kinases. *J Biomech* 2009 Nov 13;42(15):2611-5.
- (213) Cook SB, Clark BC, Ploutz-Snyder LL. Accelerometry as a measure of subject compliance in unilateral lower limb suspension. *Aviat Space Environ Med* 2006 Sep;77(9):953-6.

- (214) Clark BC, Fernhall B, Ploutz-Snyder LL. Adaptations in human neuromuscular function following prolonged unweighting: I. Skeletal muscle contractile properties and applied ischemia efficacy. *J Appl Physiol* 2006 Jul;101(1):256-63.

**C. Couppé, M. Kongsgaard, P. Aagaard, P. Hansen, J. Bojsen-Moller, M. Kjaer and S. P. Magnusson**

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C. Couppe, P. Hansen, M. Kongsgaard, V. Kovanen, C. Suetta, P. Aagaard, M. Kjaer and S. P. Magnusson

*J Appl Physiol*, September 1, 2009; 107 (3): 880-886.

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**Training-induced changes in structural and mechanical properties of the patellar tendon are related to muscle hypertrophy but not to strength gains**

O. R. Seynnes, R. M. Erskine, C. N. Maganaris, S. Longo, E. M. Simoneau, J. F. Grosset and M. V. Narici

*J Appl Physiol*, August 1, 2009; 107 (2): 523-530.

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## Habitual loading results in tendon hypertrophy and increased stiffness of the human patellar tendon

C. Couppe, M. Kongsgaard, P. Aagaard, P. Hansen, J. Bojsen-Moller, M. Kjaer, and S. P. Magnusson

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**Couppé C, Kongsgaard M, Aagaard P, Hansen P, Bojsen-Moller J, Kjaer M, Magnusson SP.** Habitual loading results in tendon hypertrophy and increased stiffness of the human patellar tendon. *J Appl Physiol* 105: 805–810, 2008. First published June 12, 2008; doi:10.1152/jappphysiol.90361.2008.—The purpose of this study was to examine patellar tendon (PT) size and mechanical properties in subjects with a side-to-side strength difference of  $\geq 15\%$  due to sport-induced loading. Seven elite fencers and badminton players were included. Cross-sectional area (CSA) of the PT obtained from MRI and ultrasonography-based measurement of tibial and patellar movement together with PT force during isometric contractions were used to estimate mechanical properties of the PT bilaterally. We found that distal tendon and PT, but not mid-tendon, CSA were greater on the lead extremity compared with the nonlead extremity (distal:  $139 \pm 11$  vs.  $116 \pm 7$  mm<sup>2</sup>; mid-tendon:  $85 \pm 5$  vs.  $77 \pm 3$  mm<sup>2</sup>; proximal:  $106 \pm 7$  vs.  $83 \pm 4$  mm<sup>2</sup>;  $P < 0.05$ ). Distal tendon CSA was greater than proximal and mid-tendon CSA on both the lead and nonlead extremity ( $P < 0.05$ ). For a given common force, stress was lower on the lead extremity ( $52.9 \pm 4.8$  MPa) compared with the nonlead extremity ( $66.0 \pm 8.0$  MPa;  $P < 0.05$ ). PT stiffness was also higher in the lead extremity ( $4,766 \pm 716$  N/mm) compared with the nonlead extremity ( $3,494 \pm 446$  N/mm) ( $P < 0.05$ ), whereas the modulus did not differ (lead  $2.27 \pm 0.27$  GPa vs. nonlead  $2.16 \pm 0.28$  GPa) at a common force. These data show that a habitual loading is associated with a significant increase in PT size and mechanical properties.

unilateral; strength; tendon size

HUMAN MOVEMENT COMES ABOUT from the force created by contracting muscles, which is transmitted to bone via aponeurosis and tendon. Recent data in human models suggest that tendon tissue is quite metabolically responsive to tensile loading (4, 12, 16). In fact, it has been shown that both a single loading bout as well as long-term habitual loading produce a markedly elevated collagen synthesis response (20, 21, 26). However, to what extent this elevated synthesis yields incorporation of collagen into the load-bearing structure of tendon, and therefore either an increase in tendon size (hypertrophy) or an altered composition and a change in mechanical function, remains ambiguous.

Studies that address the influence of exercise on the mechanical properties of tendon have until recently largely been conducted in animal models. These data show that endurance-like exercise is associated with an increase (1, 35, 39), decrease (39), or unchanged (5, 36) tendon size, and thus they do not provide a coherent picture. In humans, cross-sectional data suggest that endurance training is associated with a larger

Achilles tendon cross-sectional area (CSA) (17, 24, 33), which appears to be site specific (17, 24). However, in a recent intervention study it was shown that 9 mo of endurance training in untrained persons left the Achilles tendon CSA unchanged (13). On the other hand, animal data have shown that muscle size is related to the tendon size (8), suggesting that perhaps the magnitude of loading influences tendon size. Several human studies have shown that resistance training over 12–14 wk that produces increases in muscle strength of up to 21% does not result in an accompanying increase in tendon CSA (19, 30, 31), but rather a markedly altered modulus, which implies that there is a change in the composition of the structure rather than the size. However, it was recently shown in humans that resistance training for 12 wk yielded region-specific increases in patellar tendon (PT) CSA, without a change in modulus (18).

Conclusions from studies of cross-sectional design are inevitably hampered by issues of training history, selection bias, and intersubject variations. Additionally, longitudinal training studies may have been of insufficient duration to produce a robust tendon hypertrophy response. Finally, existing training studies (19, 30, 31) have examined tendon size in a region that appears unresponsive to training-associated adaptation. However, some of these limitations may be partially circumscribed by examination of region-specific PT properties in persons who engage in sport where one lower extremity is habitually subjected to more loading than the contralateral (“control”) side, such as in fencing or badminton. Therefore, the purpose of the present study was to examine region-specific PT size and mechanical properties in subjects who display a side-to-side strength difference of  $\geq 15\%$  due to persistent sport-induced loading over several years.

### MATERIALS AND METHODS

**Subjects.** Both badminton and fencing involve repeatedly performing rapid forward lunges with a preferred and thus more loaded lead extremity compared with the nonlead extremity, which in this study served as a within-subject control. Sapega et al. (34) and Nystrom et al. (28) found a  $>14\%$  strength difference between the legs in elite fencers, reflecting a sport-specific loading. The sport-specific loading in badminton and fencing creates a higher impact on the leg and therefore also on the PT compared with other sports without jumping and unilateral rapid lunges (27). A total of 22 athletes volunteered to participate in the study. They were recruited from the Danish National Fencing Center and the International Badminton Academy in Copenhagen, Denmark. Seven elite fencers ( $n = 4$ ) and badminton ( $n = 3$ ) players met the criteria of a side-to-side isometric knee extensor

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strength difference of  $\geq 15\%$  ( $23 \pm 2$  yr,  $74 \pm 3$  kg,  $184 \pm 3$  cm). They had participated in their respective sport for  $>5$  yr at an elite level. In addition, seven recreational athletes ( $24 \pm 1$  yr,  $77 \pm 4$  kg,  $182 \pm 2$  cm) who did not engage in fencing or badminton were examined with respect to thigh strength and tendon morphology, but not mechanical properties, to ensure that there was no side-to-side difference between the dominant and nondominant side as determined by preferred kicking leg. All were healthy and without knee pathology. The study complied with the Declaration of Helsinki and was approved by the local ethics committee. All subjects gave their informed consent before the experiment.

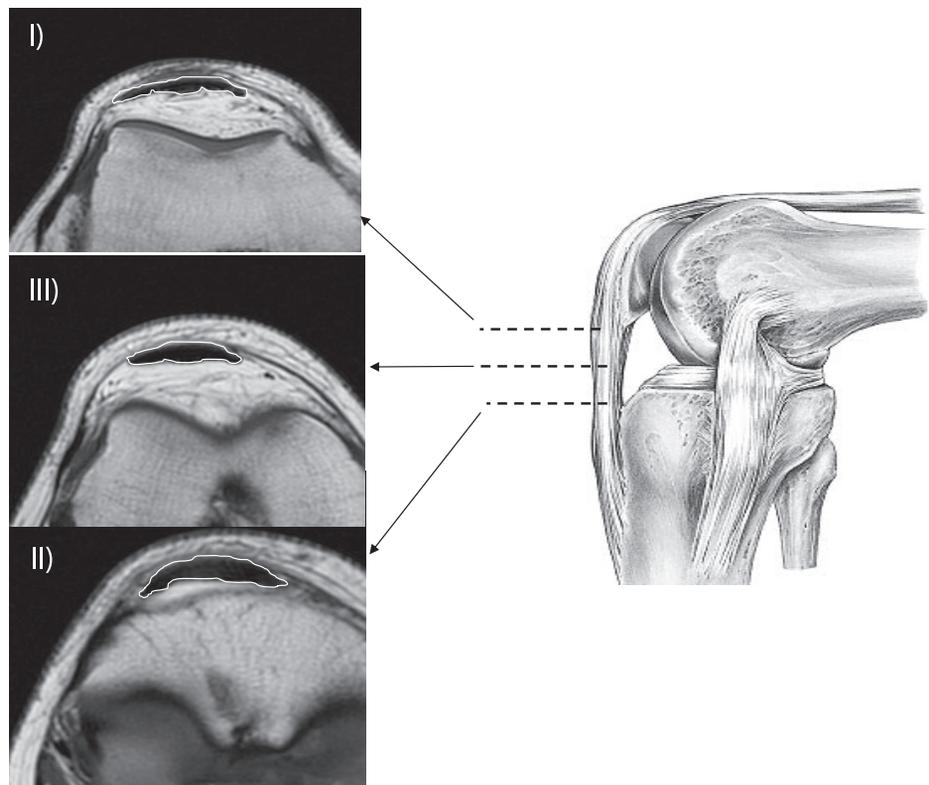
**Maximal voluntary contraction.** Maximal voluntary strength was determined by an isometric knee extension. Subjects performed a 10-min warm-up on a stationary bike (5 min at 150 W and 5 min at 200 W) before testing. The subjects were seated in a custom-made rigid chair with both hips and knees flexed to an angle of  $90^\circ$ . A leg cuff was connected to a strain gauge (model KRG-4, Bofors, Bofors, Sweden) through a rigid steel rod perpendicular to the lower leg and was mounted on the leg just above the medial malleolus. The tibia moment arm was measured (from the point of fixation to the lateral epicondyle of the knee) to calculate the knee extensor moment. Force was sampled on a personal computer (PC) at 50 Hz via a 12-bit analog-to-digital converter (model dt 2810A, Data Translation, Marlboro, MA). The subjects performed four to five maximal voluntary isometric knee extension contractions (MVC) separated by 1 min (sampled at 1,000 Hz) (3). The tests were conducted on both legs. A side-to-side difference in strength of  $\geq 15\%$  was defined as follows:  $(\text{lead extremity} - \text{nonlead extremity}) / \text{nonlead extremity} \cdot 100 \geq 15\%$ .

**Muscle and tendon morphological measurements.** The anatomic CSA of the quadriceps femoris muscle was measured 20 cm proximal from the tibia plateau (mid-thigh level) by magnetic resonance imaging (MRI) [Signa Horizon LX 1.5 T, General Electric, longitudinal relaxation time (T1)-weighted spin echo (SE)] using a lower extremity coil. The images were obtained using the following parameters: transverse relaxation time (TR)/echo time (TE) = 500/14 ms, field of view (FOV) 18, matrix  $512 \times 512$ , and slice thickness 6 mm (18).

Subsequently, the lean muscle mass of the quadriceps muscle (subcutaneous and intermuscular noncontractile tissue were not included in the measurement) was manually outlined using the software program Osiris 4.19 (<http://www.sim.hcuge.ch/osiris/>). The mean value of three measurements of the same image was used for analysis. PT CSA and PT length were determined with the use of MRI and the lower extremity coil (Signa Horizon LX 1.5 T, General Electric, T1-weighted SE) (18). PT CSA was determined by axial plane magnetic resonance using the following parameters: TR/TE 400/14 ms, FOV 20, matrix  $256 \times 256$ , slice thickness 5.0 mm, and spacing 0 mm. The axial scans were performed perpendicular to the PT. The tendon CSA was measured 1) just distal to the patellar insertion, 2) just proximal to the tibia insertion, and 3) midway between these two sites (Fig. 1). The PT length was determined from sagittal plane MRI using the following parameters: TR 500, echo train  $3 \times$  (TE 12.4 ms), FOV 16, matrix  $256 \times 192$ , slice thickness 4.0 mm, and no spacing. The PT length was obtained by measuring the distance from the dorsal insertion at the patella apex to the dorsal insertion on the tibia. The PT CSA and PT length were manually outlined using the software program Osiris 4.19 (<http://www.sim.hcuge.ch/osiris/>). The color intensity of each image was adjusted using the National Institutes of Health color scale mode of the software. Tendon CSA and length were measured using the gray-scale image display. The mean value of three measurements of the same image was used for analysis. The reproducibility data showed that the typical error percentage of repeated measures of tendon CSA was 2.5% for repeated measurements at the proximal tendon level, 2.5% for repeated measurements at the mid-tendon level, and 2.0% for repeated measures at the most distal tendon level. The MRI investigator was blinded with regards to the subjects and side.

**Assessment of PT mechanical properties.** The details of the measurement, including the reliability of the method in our laboratory, has been reported previously (14). The within-day correlation coefficients and typical error percent of repeated measures were 0.95 and 9.9% for stiffness, 0.97 and 5.5% for strain, and 0.94 and 9.4% for modulus. Subjects performed a 5-min warm-up on a stationary bike to secure

Fig. 1. Axial scans were performed perpendicular to the patellar tendon. Tendon cross-sectional area (CSA) was measured just distal to the patellar insertion (I), just proximal to the tibia insertion (II), and midway between these 2 sites (III).



proper preconditioning of the tendon before testing. Thereafter, the subjects were seated in a custom made rigid chair with both hips and knees flexed to an angle of 90°. A leg cuff, which was connected to a strain gauge (model KRG-4, Bofors) through a rigid steel rod perpendicular to the lower leg, was mounted on the leg just above the medial malleolus. An ultrasound probe (7.5 MHz, linear array B-mode, Sonoline Sienna, Siemens, Erlangen, Germany) was fitted into a custom-made rigid cast that was secured to the skin above the PT in the sagittal plane. The ultrasound probe and cast was positioned so that the patella, the PT and the tibia were all visible within the viewing field throughout the ramped contractions (Fig. 3).

The ultrasound S-VHS video images obtained during the ramp trials were sampled at 50 Hz on a PC using frame-by-frame capturing software (model G400-TV, Matrox Marvel, Dorval, Canada). Force was sampled on two separate PCs at 50 Hz via a 12-bit analog-to-digital converter (model dt 2810A, Data Translation). The two computers were interconnected to permit synchronous sampling of all data using a custom-built trigger device (3). The subjects performed four to five slow isometric ramp contractions by applying gradually increasing force on the cuff over a 10-s period while PT displacement and knee extension force were measured simultaneously. Each ramp was separated by a 2-min rest. All measurements were performed on both legs. During the ramp contractions, force was sampled at 50 Hz and filtered at a 1.0-Hz cutoff frequency using a fourth-order zero-log Butterworth filter.

PT force was calculated by dividing the estimated total knee extension moment by the internal moment arm, which was estimated from individually measured femur lengths (37). PT stress was calculated by dividing tendon force by the proximal, mid-, and distal tendon CSA determined from the MRI. PT deformation was defined as the change in distance between the patellar apex and the tibia (14, 23). Tendon strain was calculated as the change in length related to the original length. Each force-deformation curve was fitted to a second- or third-order polynomial fit, which yielded  $R^2 > 0.97$ . Tendon stiffness ( $\Delta\text{force}/\Delta\text{deformation}$ ) and modulus ( $\Delta\text{stress}/\Delta\text{strain}$ ) were calculated in the final 10% of the force-deformation and stress-strain curves, respectively (22, 23).

**Statistical analysis.** The two isometric ramp contractions that yielded the greatest force were used for further analysis. To make side-to-side comparisons and thereby account for differences in isometric ramp contractions from side to side, the trials for both sides were subsequently analyzed to the lowest common force for each individual subject. Wilcoxon matched-pairs signed-ranks tests were used to examine whether there was a side-to-side difference in measured variables. Friedman's analysis of variance, including the Dunn's multiple comparison test, was used to detect possible tendon region-specific differences along its length. An alpha level of  $P < 0.05$  was considered significant. Results are reported as means  $\pm$  SE.

## RESULTS

**Quadriceps strength and CSA.** In the recreational athletes, there was no side-to-side difference in MVC (dominant  $217 \pm 14$  N·m, nondominant  $203 \pm 16$  N·m) ( $P > 0.05$ ), or quadriceps CSA (dominant  $7,159 \pm 323$  mm<sup>2</sup>, nondominant  $6,951 \pm 261$  mm<sup>2</sup>) ( $P > 0.05$ ). In the elite athletes, MVC was significantly greater in the lead extremity ( $239 \pm 26$  N·m) compared with the nonlead extremity ( $197 \pm 23$  N·m) ( $P < 0.01$ ). Similarly quadriceps femoris CSA was significantly greater in the lead extremity ( $7,907 \pm 656$  mm<sup>2</sup>) compared with the nonlead extremity ( $7,410 \pm 561$  mm<sup>2</sup>) ( $P < 0.05$ ).

**PT CSA.** In the recreational athletes there was no side-to-side differences in PT CSA for the distal (dominant  $124 \pm 7$  mm<sup>2</sup>, nondominant  $120 \pm 10$  mm<sup>2</sup>), mid- (dominant  $75 \pm 8$  mm<sup>2</sup>, nondominant  $74 \pm 9$  mm<sup>2</sup>), or proximal (dominant  $84 \pm 8$  mm<sup>2</sup>, nondominant  $89 \pm 7$  mm<sup>2</sup>) tendon. The CSA of the

tendon for the elite athletes are shown in Fig. 2. There was a significant side-to-side difference in CSA at the distal (lead  $139 \pm 11$  mm<sup>2</sup>, nonlead  $116 \pm 7$  mm<sup>2</sup>) and proximal tendon (lead  $106 \pm 7$  mm<sup>2</sup>, nonlead  $83 \pm 4$  mm<sup>2</sup>) ( $P < 0.05$ ) but not at the mid-tendon section (lead  $85 \pm 5$  mm<sup>2</sup>, nonlead  $77 \pm 3$  mm<sup>2</sup>) ( $P = 0.218$ ). The PT CSA was greater at the distal tendon compared with mid- and proximal tendon on both the lead extremity and nonlead extremity ( $P < 0.05$ ).

**PT mechanical properties.** Mechanical properties determined at maximal force are shown in Table 1. Tendon force and stiffness were higher on the lead extremity compared with the nonlead extremity ( $P < 0.05$ ). In addition, maximal stress was lower on the lead extremity compared with the nonlead extremity at the proximal tendon level ( $P < 0.05$ ). There was no side-to-side difference for deformation, strain, or Young's modulus based on proximal, mid-, or distal tendon CSA. Stress and modulus were lower at the distal tendon compared with mid- and proximal tendon ( $P < 0.01$ ) on both sides.

Mechanical properties at a common force are shown in Table 2. Tendon stiffness was higher on the lead extremity compared with the nonlead extremity ( $P < 0.05$ ) (Fig. 3). Stress was lower on the lead extremity compared with the nonlead extremity at the proximal (Fig. 4) and distal tendon level ( $P < 0.05$ ). There was no side-to-side difference for deformation, strain, or Young's modulus based on proximal, mid-, or distal tendon CSA. Tendon stress and modulus were lower at the distal level compared with mid- and proximal tendon on both sides ( $P < 0.01$ ). There was no difference in modulus when an average of the three levels of CSA was used (lead extremity  $2.31 \pm 0.29$  GPa, nonlead extremity  $1.99 \pm 0.23$  GPa).

## DISCUSSION

The present study examined PT size and mechanical properties in subjects who had a side-to-side difference in knee extensor strength as a result of habitual sport-specific loading. The main findings were that the lead extremity, which was on average 22% stronger than the nonlead extremity, had a greater distal and proximal PT CSA, which was not evident at the mid-tendon tendon level, indicating a region-specific tendon

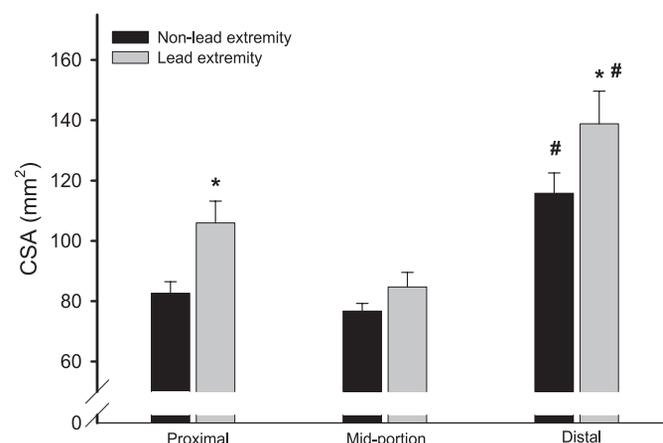


Fig. 2. Patellar tendon CSA distribution. Values are means  $\pm$  SE. \*There was a significant side-to-side difference in CSA at the distal and proximal level but not at the mid-tendon section,  $P < 0.05$ . #CSA was greater at the distal level compared with mid- and proximal tendon on both the lead extremity and nonlead extremity,  $P < 0.05$ .

Table 1. *Patella tendon mechanical properties based on maximum force*

	Lead Extremity	Nonlead Extremity
Force, N	6,886±833*	5,832±631
Deformation, mm	2.8±0.3	3.1±0.4
Stiffness, N/mm	6,011±906*	4,436±570
Stress, MPa		
Proximal tendon	62.1±5.1*	73.0±8.4
Mid-tendon	77.0±6.4	77.1±8.7
Distal tendon	47.9±4.8†	50.8±3.9†
Strain, %	5.5±0.5	6.1±0.6
Modulus, GPa		
Proximal tendon	2.88±0.35	2.76±0.33
Mid-tendon	3.65±0.53	2.92±0.34
Distal tendon	2.22±0.35†	1.98±0.28†

Values are means ± SE. \*Significantly different from nonlead extremity,  $P < 0.05$ . †Significantly different from mid- and proximal tendon,  $P < 0.01$ .

hypertrophy. Furthermore, the lead extremity displayed greater tendon stiffness without a significant difference in modulus, suggesting that the change in mechanical properties was largely the result of a change in size.

The present data show a tendon hypertrophy response associated with chronic loading as evidenced by the greater tendon CSA (20–28%) on the lead extremity. Although tendon tissue until recently has been considered basically inert, human in vivo data persuasively demonstrate that human tendon tissue is very responsive to single bouts of loading and chronic mechanical loading (4, 20, 21, 26). However, whether the elevated synthesis generates an increase in tendon size (hypertrophy) or an altered composition and a change in mechanical function remains largely unknown. Data from animal models show increased (1, 35, 39), decreased (39), or unchanged (5, 36) tendon size in response to endurance exercise. In humans, cross-sectional data show that endurance-trained persons have a larger Achilles tendon CSA compared with that of untrained persons (17, 24, 33). On the other hand, it has been shown that 9 mo of endurance training did not produce any measurable increase in the Achilles tendon CSA or mechanical properties (13). It is possible that duration of the endurance training may explain this apparent discrepancy.

Animal studies have shown a positive relationship between muscle strength/size and tendon CSA (8), which suggests that

Table 2. *Patella tendon mechanical properties based on common force*

	Lead Extremity	Nonlead Extremity
Deformation, mm	2.5±0.3	3.0±0.4
Stiffness, N/mm	4,766±716*	3,494±446
Stress, MPa		
Proximal tendon	52.9±4.8†	66.0±8.0
Mid-tendon	65.6±5.6	71.2±8.2
Distal tendon	40.9±4.4*‡	46.6±4.4‡
Strain, %	5.0±0.5	5.9±0.6
Modulus, GPa		
Proximal tendon	2.27±0.27	2.16±0.28
Mid-tendon	2.87±0.39	2.26±0.25
Distal tendon	1.79±0.25‡	1.54±0.19‡

Values are means ± SE. \*Significantly different from nonlead extremity,  $P < 0.05$ . †Significantly different from nonlead extremity,  $P < 0.01$ . ‡Significantly different from mid- and proximal tendon,  $P < 0.01$ .

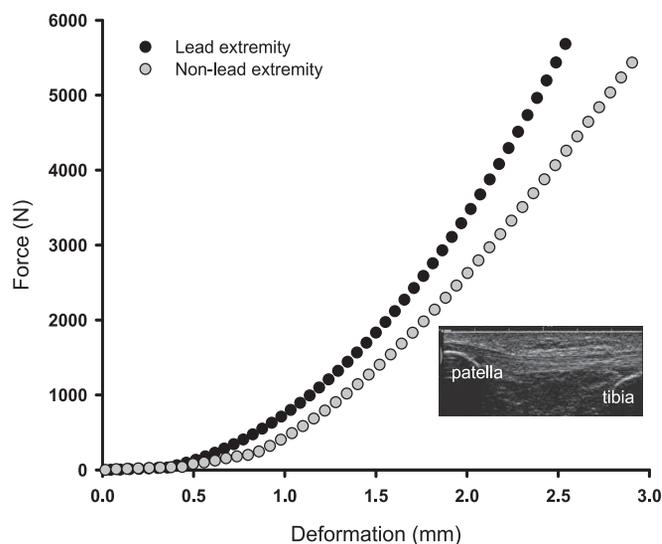


Fig. 3. Patellar tendon force-deformation relationship to a common force. Values are means of all subjects. Stiffness was higher on the lead extremity compared with the nonlead extremity ( $P < 0.05$ ).

the magnitude of loading may also be an important variable with respect to tendon response and adaptation. It was recently shown (30, 31) that 14 wk of resistance training in elderly individuals that produced increases in muscle strength of up to 21% did not result in an increase in tendon CSA measured at the mid-tendon level. In contrast, a recent investigation showed that young men who engaged in resistance training for 12 wk that produced a 15% increase in strength and a 6% increase in muscle CSA had a significant increase (6%) in PT CSA. It is important to note that this tendon hypertrophy was observed in the distal and proximal portion but not in the middle of the tendon (18). Thus possible factors for the inconsistency between these studies include the age of the subjects and the location and method of tendon CSA measurement. The subjects of the present study were elite badminton players and

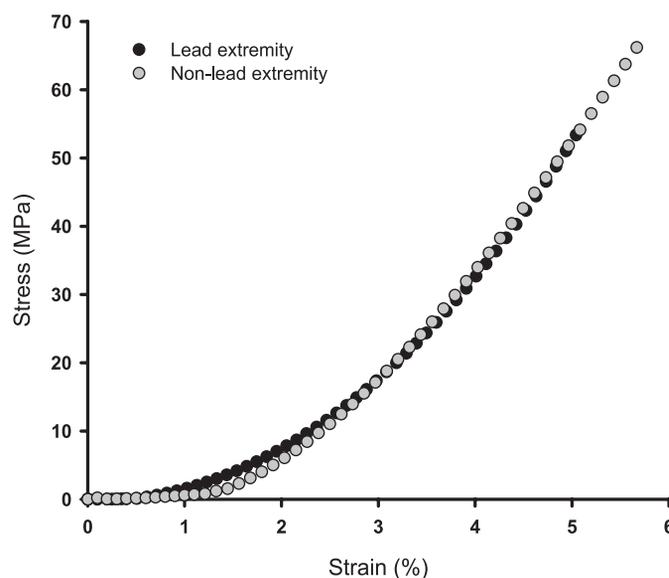


Fig. 4. Patellar tendon stress-strain relationship based on common force. Values are means of all subjects. Stress was lower on the lead extremity compared with the nonlead extremity at the proximal tendon level ( $P < 0.05$ ).

fencers who had participated in their respective sport for several years. These sports involve a sport-specific loading whereby the athletes frequently perform rapid forward lunges with the preferred lead extremity, which places a considerable eccentric load on the knee extensors. In an elite badminton match, >70 such forward lunges may be performed on the lead extremity (C. Couppe, unpublished data). The sport-specific loading was substantiated by the sizeable side-to-side difference in strength (22%) and quadriceps CSA (7%), which was accompanied by a markedly greater tendon CSA (20–28%) of the lead extremity. The magnitude of the side-specific PT CSA (20–28%), and the lack thereof in recreational athletes, strengthens the notion that mechanical loading is associated with tendon hypertrophy as observed by Kongsgaard et al. (18). The magnitude of the CSA in the present study was similar to previous cross-sectional observations, suggesting that perhaps months to years of loading is required for tendon hypertrophy, because only a smaller magnitude of tendon hypertrophy was observed after 12 wk of strength training in the Kongsgaard et al. study.

The data of the present study show that the PT on the lead extremity had a greater stiffness (36%) compared with the nonlead extremity (see Tables 1 and 2), whereas there was no significant difference in modulus. The load-associated increase in stiffness, and lack thereof in modulus, is in accordance with that recently found in young men following resistance training (18). However, others have demonstrated a marked increase in stiffness and in modulus after strength training in elderly individuals (30, 31). Although these diverging findings are difficult to reconcile, they may partly be related to age and the method of determining mechanical properties (14). The present findings and those of Kongsgaard et al. (18) suggest that the change in mechanical properties in young men is largely a function of increased in size, rather than a material change, although this needs to be confirmed.

The present data show that the lead extremity had a greater PT CSA in the distal (20%) and proximal region (28%) but not in the midsection of the tendon. Furthermore, there were no PT CSA side-to-side differences in recreational athletes. The magnitude of this region-specific human tendon hypertrophy confirms and reinforces previous cross-sectional data (17, 24, 33) and more recent training data (18). Albeit speculative, perhaps compression in these regions contributes to the synthesis of extracellular matrix proteins in tendinous tissue (25, 32). The midsection of the tendon had the narrowest CSA and thus a greater average stress (~25%) for a given load. Yet, in contrast to the proximal and distal tendon, the middle part of the tendon did not significantly increase in CSA as a result of the sport-specific loading. Interestingly, many PT clinical conditions occur at the insertions (9, 10) where the tendon CSA is larger and where it appears most likely to hypertrophy, and therefore it is doubtful whether whole tendon region specific average stress (and “safety factor”) per se is the most important variable with respect to injury and adaptation. In contrast, the mid-tendon, which is rarely injured, did not appear to display a pronounced hypertrophy response, and it is therefore likely well designed for the imposed loads.

The knee extensor strength and tendon force of the nonlead extremity in the elite athletes in the present study are comparable to that of the recreational athletes examined herein and that reported by others (7, 18, 19). The PT deformation and

strain in the present study (~3.1 mm, 5–6%) are also similar to previous reports (18, 38), but markedly different from that reported by others (~4.3 mm, ~9%) (7), which is likely related to dissimilarities in measurement methodology (14). In the present study, the average stress of the PT was 53–73 MPa. However, similar to previous reports (7, 29), the data show that the calculation of tendon stress will largely depend on where along the length of the tendon CSA is obtained. Nevertheless, the considerable variation in PT stress (30–91 MPa) reported in earlier studies (2, 14, 18, 29, 38) in the presence of reasonably comparable tendon forces (5,600–7,000 N) is most likely a function of both measurement methodology (ultrasonography and MRI) and analysis method when MRI is used. The modulus values reported herein are comparable to previous animal data on isolated tendon (6, 11, 15), but they are noticeably higher than previously reported human in vivo data (2, 18, 38). This discrepancy can most likely also be explained by the combination of differences in measuring deformation and CSA. It is noteworthy that the modulus at the proximal insertions is strikingly similar between the lead and nonlead extremity (Tables 1 and 2), which suggests that the sport-specific loading has not altered the material properties of the tendon, although this needs to be confirmed with other investigative techniques.

There are inherent limitations associated with the present study. The sample size was limited, which was in part a function of the study design and its inclusion criteria. Furthermore, the ultrasonography-based measurement technique does not permit assessment of region-specific deformation. This means that, although average stress can be calculated for specific location along the tendon, based on CSA, which varies considerably, modulus can only be estimated assuming heterogenous deformation and strain along the tendon, which remains unknown in humans, in vivo. Estimating regional modulus for the tendon yielded similar values for the proximal and distal tendon, although the values for mid-tendon approached significance ( $P = 0.11$ ), and they may have attained significance with a larger sample. Furthermore, albeit not possible with this particular group of athletes, a study design, including tendon biopsies, may have provided some more insight into the composition of the tendons.

In summary, the present study examined region-specific PT size and mechanical properties in subjects who display a side-to-side strength difference of  $\geq 15\%$  due to persistent sport-induced loading over several years. These subjects were examined in an attempt to partly circumscribe issues such as 1) training history, selection bias, and intersubject variations in cross-sectionally designed studies; 2) the relatively short duration of earlier training studies; and 3) the lack of assessment of region specificity in existing training studies. The data showed a regional variation in CSA along the PT, which markedly influenced average tendon stress. The habitually more loaded PT of the stronger extremity had a greater CSA compared with the contralateral side. The lead extremity also displayed greater stiffness than the contralateral side, whereas the modulus did not differ significantly. In sum, these data show that a habitual loading is associated with a robust change of the PT size and mechanical properties.

## GRANTS

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## REFERENCES

- Birch HL, McLaughlin L, Smith RK, Goodship AE. Treadmill exercise-induced tendon hypertrophy: assessment of tendons with different mechanical functions. *Equine Vet J Suppl* 30: 222–226, 1999.
- Bojsen-Moller J, Brogaard K, Have MJ, Stryger HP, Kjaer M, Aagaard P, Magnusson SP. Passive knee joint range of motion is unrelated to the mechanical properties of the patellar tendon. *Scand J Med Sci Sports* 17: 415–421, 2007.
- Bojsen-Moller J, Hansen P, Aagaard P, Kjaer M, Magnusson SP. Measuring mechanical properties of the vastus lateralis tendon-aponeurosis complex in vivo by ultrasound imaging. *Scand J Med Sci Sports* 13: 259–265, 2003.
- Bojsen-Moller J, Kalliokoski KK, Seppanen M, Kjaer M, Magnusson SP. Low-intensity tensile loading increases intratendinous glucose uptake in the Achilles tendon. *J Appl Physiol* 101: 196–201, 2006.
- Buchanan CI, Marsh RL. Effects of long-term exercise on the biomechanical properties of the Achilles tendon of guinea fowl. *J Appl Physiol* 90: 164–171, 2001.
- Butler DL, Kay MD, Stouffer DC. Comparison of material properties in fascicle-bone units from human patellar tendon and knee ligaments. *J Biomech* 19: 425–432, 1986.
- de Boer MD, Maganaris CN, Seynnes OR, Rennie MJ, Narici MV. Time course of muscular, neural and tendinous adaptations to 23 day unilateral lower-limb suspension in young men. *J Physiol* 583: 1079–1091, 2007.
- Elliott DH. Structure and function of mammalian tendon. *Biol Rev Camb Philos Soc* 40: 392–421, 1965.
- Ferretti A. Epidemiology of jumper's knee. *Sports Med* 3: 289–295, 1986.
- Ferretti A, Ippolito E, Mariani P, Puddu G. Jumper's knee. *Am J Sports Med* 11: 58–62, 1983.
- Flahiff CM, Brooks AT, Hollis JM, Vander Schilden JL, Nicholas RW. Biomechanical analysis of patellar tendon allografts as a function of donor age. *Am J Sports Med* 23: 354–358, 1995.
- Hannukainen J, Kalliokoski KK, Nuutila P, Fujimoto T, Kempainen J, Viljanen T, Laaksonen MS, Parkkola R, Knuuti J, Kjaer M. In vivo measurements of glucose uptake in human Achilles tendon during different exercise intensities. *Int J Sports Med* 26: 727–731, 2005.
- Hansen P, Aagaard P, Kjaer M, Larsson B, Magnusson SP. Effect of habitual running on human Achilles tendon load-deformation properties and cross-sectional area. *J Appl Physiol* 95: 2375–2380, 2003.
- Hansen P, Bojsen-Moller J, Aagaard P, Kjaer M, Magnusson SP. Mechanical properties of the human patellar tendon, in vivo. *Clin Biomech (Bristol, Avon)* 21: 54–58, 2005.
- Johnson GA, Tramaglino DM, Levine RE, Ohno K, Choi NY, Woo SL. Tensile and viscoelastic properties of human patellar tendon. *J Orthop Res* 12: 796–803, 1994.
- Kalliokoski KK, Langberg H, Ryberg AK, Scheede-Bergdahl C, Doessing S, Kjaer A, Boushel R, Kjaer M. The effect of dynamic knee-extension exercise on patellar tendon and quadriceps femoris muscle glucose uptake in humans studied by positron emission tomography. *J Appl Physiol* 99: 1189–1192, 2005.
- Kongsgaard M, Aagaard P, Kjaer M, Magnusson SP. Structural Achilles tendon properties in athletes subjected to different exercise modes and in Achilles tendon rupture patients. *J Appl Physiol* 99: 1965–1971, 2005.
- Kongsgaard M, Reitelsheder S, Pedersen TG, Holm L, Aagaard P, Kjaer M, Magnusson SP. Region specific patellar tendon hypertrophy in humans following resistance training. *Acta Physiol (Oxf)* 191: 111–121, 2007.
- Kubo K, Yata H, Kanehisa H, Fukunaga T. Effects of isometric squat training on the tendon stiffness and jump performance. *Eur J Appl Physiol* 96: 305–314, 2006.
- Langberg H, Rosendal L, Kjaer M. Training-induced changes in peritendinous type I collagen turnover determined by microdialysis in humans. *J Physiol* 534: 297–302, 2001.
- Langberg H, Skovgaard D, Petersen LJ, Bulow J, Kjaer M. Type I collagen synthesis and degradation in peritendinous tissue after exercise determined by microdialysis in humans. *J Physiol* 521: 299–306, 1999.
- Magnusson SP, Aagaard P, Dyhre-Poulsen P, Kjaer M. Load-displacement properties of the human triceps surae aponeurosis in vivo. *J Physiol* 531: 277–288, 2001.
- Magnusson SP, Hansen P, Aagaard P, Brond J, Dyhre-Poulsen P, Bojsen-Moller J, Kjaer M. Differential strain patterns of the human gastrocnemius aponeurosis and free tendon, in vivo. *Acta Physiol Scand* 177: 185–195, 2003.
- Magnusson SP, Kjaer M. Region-specific differences in Achilles tendon cross-sectional area in runners and non-runners. *Eur J Appl Physiol* 90: 549–553, 2003.
- Malaviya P, Butler DL, Boivin GP, Smith FN, Barry FP, Murphy JM, Vogel KG. An in vivo model for load-modulated remodeling in the rabbit flexor tendon. *J Orthop Res* 18: 116–125, 2000.
- Miller BF, Olesen JL, Hansen M, Dossing S, Cramer RM, Welling RJ, Langberg H, Flyvbjerg A, Kjaer M, Babraj JA, Smith K, Rennie MJ. Coordinated collagen and muscle protein synthesis in human patella tendon and quadriceps muscle after exercise. *J Physiol* 567: 1021–1033, 2005.
- Nordstrom P, Pettersson U, Lorentzon R. Type of physical activity, muscle strength, and pubertal stage as determinants of bone mineral density and bone area in adolescent boys. *J Bone Miner Res* 13: 1141–1148, 1998.
- Nystrom J, Lindwall O, Ceci R, Harmenberg J, Svedenhag J, Ekblom B. Physiological and morphological characteristics of world class fencers. *Int J Sports Med* 11: 136–139, 1990.
- Onambele GN, Burgess K, Pearson SJ. Gender-specific in vivo measurement of the structural and mechanical properties of the human patellar tendon. *J Orthop Res* 25: 1635–1642, 2007.
- Reeves ND, Maganaris CN, Narici MV. Effect of strength training on human patella tendon mechanical properties of older individuals. *J Physiol* 548: 971–981, 2003.
- Reeves ND, Narici MV, Maganaris CN. Strength training alters the viscoelastic properties of tendons in elderly humans. *Muscle Nerve* 28: 74–81, 2003.
- Robbins JR, Evanko SP, Vogel KG. Mechanical loading and TGF-beta regulate proteoglycan synthesis in tendon. *Arch Biochem Biophys* 342: 203–211, 1997.
- Rosager S, Aagaard P, Dyhre-Poulsen P, Neergaard K, Kjaer M, Magnusson SP. Load-displacement properties of the human triceps surae aponeurosis and tendon in runners and non-runners. *Scand J Med Sci Sports* 12: 90–98, 2002.
- Sapega AA, Minkoff J, Valsamis M, Nicholas JA. Musculoskeletal performance testing and profiling of elite competitive fencers. *Clin Sports Med* 3: 231–244, 1984.
- Sommer HM. The biomechanical and metabolic effects of a running regime on the Achilles tendon in the rat. *Int Orthop* 11: 71–75, 1987.
- Viidik A. Tensile strength properties of Achilles tendon systems in trained and untrained rabbits. *Acta Orthop Scand* 40: 261–272, 1969.
- Visser JJ, Hoogkamer JE, Bobbert MF, Huijijng PA. Length and moment arm of human leg muscles as a function of knee and hip-joint angles. *Eur J Appl Physiol Occup Physiol* 61: 453–460, 1990.
- Westh E, Kongsgaard M, Bojsen-Moller J, Aagaard P, Hansen M, Kjaer M, Magnusson SP. Effect of habitual exercise on the structural and mechanical properties of human tendon, in vivo, in men and women. *Scand J Med Sci Sports* 18: 23–30, 2008.
- Woo SL, Gomez MA, Woo YK, Akeson WH. Mechanical properties of tendons and ligaments. II. The relationships of immobilization and exercise on tissue remodeling. *Biorheology* 19: 397–408, 1982.

**C. Couppé, P. Hansen, M. Kongsgaard, V. Kovanen, C. Suetta, P. Aagaard, M. Kjær and S. P. Magnusson**  
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## Mechanical properties and collagen cross-linking of the patellar tendon in old and young men

C. Couppé,<sup>1,2</sup> P. Hansen,<sup>1</sup> M. Kongsgaard,<sup>1</sup> V. Kovanen,<sup>3</sup> C. Suetta,<sup>1</sup> P. Aagaard,<sup>4</sup>  
M. Kjær,<sup>1</sup> and S. P. Magnusson<sup>1,2</sup>

<sup>1</sup>Institute of Sports Medicine, Bispebjerg Hospital and Center for Healthy Aging, Faculty of Health Sciences, University of Copenhagen, Copenhagen, Denmark; <sup>2</sup>Department of Physical Therapy, Bispebjerg Hospital, Copenhagen, Denmark; <sup>3</sup>Department of Health Sciences, University of Jyväskylä, Jyväskylä, Finland; and <sup>4</sup>The University of Southern Denmark, Odense, Denmark

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**Couppé C, Hansen P, Kongsgaard M, Kovanen V, Suetta C, Aagaard P, Kjær M, Magnusson SP.** Mechanical properties and collagen cross-linking of the patellar tendon in old and young men. *J Appl Physiol* 107: 880–886, 2009. First published June 25, 2009; doi:10.1152/jappphysiol.00291.2009.—Age-related loss in muscle mass and strength impairs daily life function in the elderly. However, it remains unknown whether tendon properties also deteriorate with age. Cross-linking of collagen molecules provides structural integrity to the tendon fibrils and has been shown to change with age in animals but has never been examined in humans in vivo. In this study, we examined the mechanical properties and pyridinoline and pentosidine cross-link and collagen concentrations of the patellar tendon in vivo in old (OM) and young men (YM). Seven OM ( $67 \pm 3$  years,  $86 \pm 10$  kg) and 10 YM ( $27 \pm 2$  years,  $81 \pm 8$  kg) with a similar physical activity level (OM  $5 \pm 6$  h/wk, YM  $5 \pm 2$  h/wk) were examined. MRI was used to assess whole tendon dimensions. Tendon mechanical properties were assessed with the use of simultaneous force and ultrasonographic measurements during ramped isometric contractions. Percutaneous tendon biopsies were taken and analyzed for hydroxylysyl pyridinoline (HP), lysyl pyridinoline (LP), pentosidine, and collagen concentrations. We found no significant differences in the dimensions or mechanical properties of the tendon between OM and YM. Collagen concentrations were lower in OM than in YM ( $0.49 \pm 0.27$  vs.  $0.73 \pm 0.14$  mg/mg dry wt;  $P < 0.05$ ). HP concentrations were higher in OM than in YM ( $898 \pm 172$  vs.  $645 \pm 183$  mmol/mol;  $P < 0.05$ ). LP concentrations were higher in OM than in YM ( $49 \pm 38$  vs.  $16 \pm 8$  mmol/mol;  $P < 0.01$ ), and pentosidine concentrations were higher in OM than in YM ( $73 \pm 13$  vs.  $11 \pm 2$  mmol/mol;  $P < 0.01$ ). These cross-sectional data raise the possibility that age may not appreciably influence the dimensions or mechanical properties of the human patellar tendon in vivo. Collagen concentration was reduced, whereas both enzymatic and nonenzymatic cross-linking of concentration was elevated in OM vs. in YM, which may be a mechanism to maintain the mechanical properties of tendon with aging.

tendon dimension; tendon mechanical properties; aging; collagen; hydroxylysyl pyridinoline; lysyl pyridinoline; advanced glycation end products

FORCE GENERATED BY MUSCLE is transferred to bones via tendons to produce movement. However, tendons are not entirely inextensible, but exhibit elastic and time-dependant properties that serve to influence the overall function of the muscle-tendon complex (3, 13, 17, 29, 58). Tendons have traditionally been considered relatively inert structures, but several recent

reports have demonstrated that human tendons respond directly to physical activity by increased metabolic activity (15, 35, 42) and increased collagen synthesis (50, 51). Furthermore, strength training and habitual loading of tendons appear to be associated with increased tendon size (6, 21, 46), confirming that the aforementioned response to elevated loading results in a net increase of tendon tissue. These recent findings show that tendons respond and adapt to their specific loading history.

Aging is associated with a decline in muscle mass, strength, and physical function (66). However, although tendon properties influence the overall function of the muscle-tendon complex (58, 67), there is a relative lack of human data that describe possible age-associated changes in mechanical properties of tendon. Animal data show that aging yields a stronger and stiffer tendon (34, 69, 83), a weaker and more compliant tendon (27, 90, 91), or leaves the tendon unchanged (39); thus these data are inconclusive. In contrast, data on isolated human cadaver tendon suggest that the aging process largely leaves the mechanical properties unaltered (31, 40, 41). Despite the development of ultrasonography-based methods to evaluate human tendon properties in vivo (33, 37, 53), the effect of aging on the mechanical properties of human tendon in vivo remains elusive. Tendon strain in humans has been shown to decrease (47–49) or increase (43, 52, 61, 63, 70) with aging. Yet others have shown that aging leaves the mechanical properties of the patellar tendon unchanged (18). Thus the picture is incoherent, which may partly be related to methodological and design differences, the physical activity level of the sample population, and the type of tendon tested, i.e., the tendon-aponeurosis complex or the free tendon alone.

In tendon, the trivalent intermolecular pyridinoline cross-links [primarily hydroxylysyl pyridinoline (HP) and lysyl pyridinoline (LP)] stabilize the fibrillar structure of collagen and thus contribute to the mechanical properties of the tendon (7, 9, 12). These cross-links are formed from enzymatically derived covalent immature cross-links, which undergo a spontaneous conversion into more mature trivalent cross-links with collagen maturation (7). The slow turnover of mature collagen allows further cross-linking via the adventitious nonenzymatic reactions of glucose with the lysyl and arginine amino acid residues in the collagen triple helix as a true aging process (7, 59, 62). This nonenzymatic process results in the accumulation of advanced glycation end products (AGE) in tendon tissue. The most widely studied AGE is pentosidine. AGE accumulation is known to accelerate with aging and diabetes (16, 28, 62) and is believed to yield a stiffer and more load-resistant tendon (78, 79). It has been shown that AGE cross-link density in collag-

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enous tissues (19, 38, 81, 86) and in human tendons (11, 84) is markedly higher in older than in younger individuals, and there are only sparse human data on mature cross-link density of tendon and how these are influenced by aging (11). However, recent reports of an age-associated reduction in human tendon stiffness *in vivo* (43, 52, 61, 63, 70) in conjunction with the notion of simultaneously elevated cross-link density and AGE accumulation are difficult to reconcile. To the best of our knowledge, there are currently no human studies examining collagen cross-linking and mechanical properties of the patellar tendon *in vivo* in old and young men. Therefore, the purpose of this study was to examine both the mechanical properties of human patellar tendon *in vivo* and collagen cross-link composition in old (OM) and young men (YM) subjects.

## MATERIALS AND METHODS

**Subjects.** Seven OM ( $67 \pm 3$  years,  $86 \pm 10$  kg) and 10 YM ( $27 \pm 2$  years,  $81 \pm 8$  kg) with similar activity levels volunteered for the study (Table 1). With the use of a standardized form, participants were interviewed regarding how many hours per week they were performing organized sport or exercise on a regular basis (OM:  $5 \pm 6$  h/wk, YM:  $5 \pm 2$  h/wk). There were no differences in physical characteristics between OM and YM. All were healthy, did not take prescription medicine, and had no overt signs or symptoms of diabetes. In addition, the subjects had no known knee or tendon pathology. The study complied with the Declaration of Helsinki and was approved by the local ethics committee. All subjects gave their informed consent before the experiments.

**Muscle and tendon dimensions.** The anatomic cross-sectional area (CSA) of the quadriceps femoris muscle was measured 20 cm proximal to the tibia plateau (mid-thigh level) by magnetic resonance imaging (MRI) (General Electric, Sigma Horizon LX 1.5-Tesla, T1 weighted SE) using a lower extremity coil. The images were obtained using the following parameters: TR/TE = 500/14 ms, field of view (FOV) = 18, matrix =  $512 \times 512$ , and slice thickness = 6 mm (21, 46). Subsequently, the lean muscle mass of the quadriceps muscle (subcutaneous and intermuscular noncontractile tissues were excluded from the measurement) was manually outlined using the software program Osiris 4.19 (<http://www.sim.hcuge.ch/osiris/>). The mean value of three measurements of the same image was used for analysis. Patellar tendon CSA and length were determined with the use of MRI (General Electric, Sigma Horizon LX, 1.5-Tesla, T1 weighted SE) (21, 46). Patellar tendon CSA was determined by axial plane MRI using the following parameters: TR/TE = 400/14 ms, FOV = 20, matrix =  $256 \times 256$ , slice thickness = 5.0 mm, and spacing = 0 mm. The axial scans were performed perpendicular to the patellar tendon. As described in detail previously, the tendon CSA was measured 1) just distal to the patellar insertion, 2) just proximal to the tibia insertion, and 3) midway between these two sites (21, 46). The patellar tendon length was determined from sagittal plane MRI using the following parameters: TR = 500, echo time (ET) =  $3 \times$  (TE: 12.4 ms), FOV = 16, matrix =  $256 \times 192$ , slice thickness = 4.0 mm, and no spacing. The patellar tendon length was obtained by measuring the distance from the dorsal insertion at the patella apex to the dorsal

insertion on the tibia. Patellar tendon CSA and length were manually outlined using the software program Osiris 4.19 (<http://www.sim.hcuge.ch/osiris/>). The color intensity of each image was adjusted using the National Institutes of Health color scale mode of the software. Tendon CSA and length were measured using the grayscale image display. The average tendon CSA was calculated from the three levels (proximal, mid, and distal CSA) and used for analysis. The typical error percent of repeated measures of site-specific tendon CSA was 2–2.5% (21).

**Mechanical properties of tendon.** The details of the measurement, including the reliability of the method in our laboratory, has been reported previously (37). The within-day correlation coefficient and typical error percent results for repeated measures were 0.95 and 9.9% for tendon stiffness, 0.97 and 5.5% for tendon strain, and 0.94 and 9.4% for Young's modulus. Subjects performed a 5-min warm-up on a stationary bike to secure proper preconditioning of the tendon before testing. Thereafter, the subjects were seated in a custom-made rigid chair with both hips and knees flexed to an angle of 90°. A leg cuff, which was connected to a strain gauge (Bofors KRG-4, Bofors, Sweden) through a rigid steel rod perpendicular to the lower leg, was mounted on the leg just above the medial malleolus. An ultrasound probe (7.5 MHz, linear array B-mode; Sonoline Sienna, Siemens, Erlangen, Germany) was fitted into a custom-made rigid cast that was secured to the skin above the patellar tendon in the sagittal plane. The ultrasound probe and cast were positioned so that the patella, the patellar tendon, and the tibia were all visible within the viewing field throughout the ramped contractions (Fig. 1).

The ultrasound S-VHS video images obtained during the ramp trials were sampled at 50 Hz on a personal computer using frame-by-frame capturing software (Matrox Marvel G400-TV, Dorval, Quebec, Canada). Force was sampled on two separate personal computers at 50 Hz via a 12-bit analog-to-digital converter (dt2810A; Data Translation). The two computers were interconnected to permit synchronous sampling of all data using a custom-built trigger device (14). The subjects performed four to five slow isometric knee extensions ramps by applying gradually increasing force until maximum over a 10-s period during which patellar tendon displacement and knee extension force were measured simultaneously. Each ramp was separated by a 2-min rest period. All measurements were performed on one side, randomized to either the right or left knee. During the ramp contractions, force was sampled at 50 Hz and low-pass filtered at a 1.0-Hz cutoff frequency using a fourth-order zero-lag Butterworth filter.

Tendon force was calculated by dividing the estimated total knee extension moment by the internal moment arm, which was estimated from individually measured femur lengths (87). Tendon stress was calculated by dividing tendon force with the average of the three levels (proximal, mid, and distal) of the patellar tendon CSA determined from MRI. Tendon deformation was defined as the change in distance between the patellar apex and the tibia (37, 57). Tendon strain was calculated as the change in length related to the initial tendon length. Each single force-deformation curve was fitted to a second- or third-order polynomial fit, which yielded  $R^2 > 0.98$ . Tendon stiffness ( $\Delta$ force/ $\Delta$ deformation) and Young's modulus ( $\Delta$ stress/ $\Delta$ strain) based on common force were calculated in the final 20% of the force-deformation and stress-strain curves, respectively (57). To compare tendon dimensions between the subjects of various body size, tendon CSA data were normalized to body weight and raised to the power of 2/3 (60).

**Patellar tendon biopsies.** A Bard MAGNUM biopsy instrument (C.R. Bard, Covington, GA) with a disposable core biopsy needle (14 gauge) was used. After sterilization, the skin was injected with local anesthetic (1% lidocaine), and a 3- to 5-mm-long incision was created just distal to the patella apex. The biopsy needle was inserted into the tendon surface at an  $\sim 30^\circ$  angle and fired, securing a tissue sample of  $\sim 8$  mg. Samples were snap-frozen in liquid nitrogen and stored at  $-80^\circ\text{C}$ . Tendon biopsies were taken from the same side as tendon mechanical properties assessments were performed. No previous bi-

Table 1. Physical characteristics of OM and YM

	OM (n = 7)	YM (n = 10)
Age, years	67 $\pm$ 3	27 $\pm$ 2
Height, cm	178 $\pm$ 9	183 $\pm$ 4
Weight, kg	86 $\pm$ 10	81 $\pm$ 8
Activity level, h/wk	5 $\pm$ 6	5 $\pm$ 2

Values are means  $\pm$  SD. There were no differences in physical characteristics between old men (OM) and young men (YM).

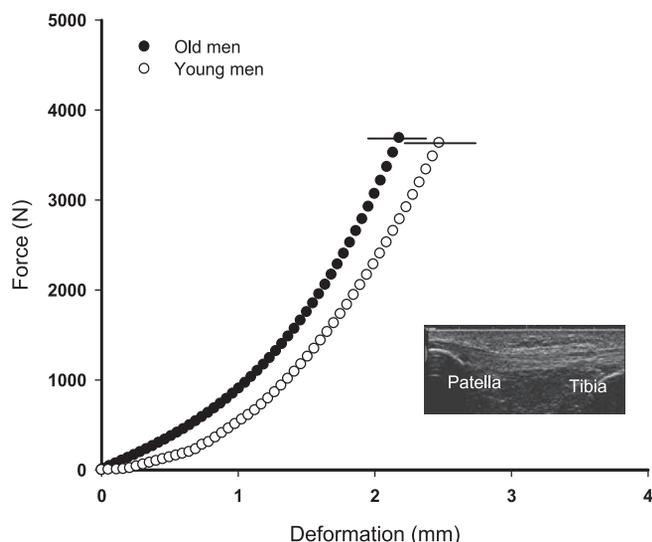


Fig. 1. The patellar tendon force-deformation relationship based on common force. Values are means  $\pm$  SD of all subjects. There were no differences between old (OM) and young men (YM) with respect to tendon deformation or stiffness ( $P > 0.05$ ).

opsy had been taken from that site in all subjects. All biopsy samples were analyzed in an investigator-blinded fashion.

**Biochemical analysis.** Freeze-dried tendon samples were hydrolyzed in 6 M HCl (+108 C, 24 h) and evaporated into dryness and dissolved in H<sub>2</sub>O. Hydroxyproline, the collagen-specific amino acid, was measured spectrophotometrically (23) to quantify collagen protein (24). HP, LP, and pentosidine were analyzed via a single reversed-phase high-performance liquid chromatography (HPLC) run and detected on the basis of their natural fluorescence (10). At 0–16 min, the wavelength for HP and LP fluorescence was 400 nm for emission and 295 nm for excitation. The wavelengths were changed at 16–60 min to 328/378 nm to measure pentosidine. For the elution of the cross-links, a gradient was built up to contain 17% eluent B (75% acetonitrile with 0.13% heptafluorobutyric anhydride) at 0 min and 25% eluent B at 30 min. Eluent A was 0.13% heptafluorobutyric anhydride. Flow rate was 1 ml/min. HP was eluted at 12 min, LP at 13.5 min, and pentosidine at 23 min. The HPLC system used included Quaternary Gradient Pump unit, PU-2089 Plus, Intelligent Autosampler AS-2057 Plus, and Intelligent Fluorescence Detector FP-2020 by Jasco. Data processing software was Jasco Chrompass. The LiChroCART 125-4 column was from Merck Hitachi. The results for HP, LP, and pentosidine are given compared with the standards injected at four different concentrations in each HPLC run. The intra-assay coefficient of variations based on duplicates within a run were 2.6%, 3.7%, and 3.9% for HP, LP, and pentosidine, respectively. The detection limit for HP and LP is 0.4 pmol and 0.05 pmol for pentosidine.

**Data reduction and analysis.** The two isometric ramp contractions that yielded the greatest maximum force were selected for further analysis. To make group comparisons and thereby account for differences in magnitude in isometric ramp contraction force, the trials for all subjects were subsequently analyzed to the lowest common force, as determined by the weakest subject (3953 N). Mann-Whitney *U*-tests were used to examine whether there were differences between the groups in the measured variables. Spearman rank-order correlation was used to analyze the strength of relationships between variables.  $P < 0.05$  was considered significant. Results are reported as means  $\pm$  SD.

## RESULTS

Peak knee extensor moment was lower in OM ( $154 \pm 41$  N·m) than in YM ( $216 \pm 62$  N·m) ( $P < 0.05$ ). Similarly, quadriceps femoris CSA was lower in OM than in YM (Table 2) ( $P < 0.01$ ).

Table 2. Patellar tendon dimensions and quadriceps muscle CSA for OM and YM

	OM ( $n = 7$ )	YM ( $n = 10$ )
Tendon length, mm	43 $\pm$ 4	43 $\pm$ 5
Tendon CSA, mm <sup>2</sup>	101 $\pm$ 17	103 $\pm$ 8
Tendon CSA, mm <sup>2</sup> /body wt <sup>2/3</sup>	5.34 $\pm$ 0.48	5.47 $\pm$ 0.25
Quadriceps muscle CSA, mm <sup>2</sup>	6,214 $\pm$ 705*	8,431 $\pm$ 522

Values are means  $\pm$  SD. CSA, cross-sectional area. \*Significantly different from YM,  $P < 0.01$ .

Tendon dimensions are shown in Table 2. Tendon length did not differ between YM and OM. Absolute average tendon CSA did not differ between OM and YM. Similarly, there was no between group difference in average tendon CSA normalized for body weight.

Mechanical properties determined at maximal force are shown in Table 3. Maximal tendon force was lower in OM than in YM ( $P < 0.05$ ). There were no differences between OM and YM with respect to tendon deformation, stiffness, strain, stress, or Young's modulus based on average tendon CSA. Mechanical properties at a common force are shown in Table 4. Again, there were no differences between OM and YM for any of the variables (Figs. 1 and 2).

Collagen concentration and cross-link density data are shown in Table 5. Collagen concentration was lower in OM than in YM ( $P < 0.05$ ). Both HP and LP ( $P < 0.05$ ) as well as pentosidine ( $P < 0.01$ ) concentrations in collagen were higher in OM than in YM. Pentosidine was positively related to age in YM ( $r = 0.74$ ,  $P < 0.01$ , Fig. 3) but not in OM ( $r = 0.65$ ,  $P = 0.11$ ). There were no significant correlations between the mechanical and biochemical variables.

## DISCUSSION

To the best of our knowledge, this is the first study that has examined both the mechanical properties of the human patellar tendon in vivo and collagen cross-link densities in YM and OM with similar physical activity levels. The main findings were that the collagen concentration was lower in OM than in YM, whereas the enzymatically derived cross-links (HP and LP) were greater in OM than in YM. At the same time, the nonenzymatically derived AGE marker (pentosidine) was markedly more abundant in OM than in YM. However, despite these apparent age-related differences in the tendon collagen properties, the tendon mechanical properties in the two age groups did not diverge appreciably.

It is well known that age is associated with a loss in muscle mass and consequently a reduction in muscle function (66). It

Table 3. Patellar tendon mechanical properties for OM and YM based on maximum force

	OM ( $n = 7$ )	YM ( $n = 10$ )
Force, N	5,161 $\pm$ 737*	7,415 $\pm$ 2184
Deformation, mm	2.6 $\pm$ 0.4	2.9 $\pm$ 0.9
Stiffness, N/mm	3,926 $\pm$ 1091	5,546 $\pm$ 1871
Stress, MPa	51 $\pm$ 8	65 $\pm$ 24
Strain, %	6.1 $\pm$ 0.9	6.9 $\pm$ 2.3
Modulus, GPa	1.7 $\pm$ 0.3	2.2 $\pm$ 0.7

Values are means  $\pm$  SD. \*Significantly different from YM,  $P < 0.05$ .

Table 4. Patellar tendon mechanical properties for OM and YM based on common force

	OM (n = 7)	YM (n = 10)
Deformation, mm	2.3±0.4	2.4±0.6
Stiffness, N/mm	3,511±837	3,290±869
Stress, MPa	41±7	37±5
Strain, %	5.3±0.9	5.9±1.7
Modulus, GPa	1.5±0.4	1.4±0.4

Values are means ± SD.

has also been suggested that tendon compliance and electro-mechanical delay increase with aging (80), which would reduce efficient transfer of contractile force and therefore amplify the age-associated decline in muscle function. However, to what extent the mechanical properties of human tendon change with aging remains unclear. Data based on animal models are disjointed as they suggest that aging reduces (27, 90, 91), augments (34, 69, 83), or leaves the mechanical properties unchanged (39). Similarly, data based on human in vivo models are inconclusive. These studies show that, with aging, the tendon becomes more compliant (43, 52, 61, 63, 70), less compliant (47–49), or remains unchanged (18, 43). In contrast, data on isolated human cadaver preparations consistently suggest that aging does not influence the mechanical properties (31, 40, 41), which is in agreement with the present data and that of Carroll et al. (18) on human patellar tendon in vivo.

The present mechanical data diverge from those of others based on the human in vivo model with one exception (18), and this may be related to methodological differences and study design. It has been shown that strength training and habitual loading pattern may result in tendon hypertrophy (6, 21, 46), which would influence the mechanical properties of the tendon (80). In contrast to previous in vivo studies (18, 43, 47, 52, 61, 63, 70), we have taken this aspect into account by comparing age groups with similar activity levels. However, it cannot be

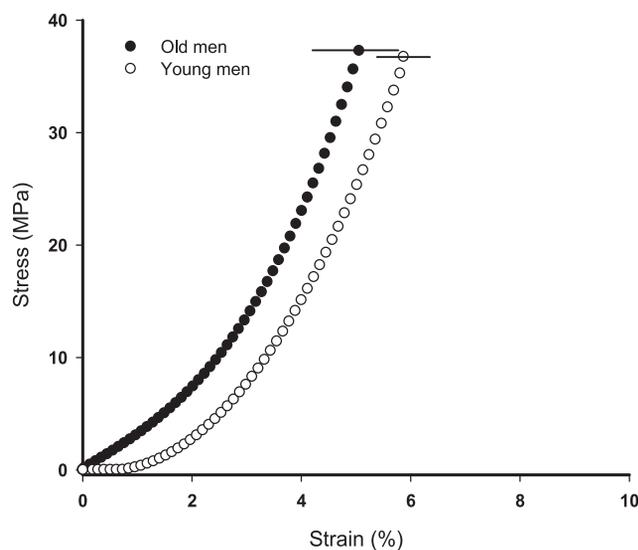


Fig. 2. The patellar tendon stress-strain relationship based on common force. Values are means ± SD of all subjects. There were no differences between OM and YM with respect to tendon strain, stress, or Young's modulus based on average tendon cross-sectional area ( $P > 0.05$ ).

Table 5. Concentration of collagen and hydroxyllysyl pyridinoline, lysyl pyridinoline, and pentosidine cross-links in the patellar tendons of OM and YM

	OM (n = 7)	YM (n = 10)
Collagen, mg/mg dry wt	0.49±0.27*	0.73±0.14
Hydroxyllysyl pyridinoline, mmol/mol collagen	898±172*	645±183
Lysyl pyridinoline, mmol/mol collagen	49±38†	16±8
Pentosidine, mmol/mol collagen	73±13†	11±2

Values are means ± SD. \*,†Significantly different from the values of YM ( $P < 0.05$  and  $P < 0.01$ , respectively).

ruled out that prior life-long training history, including exercise mode and intensity, which was unaccounted for in the study, may have influenced the data. Furthermore, it has been shown that the ultrasonography-based method of obtaining patellar tendon deformation requires that the movement of the tibia also has to be considered (37, 71), which was achieved in the present study and in the study by Carroll et al. (18). Notably, the present data and that of Carroll et al. (18) cannot demonstrate any age-associated difference in mechanical properties of the patellar tendon. Moreover, several studies have investigated the effect of aging on the mechanical properties based on a composite measure of deformation that includes both that of the tendon and aponeurosis, rather than the tendon per se (43, 52, 61, 63, 70). This makes direct comparisons of results difficult since the free tendon and aponeurosis have dissimilar mechanical properties and because the stiffness of the aponeurosis can be modulated during contraction (30, 57). It should also be recognized that the present data and that of others are based on cross-sectional designs with inherent limitations, including the striking variations in tendon mechanical properties between subjects (54); therefore, a type II error cannot be ruled out.

The densities of mature lysyl oxidase-derived intermolecular covalent cross-links, such as HP and LP, gradually increase during tendon tissue maturation, and it is commonly believed that these cross-links are the chief contributors to the function and mechanical properties of the tendon (7, 9, 12, 76). In animal models, it has been shown that there is a positive

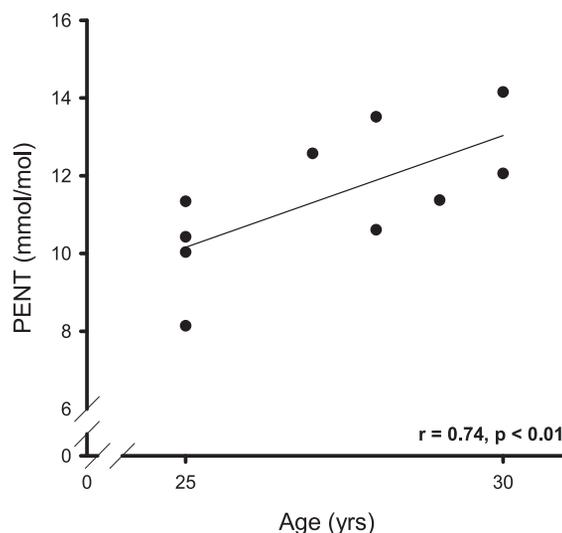


Fig. 3. Pentosidine was positively related to age in YM ( $r = 0.74$ ,  $P < 0.01$ ).

relationship between HP cross-link density and mechanical strength of the healing medial collateral ligament tissue and anterior cruciate ligament graft (32, 68) and furthermore that changes in HP and LP density will result in altered mechanical properties in collagenous tissue (7, 72). However, there are only sparse human data on mature cross-link density in tendon and how these are influenced by aging (11). A small but significant age-related increase of LP has been demonstrated in the supraspinatus tendon in vitro (11). The present data on LP and HP densities of YM correspond well with previously reported data in a similar population (45). However, the present data also demonstrate that HP and LP densities of the human patellar tendon were  $\sim 40\%$  and threefold higher, respectively, in OM than in YM, suggesting a rather marked age-associated elevation in these enzymatic cross-links. It is noteworthy that, despite the rather robust difference in both HP and LP, there was no difference in the mechanical properties of the tendon. However, it should be noted that fibril length (22, 77), fibril diameter (74), and proteoglycans and glycosaminoglycans (44, 73, 75) have all been implicated in contributing to the tendon mechanical properties, and the relative contribution of these factors and that of mature cross-links remains elusive.

To our knowledge, these are the first data showing that AGE is markedly increased in the human patellar tendon of OM vs. in that of YM. AGE cross-links, including pentosidine, are formed when lysine amino acid residues in the collagen triple helix come into contact with glucose (7, 62), and the accumulation of these cross-links is known to accelerate with aging (7, 44) and disease processes such as diabetes (16, 28, 62), atherosclerosis (88), Alzheimer (20, 64), and renal failure (89). AGE products are also used as markers of tissue turnover (11). It has been shown that the difference between young and older individuals in AGE cross-link density in collagen is  $\sim 2$ -fold in human skeletal muscle (38), 5-fold in human bone (81), 7-fold in human tendon (present data), 9-fold in human ligaments (19), and 33-fold in human cadaver cartilage (86), demonstrating the tissue-specific turnover. The fact that pentosidine density is sevenfold greater in tendon of OM than in YM (Table 5), coupled with the fact that pentosidine appears to be related to age in a narrow age span (Fig. 3), firmly demonstrates the positive relationship between AGE cross-linking of human patellar tendon collagen and aging *ex vivo*. These data corroborate and extend those previously reported on cadaver tissue (11, 84). From a functional standpoint, an elevated AGE cross-link density has been suggested to result in increased tensile stress and tendon stiffness in animal models (4, 5, 8, 34, 78, 79, 85). However, in the present study, both the stiffness (Fig. 1) and the Young's modulus (Fig. 2) of the tendon did not differ between OM and YM despite the sevenfold difference in pentosidine, suggesting that factors other than AGE may also play a major role in determining the mechanical properties of human tendon.

In the present study, the total collagen concentration of the patellar tendon was  $\sim 34\%$  lower in OM than in YM (Table 5), and this age-linked reduction is in accordance with that found in the canine patellar tendon (39) and rat tail tendon (90). It is possible that the lower collagen concentration with aging may represent the reduced size and/or density of collagen fibrils that is known to occur with aging (26, 65, 74, 76, 82). It was recently reported that MRI signal intensity of the patellar tendon was reduced with aging (18), which may be a function

of the reduction in collagen concentration observed in the present study. Unfortunately, the size of the obtained biopsy in the present study precluded transmission electron microscopy analysis for fibril size and density. The lower collagen concentration in OM may be an age-related change in the tendon *per se* and/or a function of reduced tendon loading due to an age-related loss of muscle mass/strength. Interestingly, the average whole tendon CSA did not differ between OM and YM (Table 2), which in light of the reduced collagen concentration may be related to increased amounts of other extracellular matrix components, such as proteoglycans and glycosaminoglycans. Alternatively, the retained tendon CSA in OM may result from tendon intrafibrillar fat that can accumulate with aging (1, 2, 25). It has previously been demonstrated that the Achilles tendon CSA is larger in postmenopausal women than in young women (55), but it should be noted that female hormones may significantly impact collagen synthesis (36, 56). This potential sex-dependent factor is the reason that we have undertaken the present study in men only.

Albeit speculative, it is possible that the elevated enzymatic and/or nonenzymatic cross-link density in OM vs. that shown in YM served to maintain tendon stiffness and Young's modulus despite the diminished collagen concentration. Such maintenance of tendon stiffness would serve to maintain effective transfer of muscle force despite a lower absolute muscle size (Table 2) and strength (Table 3). In this context, it should be noted that the present data were obtained in moderately physically active individuals, and future studies will need to address the effect of training *per se* in elderly.

In conclusion, results from the present study raise the possibility that the dimensions and mechanical properties of the human patellar tendon *in vivo* may not differ between OM and YM. On the other hand, the OM group displayed lower collagen concentration, but greater enzymatic (HP and LP) and nonenzymatic (pentosidine) collagen cross-links, than YM. This age-related increase in both enzymatic and nonenzymatic cross-linking compounds may serve to maintain the mechanical properties of tendon with aging.

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#### REFERENCES

1. Adams CW, Bayliss OB. Acid mucosubstances underlying lipid deposits in ageing tendons and atherosclerotic arteries. *Atherosclerosis* 18: 191–195, 1973.
2. Adams CW, Bayliss OB, Baker RW, Abdulla YH, and Hunter-Craig CJ. Lipid deposits in ageing human arteries, tendons and fascia. *Atherosclerosis* 19: 429–440, 1974.
3. Alexander RM, Net-Clark HC. Storage of elastic strain energy in muscle and other tissues. *Nature* 265: 114–117, 1977.
4. Andreassen TT, Oxlund H, Danielsen CC. The influence of non-enzymatic glycosylation and formation of fluorescent reaction products on the mechanical properties of rat tail tendons. *Connect Tissue Res* 17: 1–9, 1988.
5. Andreassen TT, Seyer-Hansen K, Bailey AJ. Thermal stability, mechanical properties and reducible cross-links of rat tail tendon in experimental diabetes. *Biochim Biophys Acta* 677: 313–317, 1981.

6. Arampatzis A, Karamanidis K, Albracht K. Adaptational responses of the human Achilles tendon by modulation of the applied cyclic strain magnitude. *J Exp Biol* 210: 2743–2753, 2007.
7. Avery NC, Bailey AJ. Enzymic and non-enzymic cross-linking mechanisms in relation to turnover of collagen: relevance to aging and exercise. *Scand J Med Sci Sports* 15: 231–240, 2005.
8. Bai P, Phua K, Hardt T, Cernadas M, Brodsky B. Glycation alters collagen fibril organization. *Connect Tissue Res* 28: 1–12, 1992.
9. Bailey AJ. Molecular mechanisms of ageing in connective tissues. *Mech Ageing Dev* 122: 735–755, 2001.
10. Bank RA, Beekman B, Verzijl N, de Roos JA, Sakkee AN, TeKoppele JM. Sensitive fluorimetric quantitation of pyridinium and pentosidine crosslinks in biological samples in a single high-performance liquid chromatographic run. *J Chromatogr B Biomed Sci Appl* 703: 37–44, 1997.
11. Bank RA, TeKoppele JM, Oostingh G, Hazleman BL, Riley GP. Lysylhydroxylation and non-reducible crosslinking of human supraspinatus tendon collagen: changes with age and in chronic rotator cuff tendinitis. *Ann Rheum Dis* 58: 35–41, 1999.
12. Barnard K, Light ND, Sims TJ, Bailey AJ. Chemistry of the collagen cross-links. Origin and partial characterization of a putative mature cross-link of collagen. *Biochem J* 244: 303–309, 1987.
13. Biewener A, Baudinette R. In vivo muscle force and elastic energy storage during steady-speed hopping of tamar wallabies (*Macropus eugenii*). *J Exp Biol* 198: 1829–1841, 1995.
14. Bojsen-Moller J, Hansen P, Aagaard P, Kjaer M, Magnusson SP. Measuring mechanical properties of the vastus lateralis tendon-aponeurosis complex in vivo by ultrasound imaging. *Scand J Med Sci Sports* 13: 259–265, 2003.
15. Bojsen-Moller J, Kalliokoski KK, Seppanen M, Kjaer M, Magnusson SP. Low-intensity tensile loading increases intratendinous glucose uptake in the Achilles tendon. *J Appl Physiol* 101: 196–201, 2006.
16. Brownlee M. Advanced protein glycosylation in diabetes and aging. *Annu Rev Med* 46: 223–234, 1995.
17. Butler DL, Grood ES, Noyes FR, Zernicke RF. Biomechanics of ligaments and tendons. *Exerc Sport Sci Rev* 6: 125–181, 1978.
18. Carroll CC, Dickinson JM, Haus JM, Lee GA, Hollon CJ, Aagaard P, Magnusson SP, Trappe TA. Influence of aging on the in vivo properties of human patellar tendon. *J Appl Physiol* 105: 1907–1915, 2008.
19. Chen JR, Takahashi M, Kushida K, Suzuki M, Suzuki K, Horiuchi K, Nagano A. Direct detection of crosslinks of collagen and elastin in the hydrolysates of human yellow ligament using single-column high performance liquid chromatography. *Anal Biochem* 278: 99–105, 2000.
20. Colaco CA, Ledesma MD, Harrington CR, Avila J. The role of the Maillard reaction in other pathologies: Alzheimer's disease. *Nephrol Dial Transplant* 11, Suppl 5: 7–12, 1996.
21. Coupe C, Kongsgaard M, Aagaard P, Hansen P, Bojsen-Moller J, Kjaer M, Magnusson SP. Habitual loading results in tendon hypertrophy and increased stiffness of the human patellar tendon. *J Appl Physiol* 105: 805–810, 2008.
22. Craig AS, Birtles MJ, Conway JF, Parry DA. An estimate of the mean length of collagen fibrils in rat tail-tendon as a function of age. *Connect Tissue Res* 19: 51–62, 1989.
23. Creemers LB, Jansen DC, van Veen-Reurings A, van den BT, Everts V. Microassay for the assessment of low levels of hydroxyproline. *Bio-techniques* 22: 656–658, 1997.
24. Creemers LB, Jansen DC, van Veen-Reurings A, van den BT, Everts V. Microassay for the assessment of low levels of hydroxyproline. *Bio-techniques* 22: 656–658, 1997.
25. Crouse JR, Grundy SM, Ahrens EH Jr. Cholesterol distribution in the bulk tissues of man: variation with age. *J Clin Invest* 51: 1292–1296, 1972.
26. Curwin SL, Roy RR, Vailas AC. Regional and age variations in growing tendon. *J Morphol* 221: 309–320, 1994.
27. Dressler MR, Butler DL, Wenstrup R, Awad HA, Smith F, Boivin GP. A potential mechanism for age-related declines in patellar tendon biomechanics. *J Orthop Res* 20: 1315–1322, 2002.
28. Dyer DG, Dunn JA, Thorpe SR, Bailie KE, Lyons TJ, McCance DR, Baynes JW. Accumulation of Maillard reaction products in skin collagen in diabetes and aging. *J Clin Invest* 91: 2463–2469, 1993.
29. Elliott DH. Structure and function of mammalian tendon. *Biol Rev Camb Philos Soc* 40: 392–421, 1965.
30. Finni T, Hodgson JA, Lai AM, Edgerton VR, Sinha S. Nonuniform strain of human soleus aponeurosis-tendon complex during submaximal voluntary contractions in vivo. *J Appl Physiol* 95: 829–837, 2003.
31. Flahiff CM, Brooks AT, Hollis JM, Vander Schilden JL, Nicholas RW. Biomechanical analysis of patellar tendon allografts as a function of donor age. *Am J Sports Med* 23: 354–358, 1995.
32. Frank C, McDonald D, Wilson J, Eyre D, Shrive N. Rabbit medial collateral ligament scar weakness is associated with decreased collagen pyridinoline crosslink density. *J Orthop Res* 13: 157–165, 1995.
33. Fukashiro S, Itoh M, Ichinose Y, Kawakami Y, Fukunaga T. Ultrasonography gives directly but noninvasively elastic characteristic of human tendon in vivo. *Eur J Appl Physiol Occup Physiol* 71: 555–557, 1995.
34. Galeski A, Kastelic J, Baer E, Kohn RR. Mechanical and structural changes in rat tail tendon induced by alloxan diabetes and aging. *J Biomech* 10: 775–782, 1977.
35. Hannukainen J, Kalliokoski KK, Nuutila P, Fujimoto T, Kemppainen J, Viljanen T, Laaksonen MS, Parkkola R, Knuuti J, Kjaer M. In vivo measurements of glucose uptake in human Achilles tendon during different exercise intensities. *Int J Sports Med* 26: 727–731, 2005.
36. Hansen M, Kongsgaard M, Holm L, Skovgaard D, Magnusson SP, Qvortrup K, Larsen JO, Dahl M, Serup A, Frystyk J, Flyvbjerg A, Langberg H, Kjaer M. Effect of estrogen on tendon collagen synthesis, tendon structural characteristics, and biomechanical properties in postmenopausal women. *J Appl Physiol* 106: 1385–1393, 2009.
37. Hansen P, Bojsen-Moller J, Aagaard P, Kjaer M, Magnusson SP. Mechanical properties of the human patellar tendon, in vivo. *Clin Biomech (Bristol, Avon)* 21: 54–58, 2006.
38. Haus JM, Carrithers JA, Trappe SW, Trappe TA. Collagen, cross-linking, and advanced glycation end products in aging human skeletal muscle. *J Appl Physiol* 103: 2068–2076, 2007.
39. Haut RC, Lancaster RL, DeCamp CE. Mechanical properties of the canine patellar tendon: some correlations with age and the content of collagen. *J Biomech* 25: 163–173, 1992.
40. Hubbard RP, Soutas-Little RW. Mechanical properties of human tendon and their age dependence. *J Biomech Eng* 106: 144–150, 1984.
41. Johnson GA, Tramaglini DM, Levine RE, Ohno K, Choi NY, Woo SL. Tensile and viscoelastic properties of human patellar tendon. *J Orthop Res* 12: 796–803, 1994.
42. Kalliokoski KK, Langberg H, Ryberg AK, Scheede-Bergdahl C, Doessing S, Kjaer A, Boushel R, Kjaer M. The effect of dynamic knee-extension exercise on patellar tendon and quadriceps femoris muscle glucose uptake in humans studied by positron emission tomography. *J Appl Physiol* 99: 1189–1192, 2005.
43. Karamanidis K, Arampatzis A. Mechanical and morphological properties of human quadriceps femoris and triceps surae muscle-tendon unit in relation to aging and running. *J Biomech* 39: 406–417, 2006.
44. Kjaer M. Role of extracellular matrix in adaptation of tendon and skeletal muscle to mechanical loading. *Physiol Rev* 84: 649–698, 2004.
45. Kongsgaard M, Kovanen V, Aagaard P, Doessing S, Hansen P, Kjaer M, Magnusson SP. Peritendinous corticosteroid injections, eccentric decline squat training and heavy slow resistance training in patellar tendinopathy. *Scand J Med Sci Sports* In press.
46. Kongsgaard M, Reitelseder S, Pedersen TG, Holm L, Aagaard P, Kjaer M, Magnusson SP. Region specific patellar tendon hypertrophy in humans following resistance training. *Acta Physiol (Oxf)* 191: 111–121, 2007.
47. Kubo K, Ishida Y, Komuro T, Tsunoda N, Kanehisa H, Fukunaga T. Age-related differences in the force generation capabilities and tendon extensibilities of knee extensors and plantar flexors in men. *J Gerontol A Biol Sci Med Sci* 62: 1252–1258, 2007.
48. Kubo K, Kanehisa H, Miyatani M, Tachi M, Fukunaga T. Effect of low-load resistance training on the tendon properties in middle-aged and elderly women. *Acta Physiol Scand* 178: 25–32, 2003.
49. Kubo K, Morimoto M, Komuro T, Tsunoda N, Kanehisa H, Fukunaga T. Age-related differences in the properties of the plantar flexor muscles and tendons. *Med Sci Sports Exerc* 39: 541–547, 2007.
50. Langberg H, Rosendal L, Kjaer M. Training-induced changes in peritendinous type I collagen turnover determined by microdialysis in humans. *J Physiol* 534: 297–302, 2001.
51. Langberg H, Skovgaard D, Petersen LJ, Bulow J, Kjaer M. Type I collagen synthesis and degradation in peritendinous tissue after exercise determined by microdialysis in humans. *J Physiol* 521: 299–306, 1999.
52. Mademli L, Arampatzis A, Walsh M. Age-related effect of static and cyclic loadings on the strain-force curve of the vastus lateralis tendon and aponeurosis. *J Biomech Eng* 130: 011007, 2008.
53. Maganaris CN, Paul JP. In vivo human tendon mechanical properties. *J Physiol* 521: 307–313, 1999.

54. Magnusson SP, Aagaard P, Dyhre-Poulsen P, Kjaer M. Load-displacement properties of the human triceps surae aponeurosis in vivo. *J Physiol* 531: 277–288, 2001.
55. Magnusson SP, Beyer N, Abrahamsen H, Aagaard P, Neergaard K, Kjaer M. Increased cross-sectional area and reduced tensile stress of the Achilles tendon in elderly compared with young women. *J Gerontol A Biol Sci Med Sci* 58: 123–127, 2003.
56. Magnusson SP, Hansen M, Langberg H, Miller B, Haraldsson B, Westh EK, Koskinen S, Aagaard P, Kjaer M. The adaptability of tendon to loading differs in men and women. *Int J Exp Pathol* 88: 237–240, 2007.
57. Magnusson SP, Hansen P, Aagaard P, Brond J, Dyhre-Poulsen P, Bojsen-Moller J, Kjaer M. Differential strain patterns of the human gastrocnemius aponeurosis and free tendon, in vivo. *Acta Physiol Scand* 177: 185–195, 2003.
58. Magnusson SP, Narici MV, Maganaris CN, Kjaer M. Human tendon behaviour and adaptation, in vivo. *J Physiol* 586: 71–81, 2008.
59. Maillard L. Action des acides aminés sur les sucres: formation des mélanoidines par voie méthodique. *C R Acad Sci* 154: 66–68, 1912.
60. Markovic G, Jaric S. Movement performance and body size: the relationship for different groups of tests. *Eur J Appl Physiol* 92: 139–149, 2004.
61. Mian OS, Thom JM, Ardigo LP, Minetti AE, Narici MV. Gastrocnemius muscle-tendon behaviour during walking in young and older adults. *Acta Physiol (Oxf)* 189: 57–65, 2007.
62. Monnier VM. Toward a Maillard reaction theory of aging. *Prog Clin Biol Res* 304: 1–22, 1989.
63. Morse CI, Thom JM, Birch KM, Narici MV. Tendon elongation influences the amplitude of interpolated doublets in the assessment of activation in elderly men. *J Appl Physiol* 98: 221–226, 2005.
64. Munch G, Schinzel R, Loske C, Wong A, Durany N, Li JJ, Vlassara H, Smith MA, Perry G, Riederer P. Alzheimer's disease—synergistic effects of glucose deficit, oxidative stress and advanced glycation end-products. *J Neural Transm* 105: 439–461, 1998.
65. Nakagawa Y, Majima T, Nagashima K. Effect of ageing on ultrastructure of slow and fast skeletal muscle tendon in rabbit Achilles tendons. *Acta Physiol Scand* 152: 307–313, 1994.
66. Narici MV, Maganaris C, Reeves N. Myotendinous alterations and effects of resistive loading in old age. *Scand J Med Sci Sports* 15: 392–401, 2005.
67. Narici MV, Maganaris CN. Plasticity of the muscle-tendon complex with disuse and aging. *Exerc Sport Sci Rev* 35: 126–134, 2007.
68. Ng GY, Oakes BW, Deacon OW, McLean ID, Eyre DR. Long-term study of the biochemistry and biomechanics of anterior cruciate ligament-patellar tendon autografts in goats. *J Orthop Res* 14: 851–856, 1996.
69. Nielsen HM, Skalicky M, Viidik A. Influence of physical exercise on aging rats. III. Life-long exercise modifies the aging changes of the mechanical properties of limb muscle tendons. *Mech Ageing Dev* 100: 243–260, 1998.
70. Onambele GL, Narici MV, Maganaris CN. Calf muscle-tendon properties and postural balance in old age. *J Appl Physiol* 100: 2048–2056, 2006.
71. Onambele GN, Burgess K, Pearson SJ. Gender-specific in vivo measurement of the structural and mechanical properties of the human patellar tendon. *J Orthop Res* 25: 1635–1642, 2007.
72. Oxlund H, Barckman M, Ortoft G, Andreassen TT. Reduced concentrations of collagen cross-links are associated with reduced strength of bone. *Bone* 17: 365S–371S, 1995.
73. Parry DA. The molecular and fibrillar structure of collagen and its relationship to the mechanical properties of connective tissue. *Biophys Chem* 29: 195–209, 1988.
74. Parry DA, Barnes GR, Craig AS. A comparison of the size distribution of collagen fibrils in connective tissues as a function of age and a possible relation between fibril size distribution and mechanical properties. *Proc R Soc Lond B Biol Sci* 203: 305–321, 1978.
75. Partington FR, Wood GC. The role of non-collagen components in the mechanical behaviour of tendon fibres. *Biochim Biophys Acta* 69: 485–495, 1963.
76. Patterson-Kane JC, Parry DA, Birch HL, Goodship AE, Firth EC. An age-related study of morphology and cross-link composition of collagen fibrils in the digital flexor tendons of young thoroughbred horses. *Connect Tissue Res* 36: 253–260, 1997.
77. Provenzano PP, Vanderby R Jr. Collagen fibril morphology and organization: implications for force transmission in ligament and tendon. *Matrix Biol* 25: 71–84, 2006.
78. Reddy GK. Cross-linking in collagen by nonenzymatic glycation increases the matrix stiffness in rabbit achilles tendon. *Exp Diabetes Res* 5: 143–153, 2004.
79. Reddy GK, Stehno-Bittel L, Enwemeka CS. Glycation-induced matrix stability in the rabbit achilles tendon. *Arch Biochem Biophys* 399: 174–180, 2002.
80. Reeves ND, Maganaris CN, Narici MV. Effect of strength training on human patella tendon mechanical properties of older individuals. *J Physiol* 548: 971–981, 2003.
81. Saito M, Marumo K, Fujii K, Ishioka N. Single-column high-performance liquid chromatographic-fluorescence detection of immature, mature, and senescent cross-links of collagen. *Anal Biochem* 253: 26–32, 1997.
82. Sargon MF, Ozlu K, Oken F. Age-related changes in human tendo calcaneus collagen fibrils. *Saudi Med J* 26: 425–428, 2005.
83. Shadwick RE. Elastic energy storage in tendons: mechanical differences related to function and age. *J Appl Physiol* 68: 1033–1040, 1990.
84. Suzuki D, Takahashi M, Abe M, Nagano A. Biochemical study of collagen and its crosslinks in the anterior cruciate ligament and the tissues used as a graft for reconstruction of the anterior cruciate ligament. *Connect Tissue Res* 49: 42–47, 2008.
85. Verzar F. The aging of collagen. *Sci Am* 208: 104–114, 1963.
86. Verzijl N, DeGroot J, Oldehinkel E, Bank RA, Thorpe SR, Baynes JW, Bayliss MT, Bijlsma JW, Lafeber FP, TeKoppele JM. Age-related accumulation of Maillard reaction products in human articular cartilage collagen. *Biochem J* 350: 381–387, 2000.
87. Visser JJ, Hoogkamer JE, Bobbert MF, Huijting PA. Length and moment arm of human leg muscles as a function of knee and hip-joint angles. *Eur J Appl Physiol Occup Physiol* 61: 453–460, 1990.
88. Vlassara H. Advanced glycation end-products and atherosclerosis. *Ann Med* 28: 419–426, 1996.
89. Vlassara H. Protein glycation in the kidney: role in diabetes and aging. *Kidney Int* 49: 1795–1804, 1996.
90. Vogel HG. Influence of maturation and age on mechanical and biochemical parameters of connective tissue of various organs in the rat. *Connect Tissue Res* 6: 161–166, 1978.
91. Vogel HG. Age dependence of mechanical properties of rat tail tendons (hysteresis experiments). *Aktuelle Gerontol* 13: 22–27, 1983.

# The effects of immobilization on the mechanical properties of the patellar tendon in old men

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## Abstract

It remains unknown if inactivity changes the mechanical properties of the human patellar tendon in old healthy persons. **Purpose:** To examine the effects of short-term unilateral immobilization on the structural and mechanical properties of the patellar tendon in old men (OM), *in vivo*. **Methods:** Eight OM (67±4 yrs, 87±10 kg) with a physical activity level (5±6 hrs/wk) underwent 14 days of unilateral immobilization. They were assessed on the immobilized and control side before and after the intervention. Peak knee extensor moment was assessed during isometric MVC. MRI was used to assess whole patellar tendon dimensions. The mechanical properties of the patellar tendon were assessed using simultaneous force and ultrasonographic measurements during isometric ramp contractions. **Results:** Peak knee extensor moment decreased on the immobilized side ( $P<0.05$ ), while a trend ( $P=0.078$ ) was observed for the control side. On the immobilized side, tendon stiffness (Pre: 2949±799 vs. Post: 2366±774 N mm<sup>-1</sup>,  $P<0.01$ ) and Young's Modulus (Pre: 1.2±0.3 vs. Post: 1.0±0.3 GPa,  $P<0.05$ ) declined with immobilization. On the control side, tendon stiffness (Pre: 3340±1209 vs. Post: 2230±503,  $P<0.01$ ) and Young's Modulus (Pre: 1.5±0.4 vs. Post: 0.9±0.3, GPa,  $P<0.05$ ) also decreased with immobilization. **Conclusion:** The data show that the mechanical properties of the patellar tendon decreased on both sides. A decline in stiffness and Young's modulus were observed without any sizable changes of the tendon, suggesting that the material properties of the tendon were affected by unloading. These findings indicate that short-term inactivity has a substantial negative influence on the mechanical properties of the human patellar tendon in old men.

## Introduction

Human locomotion comes about when force created by contracting muscle is transmitted to the bone via aponeurosis and tendon, and the viscoelastic properties of the tendon will influence the overall function of the muscle–tendon complex (2; 8; 12; 21; 44). In contrast to previous belief, recent data show that in vivo human tendon tissue is not inert but rather responsive to both single bouts as well as habitual mechanical loading (10; 36; 37; 49). In fact, strength training and habitual exercise loading of tendons appear to be associated with an increase in tendon size and stiffness, respectively (4; 16; 32; 58). These data demonstrate that tendons respond and adapt to loading history.

It is well known that immobilization results in a rapid loss of muscle volume and strength, which will negatively influence muscle function (44; 52; 62). The effects of immobilization on the mechanical tendon properties of the tendon have been studied extensively in animal models (11; 63), and several studies show a decline of the mechanical properties of the tendon (22; 39; 47; 48; 56; 67; 68), while some demonstrate the opposite (6; 20). Human studies also suggest that the tendon undergoes a substantial reduction in the mechanical properties with immobilization (17; 33; 35; 54; 59; 60; 69), which have been principally attributed to changes in material rather than size (17; 33; 35; 54; 59; 60; 69). However, the aforementioned effects of immobilization on the mechanical properties of tendon have mainly been investigated in young persons, and thus it remains unknown if similar or more dramatic changes may occur in older healthy persons.

The purpose of the present study was therefore to examine the effects of short-term unilateral immobilization on the human patellar tendon structural and mechanical properties in old men (OM), in vivo.

## Materials & methods

### *Subjects and design*

Eight old men (OM) (67±4 yrs, 87±10 kg, 178±8) with an activity level of 5±6 hrs/wk volunteered for the study. All were healthy, did not take prescription medication, had no overt signs or symptoms of diabetes, and had no known knee or tendon pathology. The study complied with the Declaration of Helsinki and was approved by the local Ethics Committee. All subjects gave their informed consent prior to the experiment.

### *Immobilization*

All subjects had one lower limb randomly assigned to an immobilization intervention that consisted a 30 degree knee flexion full lightweight fiber leg cast from above the malleoli to the groin for 2 weeks. This model has earlier been shown to effectively prevent walking ability and induce significant muscle atrophy in short-

term immobilization studies (26-28; 62). The contralateral limb served as a control. The subjects were instructed to use crutches, avoid ground contact with the casted leg and not to activate the muscles of their casted side during the 2 week study period. All subjects were carefully instructed to actively perform plantar flexor and extensor contractions several times a day in order to prevent potential venous thrombosis. All measurements were performed before and after the immobilization period on both sides.

### *Tendon dimensions*

Patellar tendon cross-sectional area (CSA) and length were determined with the use of MRI (General Electric, Sigma Horizon LX 1.5 Tesla, T1 weighted SE) (16; 32). Patellar tendon CSA was determined by axial plane MR using the following parameters: TR/TE 400/14 ms, FOV 20, matrix 256x256, slice thickness 5.0 mm and spacing 0 mm. The axial scans were performed perpendicular to the patellar tendon. As described in detail elsewhere, the tendon CSA was measured I) just distal to the patellar insertion, II) just proximal to the tibia insertion and III) midway between these two sites (16; 32). The patellar tendon length was determined from sagittal plane MRI using the following parameters: TR 500, ET: 3 x (TE: 12.4 ms), FOV 16, matrix 256x192, slice thickness 4.0 mm and no spacing. Patellar tendon length was obtained by measuring the distance from the dorsal insertion at the patella apex to the dorsal insertion on the tibia. Patellar tendon CSA and length were manually outlined using the software program Osiris 4.19 (<http://www.sim.hcuge.ch/osiris/>). The color intensity of each image was adjusted using the NIH color scale mode of the software. Tendon CSA and length was measured using the gray scale image display. Average tendon CSA was calculated from the 3 sites (proximal, mid and distal CSA) and was used for analysis (15). The typical error percent of repeated measures of site-specific tendon CSA was 2-2.5 % (16). The MRI investigator was blinded with regards to side.

### *Maximal voluntary contraction (MVC)*

Maximal voluntary strength was determined by isometric knee extension efforts. As described in detail elsewhere (9) the subjects were seated in a custom made rigid chair with both hips and knees flexed to an angle of 90°. A leg cuff was connected to a strain gauge (Bofors KRG-4, Bofors, Sweden) through a rigid steel rod perpendicular to the lower leg and was mounted on the leg just above the medial malleolus. The tibia moment arm was measured (from the point of fixation to the lateral epicondyle of the knee) to calculate the knee extensor moment. Force was sampled on a PC at 50 Hz via a 12-bit A/D converter (dt 2810A, Data Translation, MA, USA). The subjects performed 4-5 maximal voluntary isometric knee

extension contractions (MVC) separated by one min (9). The tests were conducted on both sides.

#### *Mechanical properties of tendon*

The details of the measurement, including the reliability of the method in our laboratory, have been reported previously (23). The within-day correlation coefficients and typical error percent for repeated measures were 0.95 and 9.9% for tendon stiffness, 0.97 and 5.5% for tendon strain, and 0.94 and 9.4% for Young's Modulus, respectively. Subjects performed a 5-min warm-up on a stationary bike (~50 W) in order to secure proper preconditioning of the tendon prior to testing. Thereafter the subjects were seated in a custom made rigid chair with both hips and knees flexed 90°. A leg cuff, which was connected to a strain gauge (Bofors KRG-4, Bofors, Sweden) through a rigid steel rod perpendicular to the lower leg, was mounted on the leg just above the medial malleolus. An ultrasound probe (7.5 MHz, linear array B-mode, Sonoline Sienna, Siemens, Erlangen, Germany) was fitted into a custom made rigid cast that was secured to the skin above the patellar tendon in the sagittal plane. The ultrasound probe and cast was positioned so that the patella, the patellar tendon and the tibia were all visible within the viewing field throughout the ramped contractions (Figures 1 & 2).

Ultrasound S-VHS video images of the patella tendon and its bony attachments were sampled during the ramp trials at 50 Hz on a PC using frame-by-frame capturing software (Matrox Marvel G400-TV, Dorval, Canada). Deformation images and force signals were sampled (50 Hz) on two separate PC's, for the latter using a 12-bit A/D converter (dt 2810A, Data Translation, MA, USA). The two computers were inter-connected to permit synchronous sampling of all data using a custom-built trigger device (9). The subjects performed 4-5 slow isometric knee extensions ramps by applying gradually increasing force until maximum over a 10 s period during which patellar tendon displacement and knee extension force were measured simultaneously. Each ramp contraction was separated by a 2-minute rest period. All measurements were performed on both sides, before and after the period of immobilization. All force signals were low-pass filtered at a 1.0 Hz cutoff frequency using a 4<sup>th</sup> order zero-lag Butterworth filter.

Patella tendon force was calculated by dividing the estimated total knee extension moment by the internal moment arm, which was estimated from individually measured femur lengths (66). Tendon stress was calculated by dividing tendon force with the average patellar tendon CSA determined from MRI (proximal, mid, distal sites). Tendon deformation was defined as the change in distance between the patellar apex and the tibia (23; 43). Tendon strain was calculated as the change in length related to the initial tendon length ( $\Delta L/L_0$ ). Polynomial functions (2<sup>nd</sup> or 3<sup>rd</sup> order) were fitted to each single force-deformation

curve, which yielded  $r^2 > 0.96$ . Tendon stiffness ( $\Delta \text{force}/\Delta \text{deformation}$ ) and Young's modulus ( $\Delta \text{stress}/\Delta \text{strain}$ ) were calculated for each side within each person in the final 10% (maximum common force of both legs) of the force-deformation and stress-strain curves respectively (43).

#### *Data reduction and analysis*

The two isometric ramp contractions that yielded the greatest maximum force were selected for further analysis. In order to make comparisons before and after immobilization for the same subject and thereby account for differences in magnitude in isometric ramp contraction force, the trials (before and after immobilization) were subsequently analyzed to the maximum common force. Wilcoxon signed rank test were used to examine if there were differences between pre and post immobilization values for the measured variables. An alpha level of  $P < 0.05$  was considered significant. Results are reported as mean  $\pm$  SD.

#### **Results**

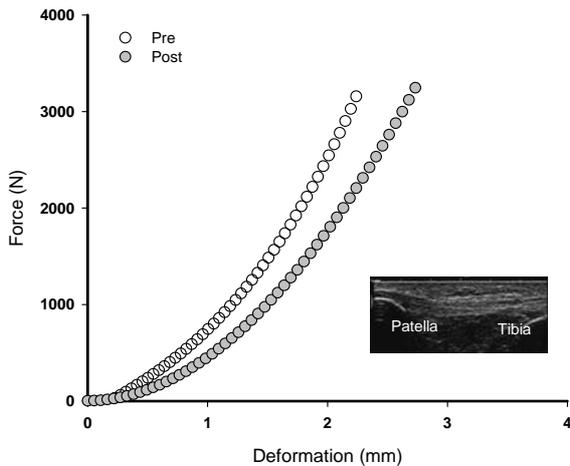
There were no side-to-side differences for any of the measured variables prior to immobilization. Peak knee extensor moment decreased on the immobilized side ( $143 \pm 25$  vs.  $119 \pm 26$  Nm,  $P < 0.05$ ). A statistical trend was observed toward a decrease in peak knee extensor moment ( $152 \pm 32$  vs.  $139 \pm 39$  Nm,  $P = 0.078$ ) on the control side.

Tendon dimensions are shown in Table 1. There were no changes with respect to tendon length or absolute tendon CSA on the immobilized or the control side.

(n = 8)	Immobilized side		Control side	
	Pre	Post	Pre	Post
Tendon length (mm)	43 $\pm$ 3	43 $\pm$ 3	43 $\pm$ 6	43 $\pm$ 3
Tendon CSA (mm <sup>2</sup> )	101 $\pm$ 11	101 $\pm$ 11	99 $\pm$ 15	102 $\pm$ 16

**Table 1.** Patellar tendon dimensions for the immobilized side and control side. Values are mean  $\pm$  SD. There were no differences in patellar tendon dimensions between Pre and Post immobilization.

Mechanical properties assessed at maximum common force are shown in Table 2. Tendon stiffness decreased both on the immobilized side ( $P < 0.01$ ) (Figure 1) and on the control side ( $P < 0.01$ ) (Figure 2). Likewise, Young's Modulus decreased on the immobilized side ( $P < 0.05$ ) (Figure 3) and on the control side ( $P < 0.05$ ) (Figure 4). There were no other differences with respect to tendon deformation, strain or stress on either the immobilized side or the control side.



**Figure 1.** The patellar tendon force-deformation relationship based on common force for the immobilized side. Values are means; SD values are stated in Table 2. Stiffness decreased after 14 days of immobilization (Pre vs. Post,  $P<0.01$ ).

## Discussion

To the best of our knowledge this is the first study that has examined the effects of short-term unilateral immobilization on the structural and mechanical properties of the human patellar tendon, *in vivo*, in old healthy individuals. The main findings were that the mechanical properties of the patellar tendon decreased on the immobilized side. A decline in stiffness and Young's modulus took place without any measurable change in the size of the tendon, suggesting that the material properties of the tendon were primarily affected by the unloading.

It is well known that the immobilization associated drop in muscle strength is greater than that of muscle mass, and that both contribute to a reduced muscle function (44; 52; 62). The larger drop in muscle strength has mainly been attributed to rapid changes in the nervous system that occur with short term unloading (19; 40; 41; 52). These changes seem more accelerated in older than young persons (61). Interestingly, recent studies report that unloading affect tendon properties to a greater extent than muscle loss, which in turn may contribute to a decline in electromechanical delay (the time lag between muscle activation and muscle force production) and reduced rate of force development (7; 17; 33; 34; 52). Nordez et al. (53) reported that the tendon stretching contributed with ~30% of the total electromechanical delay (44; 52). Thus, it appears that tendon properties play a major role in the function of the muscle-tendon complex. The present data corroborate with these earlier findings and further show that the functional effect on the muscle-tendon complex is likely similar in old persons.

Most animal immobilization studies show reduced tendon stiffness without a change in tendon size indicating that the material properties of

the tendon are affected by disuse (22; 39; 47; 48; 56; 67). However, some have shown opposite results (6; 20), which may be related to the use of different models, tendons and durations of immobilization. With a single exception (42), human data in young individuals show that mechanical properties of the tendon decrease with both short-term (17; 34; 35; 54; 59; 60) and long-term (54; 69) unloading in the absence of tendon atrophy, suggesting that the material properties of the tendon were primarily affected. In the present study similar findings were demonstrated for the first time *in vivo* for aging human individuals. Thus, 14 days of immobilization led to decreased tendon stiffness on the immobilized side, which was accompanied by a decrease in Young's Modulus. This decline occurred without changes in tendon dimensions, indicating that the material properties of the patella tendon were altered for our old subjects, which is in line with previous reports in young persons (17; 33; 34; 59; 60).

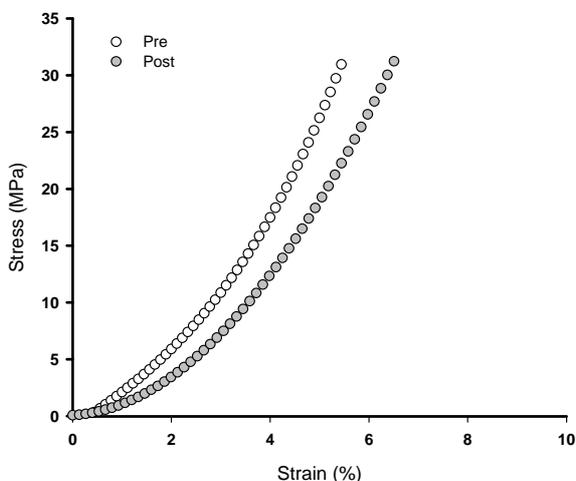
(n = 8)	Immobilized side		Control side	
	Pre	Post	Pre	Post
Deformation (mm)	2.3±0.5	2.7±0.8	2.3±0.4	2.8±0.6
Stiffness (N mm <sup>-1</sup> )	2949±799	2366±774**	3330±839	2365±436**
Stress (MPa)	32±7	33±10	34±8	32±8
Strain (%)	5.5±1.6	6.6±2.2	5.3±0.7	6.5±1.7
Modulus (GPa)	1.2±0.3	1.0±0.3*	1.5±0.4	1.0±0.3*

**Table 2.** Patellar tendon mechanical properties for the immobilized side and control side based on common force. Values are mean ± SD. Significantly different from Pre, \* $P<0.05$ , \*\* $P<0.01$ .

De Boer et al. (17) found that quadriceps strength dropped ~15% and that the mechanical properties of the patella tendon declined ~10% after 2 weeks of immobilization in young men. After an additional 9 days of unloading quadriceps strength declined with a total ~20% and the mechanical properties of the patellar tendon with a total ~30%. There was an associated ~40% reduction in the rate of force development after the 14 days of unloading, which remained unchanged after 23 days. Furthermore, it has been shown in young and middle-aged men that the mechanical properties of the achilles tendon decreased up to ~58% with 90 days of immobilization (54). Collectively these data demonstrate that short-term limb unloading has considerable effects on the mechanical properties of the tendon that to some extent mirrors that of the muscle, and that these changes can be expected to occur in the early phase of unloading. In the present study we extended these findings by showing that muscle tendon mechanics are also affected by acute disuse in the elderly, and that the magnitude of change is perhaps even greater.

The biochemical changes within the tendon due to immobilization are still largely unknown (31; 55). In animal studies unloading causes a decrease

in collagen synthesis (29; 57), collagen concentration (24; 65) and the ultrastructure (fibril size/ density (50; 51). However, others have found that collagen concentration (1; 3; 57), ultrastructure (70) and tendon tissue mass (25) remain unchanged by unloading. In humans, ~2 weeks (14) and ~7 weeks of unloading (13) did not seem to affect collagen synthesis in the achilles tendon, whereas collagen synthesis rate decreased in the patellar tendon after 10-23 days of immobilization (18), suggesting that differentiated responses may be observed between anatomically distinct tendons. Interestingly, in the latter case the decreased synthesis rate was accompanied by an almost similar reduction in tendon stiffness, Young's Modulus while tendon CSA remained unchanged, indicating that the material properties of the tendon was altered, which is line with the present findings. It should be noted that degradation and catabolic enzymes was not evaluated in the study of De Boer et al(18), and these parameters have been shown to increase with loss of tendon tension (5; 38). The observed decrease in tendon stiffness could well be associated with increased collagen degradation. The results obtained in our study, together with those of De Boer et al. (18), suggests, that the deteriorating effects of unloading are rapidly evoked both in old and young individuals.



**Figure 3.** The patellar tendon stress-strain relationship based on common force for the immobilized side. Values are means; SD values are stated in Table 2. Young's Modulus decreased after 14 days of immobilization (Pre vs. Post,  $P < 0.05$ ).

It has been proposed that the degree of tension on the patellar tendon during immobilization could influence the tendon biomechanical properties quite dramatically within one week (45). Animal studies show that if the tendon is left in a unloaded (shortened) position during 7 days of immobilization, enzyme activity of the collagen synthesis is decreased (29; 57), potentially leading to altered tendon composition and weakened mechanical properties (3). In contrast, if the tendon was kept in a neutral or chronically loaded (elongated) position while

immobilized, the decrease in collagen synthesis enzyme activity was only minor or even fully abolished (29; 57). The present study did not investigate possible biochemical changes, and it should be kept in mind that the immobilized knee was positioned in a relatively shorten position (~30° knee flexion), which likely is associated with no or only very minor passive tension in the patella tendon (43). The acute lack of tensile tendon input during the immobilization period might have influenced the material properties of the tendon, since tendon stiffness and modulus decreased without tendon atrophy. Unloading tenocytes have been shown to have severe effects on collagen expression, proteoglycan expression, growth factor secretion and metalloproteinases (5; 38). Furthermore, it has recently been demonstrated that complete tension deprivation in the rabbit patellar tendon induced fibroblast apoptosis within 24 hrs (30). Thus, it is possible that the removed tension development in the patella tendon was the initiating event for the present findings of reduced tendon stiffness and Young's Modulus.

The unchanged tendon size due to immobilization is in agreement with numerous other studies (14; 17; 33; 34; 54; 60; 69). It has been reported that there is a decrease in the number and in average diameter of collagen fibrils as result of immobilization (46; 50; 51), but it has also been shown that the fibril to fibril distance increases with immobilization compared to control tendons, suggesting a less fibril dense tendon (51). It is possible that the increased fibril to fibril distance represents greater relative content of interstitial water and/or other extracellular matrix components while whole tendon CSA remains unchanged (64; 68) as proposed by others (18). Such a mechanism would explain the altered material properties but unchanged CSA in the present study.

A number of limitations may be observed with the present study. Unfortunately, we were unable to obtain tendon biopsies in the present study, which would have allowed us to address possible changes in tendon composition. Furthermore, to our surprise changes occurred in the non-immobilized limb, which rendered it inadequate as a control. Albeit non-significant, muscle strength tended ( $P=0.078$ ) to decrease on the non-immobilized side. Furthermore, both stiffness and modulus also decreased on the non-immobilized side. Thus, although the data show that the non-immobilized side was not an ideal control in this sample, the data suggests that muscle strength changes are accompanied by changes in tendon properties. A possible explanation for the findings in the control limb is that unilateral casting and crutch walking in old persons may influence the overall activity level.

In conclusion, a decline in stiffness and Young's modulus was observed following short-term (14 days) unilateral limb immobilization in old men without any changes in tendon size, suggesting that the material properties of the tendon were affected

by unloading. These findings indicate that short-term inactivity has a major negative influence on the mechanical properties of the human patellar tendon in the elderly.

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#### Reference List

1. **Akeson WH, Woo SL, Amiel D, Coutts RD and Daniel D.** The connective tissue response to immobility: biochemical changes in periarticular connective tissue of the immobilized rabbit knee. *Clin Orthop Relat Res* 356-362, 1973.
2. **Alexander RM and net-Clark HC.** Storage of elastic strain energy in muscle and other tissues. *Nature* 265: 114-117, 1977.
3. **Amiel D, Woo SL, Harwood FL and Akeson WH.** The effect of immobilization on collagen turnover in connective tissue: a biochemical-biomechanical correlation. *Acta Orthop Scand* 53: 325-332, 1982.
4. **Arampatzis A, Karamanidis K and Albracht K.** Adaptational responses of the human Achilles tendon by modulation of the applied cyclic strain magnitude. *J Exp Biol* 210: 2743-2753, 2007.
5. **Arnoczky SP, Lavagnino M and Egerbacher M.** The mechanobiological aetiopathogenesis of tendinopathy: is it the over-stimulation or the under-stimulation of tendon cells? *Int J Exp Pathol* 88: 217-226, 2007.
6. **Arruda EM, Calve S, Dennis RG, Mundy K and Baar K.** Regional variation of tibialis anterior tendon mechanics is lost following denervation. *J Appl Physiol* 101: 1113-1117, 2006.
7. **Bamman MM, Clarke MS, Feeback DL, Talmadge RJ, Stevens BR, Lieberman SA and Greenisen MC.** Impact of resistance exercise during bed rest on skeletal muscle sarcopenia and myosin isoform distribution. *J Appl Physiol* 84: 157-163, 1998.
8. **Biewener A and Baudinette R.** In vivo muscle force and elastic energy storage during steady-speed hopping of tammar wallabies (*Macropus eugenii*). *J Exp Biol* 198: 1829-1841, 1995.
9. **Bojsen-Moller J, Hansen P, Aagaard P, Kjaer M and Magnusson SP.** Measuring mechanical properties of the vastus lateralis tendon-aponeurosis complex in vivo by ultrasound imaging. *Scand J Med Sci Sports* 13: 259-265, 2003.
10. **Bojsen-Moller J, Kalliokoski KK, Seppanen M, Kjaer M and Magnusson SP.** Low-intensity tensile loading increases intratendinous glucose uptake in the Achilles tendon. *J Appl Physiol* 101: 196-201, 2006.
11. **Booth FW and Gould EW.** Effects of training and disuse on connective tissue. *Exerc Sport Sci Rev* 3: 83-112, 1975.
12. **Butler DL, Grood ES, Noyes FR and Zernicke RF.** Biomechanics of ligaments and tendons. *Exerc Sport Sci Rev* 6: 125-181, 1978.
13. **Christensen B, Dyrberg E, Aagaard P, Enehjelm S, Krogsgaard M, Kjaer M and Langberg H.** Effects of long-term immobilization and recovery on human triceps surae and collagen turnover in the Achilles tendon in patients with healing ankle fracture. *J Appl Physiol* 105: 420-426, 2008.
14. **Christensen B, Dyrberg E, Aagaard P, Kjaer M and Langberg H.** Short-term immobilization and recovery affect skeletal muscle but not collagen tissue turnover in humans. *J Appl Physiol* 105: 1845-1851, 2008.
15. **Coupe C, Hansen P, Kongsgaard M, Kovanen V, Suetta C, Aagaard P, Kjaer M and Magnusson SP.** Mechanical properties and collagen cross-linking of the patellar tendon in old and young men. *J Appl Physiol* 107: 880-886, 2009.
16. **Coupe C, Kongsgaard M, Aagaard P, Hansen P, Bojsen-Moller J, Kjaer M and Magnusson SP.** Habitual loading results in tendon hypertrophy and increased stiffness of the human patellar tendon. *J Appl Physiol* 105: 805-810, 2008.
17. **de B, Maganaris CN, Seynnes OR, Rennie MJ and Narici MV.** Time course of muscular, neural and tendinous adaptations to 23 day unilateral lower-limb suspension in young men. *J Physiol* 583: 1079-1091, 2007.
18. **de B, Selby A, Atherton P, Smith K, Seynnes OR, Maganaris CN, Maffulli N, Movin T, Narici MV and Rennie MJ.** The temporal responses of protein synthesis, gene expression and cell signalling in human quadriceps muscle and patellar tendon to disuse. *J Physiol* 585: 241-251, 2007.
19. **Deschenes MR, Giles JA, McCoy RW, Volek JS, Gomez AL and Kraemer WJ.** Neural factors account for strength decrements observed after short-term muscle unloading. *Am J Physiol Regul Integr Comp Physiol* 282: R578-R583, 2002.

20. **Eliasson P, Fahlgren A, Pasternak B and Aspenberg P.** Unloaded rat Achilles tendons continue to grow, but lose viscoelasticity. *J Appl Physiol* 103: 459-463, 2007.
21. **ELLIOTT DH.** STRUCTURE AND FUNCTION OF MAMMALIAN TENDON. *Biol Rev Camb Philos Soc* 40: 392-421, 1965.
22. **Hannafin JA, Arnoczky SP, Hoonjan A and Torzilli PA.** Effect of stress deprivation and cyclic tensile loading on the material and morphologic properties of canine flexor digitorum profundus tendon: an in vitro study. *J Orthop Res* 13: 907-914, 1995.
23. **Hansen P, Bojsen-Moller J, Aagaard P, Kjaer M and Magnusson SP.** Mechanical properties of the human patellar tendon, in vivo. *Clin Biomech (Bristol, Avon)* 2005.
24. **Harwood FL and Amiel D.** Differential metabolic responses of periarticular ligaments and tendon to joint immobilization. *J Appl Physiol* 72: 1687-1691, 1992.
25. **Heinemeier KM, Olesen JL, Haddad F, Schjerling P, Baldwin KM and Kjaer M.** Effect of unloading followed by reloading on expression of collagen and related growth factors in rat tendon and muscle. *J Appl Physiol* 106: 178-186, 2009.
26. **Hespeel P, Op't EB, Van LM, Urso B, Greenhaff PL, Labarque V, Dymarkowski S, Van HP and Richter EA.** Oral creatine supplementation facilitates the rehabilitation of disuse atrophy and alters the expression of muscle myogenic factors in humans. *J Physiol* 536: 625-633, 2001.
27. **Hortobagyi T, Dempsey L, Fraser D, Zheng D, Hamilton G, Lambert J and Dohm L.** Changes in muscle strength, muscle fibre size and myofibrillar gene expression after immobilization and retraining in humans. *J Physiol* 524 Pt 1: 293-304, 2000.
28. **Jones SW, Hill RJ, Krasney PA, O'Conner B, Peirce N and Greenhaff PL.** Disuse atrophy and exercise rehabilitation in humans profoundly affects the expression of genes associated with the regulation of skeletal muscle mass. *FASEB J* 18: 1025-1027, 2004.
29. **Karpakka J, Vaananen K, Virtanen P, Savolainen J, Orava S and Takala TE.** The effects of remobilization and exercise on collagen biosynthesis in rat tendon. *Acta Physiol Scand* 139: 139-145, 1990.
30. **Kawabata H, Katsura T, Kondo E, Kitamura N, Miyatake S, Tanabe Y, Setoguchi T, Komiya S and Yasuda K.** Stress deprivation from the patellar tendon induces apoptosis of fibroblasts in vivo with activation of mitogen-activated protein kinases. *J Biomech* 42: 2611-2615, 2009.
31. **Kjaer M, Langberg H, Heinemeier K, Bayer ML, Hansen M, Holm L, Doessing S, Kongsgaard M, Krogsgaard MR and Magnusson SP.** From mechanical loading to collagen synthesis, structural changes and function in human tendon. *Scand J Med Sci Sports* 19: 500-510, 2009.
32. **Kongsgaard M, Reitelsheder S, Pedersen TG, Holm L, Aagaard P, Kjaer M and Magnusson SP.** Region specific patellar tendon hypertrophy in humans following resistance training. *Acta Physiol (Oxf)* 191: 111-121, 2007.
33. **Kubo K, Akima H, Kouzaki M, Ito M, Kawakami Y, Kanehisa H and Fukunaga T.** Changes in the elastic properties of tendon structures following 20 days bed-rest in humans. *Eur J Appl Physiol* 83: 463-468, 2000.
34. **Kubo K, Akima H, Ushiyama J, Tabata I, Fukuoka H, Kanehisa H and Fukunaga T.** Effects of 20 days of bed rest on the viscoelastic properties of tendon structures in lower limb muscles. *Br J Sports Med* 38: 324-330, 2004.
35. **Kubo K, Akima H, Ushiyama J, Tabata I, Fukuoka H, Kanehisa H and Fukunaga T.** Effects of resistance training during bed rest on the viscoelastic properties of tendon structures in the lower limb. *Scand J Med Sci Sports* 14: 296-302, 2004.
36. **Langberg H, Rosendal L and Kjaer M.** Training-induced changes in peritendinous type I collagen turnover determined by microdialysis in humans. *J Physiol* 534: 297-302, 2001.
37. **Langberg H, Skovgaard D, Petersen LJ, Bulow J and Kjaer M.** Type I collagen synthesis and degradation in peritendinous tissue after exercise determined by microdialysis in humans. *J Physiol* 521 Pt 1: 299-306, 1999.
38. **Lavagnino M, Arnoczky SP, Tian T and Vaupel Z.** Effect of amplitude and frequency of cyclic tensile strain on the inhibition of MMP-1 mRNA expression in tendon cells: an in vitro study. *Connect Tissue Res* 44: 181-187, 2003.
39. **Loitz BJ, Zernicke RF, Vailas AC, Kody MH and Meals RA.** Effects of short-term immobilization versus continuous passive motion on the biomechanical and biochemical properties of the rabbit tendon. *Clin Orthop Relat Res* 265-271, 1989.
40. **Lundbye-Jensen J and Nielsen JB.** Central nervous adaptations following 1 wk of wrist and hand immobilization. *J Appl Physiol* 105: 139-151, 2008.

41. **Lundbye-Jensen J and Nielsen JB.** Immobilization induces changes in presynaptic control of group Ia afferents in healthy humans. *J Physiol* 586: 4121-4135, 2008.
42. **Maganaris CN, Reeves ND, Rittweger J, Sargeant AJ, Jones DA, Gerrits K and De Haan A.** Adaptive response of human tendon to paralysis. *Muscle Nerve* 33: 85-92, 2006.
43. **Magnusson SP, Hansen P, Aagaard P, Brond J, Dyhre-Poulsen P, Bojsen-Moller J and Kjaer M.** Differential strain patterns of the human gastrocnemius aponeurosis and free tendon, in vivo. *Acta Physiol Scand* 177: 185-195, 2003.
44. **Magnusson SP, Narici MV, Maganaris CN and Kjaer M.** Human tendon behaviour and adaptation, in vivo. *J Physiol* 586: 71-81, 2008.
45. **Majima T, Yasuda K, Fujii T, Yamamoto N, Hayashi K and Kaneda K.** Biomechanical effects of stress shielding of the rabbit patellar tendon depend on the degree of stress reduction. *J Orthop Res* 14: 377-383, 1996.
46. **Majima T, Yasuda K, Tsuchida T, Tanaka K, Miyakawa K, Minami A and Hayashi K.** Stress shielding of patellar tendon: effect on small-diameter collagen fibrils in a rabbit model. *J Orthop Sci* 8: 836-841, 2003.
47. **Matsumoto F, Trudel G, Uthoff HK and Backman DS.** Mechanical effects of immobilization on the Achilles' tendon. *Arch Phys Med Rehabil* 84: 662-667, 2003.
48. **meida-Silveira MI, Lambertz D, Perot C and Goubel F.** Changes in stiffness induced by hindlimb suspension in rat Achilles tendon. *Eur J Appl Physiol* 81: 252-257, 2000.
49. **Miller BF, Olesen JL, Hansen M, Dossing S, Cramer RM, Welling RJ, Langberg H, Flyvbjerg A, Kjaer M, Babraj JA, Smith K and Rennie MJ.** Coordinated collagen and muscle protein synthesis in human patella tendon and quadriceps muscle after exercise. *J Physiol* 567: 1021-1033, 2005.
50. **Muellner T, Kwasny O, Loehnert V, Mallinger R, Unfried G, Schabus R and Plenk H, Jr.** Light and electron microscopic study of stress-shielding effects on rat patellar tendon. *Arch Orthop Trauma Surg* 121: 561-565, 2001.
51. **Nakagawa Y, Totsuka M, Sato T, Fukuda Y and Hirota K.** Effect of disuse on the ultrastructure of the achilles tendon in rats. *Eur J Appl Physiol Occup Physiol* 59: 239-242, 1989.
52. **Narici MV and Maganaris CN.** Plasticity of the muscle-tendon complex with disuse and aging. *Exerc Sport Sci Rev* 35: 126-134, 2007.
53. **Nordez A, Gallot T, Catheline S, Guevel A, Cornu C and Hug F.** Electromechanical delay revisited using very high frame rate ultrasound. *J Appl Physiol* 106: 1970-1975, 2009.
54. **Reeves ND, Maganaris CN, Ferretti G and Narici MV.** Influence of 90-day simulated microgravity on human tendon mechanical properties and the effect of resistive countermeasures. *J Appl Physiol* 98: 2278-2286, 2005.
55. **Rennie M.** Mysterious properties of tendon metabolism. *J Appl Physiol* 105: 397, 2008.
56. **Rumian AP, Draper ER, Wallace AL and Goodship AE.** The influence of the mechanical environment on remodelling of the patellar tendon. *J Bone Joint Surg Br* 91: 557-564, 2009.
57. **Savolainen J, Myllyla V, Myllyla R, Vihko V, Vaananen K and Takala TE.** Effects of denervation and immobilization on collagen synthesis in rat skeletal muscle and tendon. *Am J Physiol* 254: R897-R902, 1988.
58. **Seynnes OR, Erskine RM, Maganaris CN, Longo S, Simoneau EM, Grosset JF and Narici MV.** Training-induced changes in structural and mechanical properties of the patellar tendon are related to muscle hypertrophy, but not to strength gains. *J Appl Physiol* 2009.
59. **Seynnes OR, Maffiuletti NA, Maganaris CN, de B, Pensini M, di Prampero PE and Narici MV.** Soleus T reflex modulation in response to spinal and tendinous adaptations to unilateral lower limb suspension in humans. *Acta Physiol (Oxf)* 194: 239-251, 2008.
60. **Shin D, Finni T, Ahn S, Hodgson JA, Lee HD, Edgerton VR and Sinha S.** Effect of chronic unloading and rehabilitation on human Achilles tendon properties: a velocity-encoded phase-contrast MRI study. *J Appl Physiol* 105: 1179-1186, 2008.
61. **Suetta C, Hvid LG, Justesen L, Christensen U, Neergaard K, Simonsen L, Ortenblad N, Magnusson SP, Kjaer M and Aagaard P.** Effects of ageing on human skeletal muscle after immobilisation and re-training. *J Appl Physiol* 2009.
62. **Suetta C, Hvid LG, Justesen L, Christensen U, Neergaard K, Simonsen L, Ortenblad N, Magnusson SP, Kjaer M and Aagaard P.** Effects of ageing on human skeletal muscle after immobilisation and re-training. *J Appl Physiol* 2009.
63. **Tipton CM, Vailas AC and Matthes RD.** Experimental studies on the influences of physical activity on ligaments, tendons and joints: a brief review. *Acta Med Scand Suppl* 711:157-68.: 157-168, 1986.

64. **Tsuchida T, Yasuda K, Kaneda K, Hayashi K, Yamamoto N, Miyakawa K and Tanaka K.** Effects of in situ freezing and stress-shielding on the ultrastructure of rabbit patellar tendons. *J Orthop Res* 15: 904-910, 1997.
65. **Vailas AC, Deluna DM, Lewis LL, Curwin SL, Roy RR and Alford EK.** Adaptation of bone and tendon to prolonged hindlimb suspension in rats. *J Appl Physiol* 65: 373-376, 1988.
66. **Visser JJ, Hoogkamer JE, Bobbert MF and Huijing PA.** Length and moment arm of human leg muscles as a function of knee and hip-joint angles. *Eur J Appl Physiol Occup Physiol* 61: 453-460, 1990.
67. **Woo SL, Gomez MA, Woo YK and Akeson WH.** Mechanical properties of tendons and ligaments. II. The relationships of immobilization and exercise on tissue remodeling. *Biorheology* 19: 397-408, 1982.
68. **Yamamoto N, Ohno K, Hayashi K, Kuriyama H, Yasuda K and Kaneda K.** Effects of stress shielding on the mechanical properties of rabbit patellar tendon. *J Biomech Eng* 115: 23-28, 1993.
69. **Zhao H, Ren Y, Wu YN, Liu SQ and Zhang LQ.** Ultrasonic evaluations of Achilles tendon mechanical properties poststroke. *J Appl Physiol* 106: 843-849, 2009.
70. **Zhou J, Koike Y, Uhthoff HK and Trudel G.** Quantitative histology and ultrastructure fail to explain weakness of immobilized rabbit Achilles' tendons. *Arch Phys Med Rehabil* 88: 1177-1184, 2007.